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Research article

urn:lsid:zoobank.org:pub:CF109FEA-7812-4787-A164-4F5FF0C46E21

A new genus of Achrysonini and new records in Cerambycidae (Coleoptera, Chrysomeloidea) from Colombia

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Abstract. *Ysachron pilosus* gen. et sp. nov. is described from the Caribbean region of Colombia, and an updated key to genera of South American Achrysonini is provided. Moreover, the geographical distribution of 45 species of Cerambycidae, belonging to the subfamilies Cerambycinae, Lamiinae and Prioninae, is expanded, including nine new country records for Colombia and a further locality in Colombia for nine species.

Keywords. Cerambycinae, Lamiinae, new species, Prioninae, taxonomy.

García K., Botero J.P. & Santos-Silva A. 2021. A new genus of Achrysonini and new records in Cerambycidae (Coleoptera, Chrysomeloidea) from Colombia. *European Journal of Taxonomy* 735: 89–109.
<https://doi.org/10.5852/ejt.2021.735.1243>

Introduction

Cerambycinae Latreille, 1802 is the second largest subfamily of Cerambycidae Latreille, 1802, with worldwide distribution, more than 110 tribes, approximately 1800 genera, and 12 000 species (Tavakilian & Chevillotte 2020). This subfamily is the most speciose in South America (Švácha & Lawrence 2014; Tavakilian & Chevillotte 2020).

The tribe Achrysonini was proposed by Lacordaire (1868) to include five genera: *Achryson* Audinet-Serville, 1833; *Eurymerus* Audinet-Serville, 1833; *Allogaster* Thomson, 1864; *Icosium* Lucas, 1854; and *Nortia* Thomson, 1864. Of these, just the first two are South American and *Eurymerus* is currently allocated in Ectenessini Martins & Galileo, 1998. Lacordaire (1868) mentioned the similarity of Achrysonini with Oemini Lacordaire, 1868 but differentiated them by the procoxal cavities not strongly

angulated laterally in Achrysonini (strongly angulated in Oemini). Bates (1870) also compared both tribes and differentiated them by the shape of the anterior procoxal cavities.

Martins (1998) and Martins & Galileo (1999) mentioned that Achrysonini is characterized by the presence of setae between the ommatidia and used this character to justify the transfer of several genera among tribes of Cerambycinae.

Martins (2002) revised the South American species of Achrysonini and characterized the tribe by the presence of eyes coarsely faceted and with setae between the ommatidia. In that work, Martins provided keys for the South American genera and species. Monné & Monné (2004) described *Neoachryson castaneum*, a new genus and species from Argentina.

Currently, the tribe is composed of 22 genera (Tavakilian & Chevillotte 2020), of which 12 are distributed in the new world, and nine genera and 31 species occur in South America (Monné 2020).

Herein, we describe a new genus and a new species of Achrysonini and provide an updated key to South America genera of the tribe. Additionally, we expand the known geographical distribution for 45 species, nine of them registered for the first time for Colombia.

Material and methods

The material examined was collected in fragments of tropical dry forest from the Caribbean region of Colombia, in the departments of Atlántico (Reserva Campesina la Montaña) and Bolívar (Reserva La Flecha). These locations were sampled from February to June 2018 using a UV light trap, white light trap, manual capture and beating sheet.

The specimens are currently deposited in the following institutions, which are subsequently referred to by their acronyms:

MPUJ = Pontificia Universidad Javeriana, Bogotá, Colombia

MZSP = Museu de Zoologia, Universidade de São Paulo, São Paulo, São Paulo, Brazil

UARC = Universidad del Atlántico, Puerto Colombia, Atlántico, Colombia

Photographs were taken with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65 mm f/2.8 1–5X macro lens, controlled by Zerene Stacker focus stacking software. Measurements were taken in ‘mm’ using the software Leica Application Suite (LAS 4.0) and Leica M125 stereo microscope, also used in the study of the specimens. References and geographical distributions were verified in Martínez (2000), Monné (2020), and Tavakilian & Chevillotte (2020) catalogs. The terminology used herein for external morphological structures follows Lawrence *et al.* (2010) and for male terminalia Ehara (1954) and Švácha & Lawrence (2014).

Results

Taxonomy

Class Insecta Linnaeus, 1758
Order Coleoptera Linnaeus, 1758
Suborder Polyphaga Emery, 1886
Superfamily Chrysomeloidea Latreille, 1802
Family Cerambycidae Latreille, 1802
Subfamily Cerambycinae Latreille, 1802
Tribe Achrysonini Lacordaire, 1868

Ysachron gen. nov.

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Figs 1–12

Type species

Ysachron pilosus gen. et sp. nov., here designated.

Differential diagnosis

Ysachron gen. nov. is similar to some genera of Hesperophanini Mulsant, 1839, Oemini and Achrysonini. Despite not having the setae between ommatidia, we allocate it in Achrysonini due to the presence of the following characteristics: eyes coarsely faceted; antennomeres unarmed; antennae in males longer than body; scape shorter than antennomere III; procoxal cavities open and not strongly angulate laterally (Fig. 13, *Ysachron pilosus* gen. et sp. nov.; Fig. 14, *Achryson surinamum* (Linnaeus, 1767)); prosternal process without lateral projections apically; and metatibiae not carinate.

The presence or absence of setae between ommatidia is a characteristic that also varies among subtribes of Oemini (Martins, 1997): present in Oemina Lacordaire, 1868, absent in Methioidina Martins, 1997 (except in *Proeme* Martins, 1978).

Although we are aware of the great similarity between the tribes Achrysonini, Hesperophanini and Oemini, *Ysachron* gen. nov. differs from Hesperophanini and Oemini by having procoxal cavities not strongly angulate laterally (Figs 13–14), while they are strongly angulate in the latter two (Figs 15–16). The new genus also differs from genera of Hesperophanini by the median lobe with the apical region shorter than basal apophysis (Figs 27–29).

Among the genera of Achrysonini, *Ysachron* gen. nov. is similar to *Achryson* by the rounded prothorax sides without projections, basal antennomeres linear, scape as long as half of the length of antennomere III, and fuscous aspect. It differs as follows: eyes lacking setae between the ommatidia; surface of the pronotum alveolate; antennae 12-segmented; elytra with long setae; and meso- and metafemora subclavate. Species of *Achryson* have setae between ommatidia, surface of the pronotum microsculptured, antennae 11-segmented, elytra with short setae, and femora fusiform. Also, unlike some other genera in Achrysonini, *Ysachron* gen. nov. does not show sexual punctuation differences.

Etymology

The name *Ysachron* is an anagram of *Achryson*, a similar genus of Achrysonini. The gender is masculine.

Description

Head narrowed behind eyes, densely alveolate between upper eye lobes. Median groove well-marked between antennal tubercles. Antennal tubercles weakly elevated, with obtuse apex. Eyes coarsely

faceted, not coplanar with surface around them; without setae between ommatidia; upper eye lobes distant from each other; lower eye lobes large, about three times the length of genae. Genae small, with rounded apex. Labrum small, when together with clypeus, longer than frons. Mandibles short, with acute apex. Maxillary and labial palpi of similar length. Antennae 12-segmented, longer than body in males, as long as body in females; scape short, somewhat cylindrical, as long as about half length of antennomere III; antennomere III longer than IV. Prothorax subcylindrical; lateral margin rounded and unarmed, constricted in anterior and posterior margins. Prosternal process narrow, parallel sided, with apex curved inwards. Mesoventrite tumid. Mesoventral process emarginated at apex. Metaventricle rectangular, slightly wider than long, parallel-sided. Scutellum small. Elytra not exposing abdomen, parallel-sided; humeri rounded; base wider than posterior margin of prothorax; surface not carinate. Femora from subfusiform to slightly clavate. Tibiae slightly widened toward apex. Abdominal ventrite I the largest (including abdominal process), ventrites II–V subequal in length.

Key to South American genera of Achrysonini (modified and translated from Martins 2002 and Monné & Monné 2004)

1. Setae between ommatidia absent; antennae 12-segmented *Ysachron* gen. nov.
– Setae between ommatidia present; antennae 11-segmented 2
2. Sides of prothorax with long spine (longer than pedicel), with curved apex backward
..... *Drascalía* Fairmaire & Germain, 1864
– Sides of prothorax unarmed (without spines or with small gibbosity) or at most with short triangular tubercle 3
3. Pronotum with two tubercles on anterior third; elytra with longitudinal elevation, more evident near base, reaching apical fourth, delimiting a flattened area near suture .. *Abyarachryson* Martins, 2002
– Pronotum without tubercles (except for small gibbosities in some species of *Achryson*); elytra uniformly convex or flattened on apical $\frac{2}{3}$ 4
4. Elytra flattened on apical $\frac{2}{3}$; metafemora linear *Neoachryson* Monné & Monné, 2004
– Elytra uniformly convex, metafemora not linear 5
5. Metafemora strongly clavate 6
– Metafemora subfusiform, with elongate club 7
6. Antennomere III longer than scape and other antennomeres *Huequenía* Cerda, 1890
– Antennomere III shorter than scape and other antennomeres *Esseiachryson* Martins, 2002
7. Elytra with contrasting punctures and glabrous perimeter *Cotyachryson* Martins, 2002
– Elytra without contrasting punctures 8
8. Basal antennomeres thickened; sides of prothorax with small central gibbosity
..... *Cerdaia* Monné, 2006
– Basal antennomeres linear; sides of prothorax rounded 9
9. Scape about as long or slightly shorter than antennomere III; elytra with long setae and shining aspect *Xenocompsa* Martins, 1965
– Scape about as long as half the length of antennomere III; elytra often with short setae and general aspect not shining *Achryson* Audinet-Serville, 1833

Ysachron pilosus gen. et sp. nov.

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Figs 1–13, 17–29

Differential diagnosis

As for genus.

Etymology

The species epithet comes from the Latin ‘*pilosus*’, referring to the general appearance of the species, covered by long and erect setae.

Material examined

Holotype

COLOMBIA • 1 ♂; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 19 Feb. 2018; K. García leg; tropical dry forest, beating sheet; MPUJ_ENT 0071150.

Paratypes

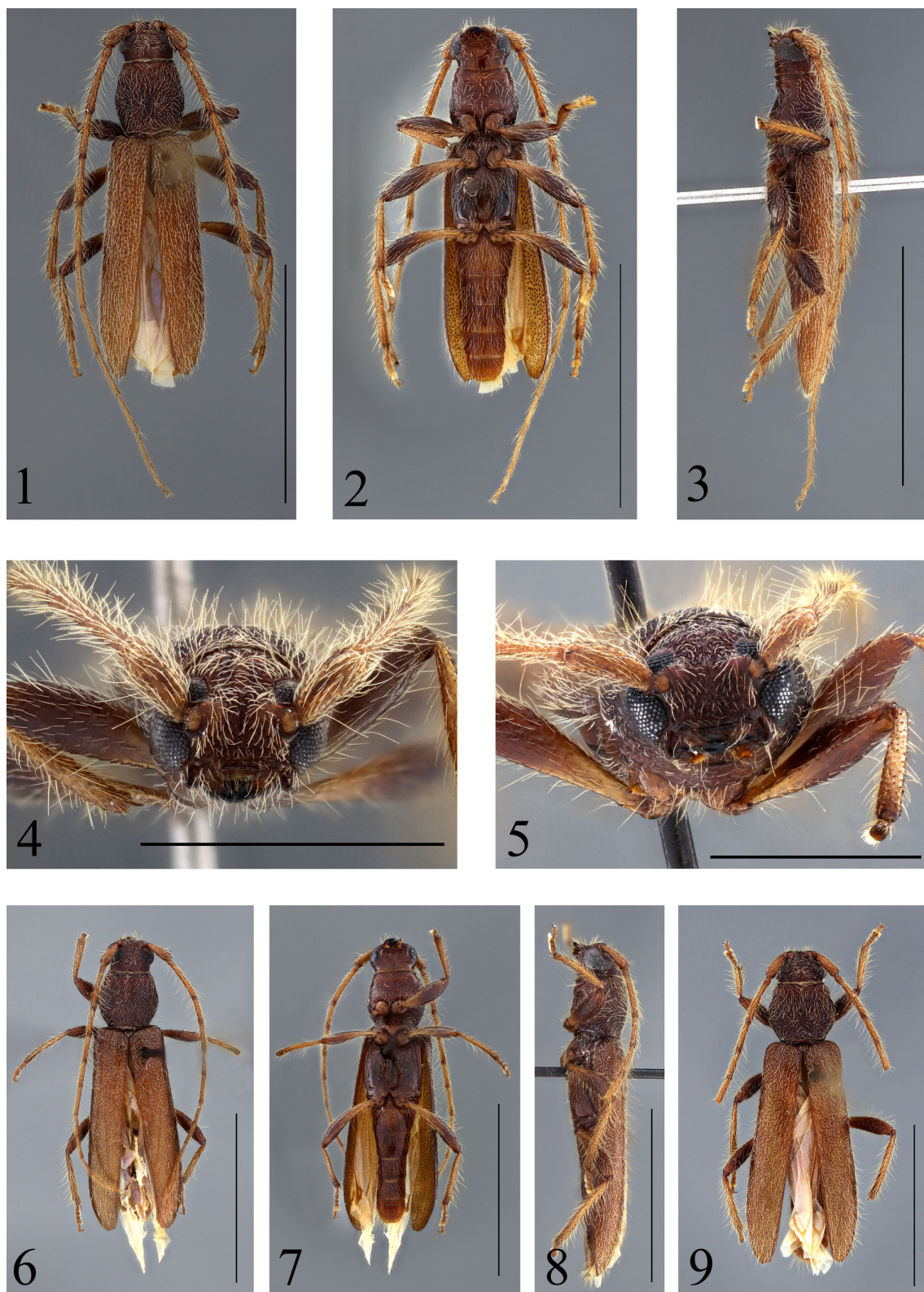
COLOMBIA • 1 ♂; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N 75°0.2'34" W; 260 m a.s.l.; 15–16 Feb. 2018; K. García leg; tropical dry forest, white light trap; MZSP • 1 ♂; same collection data as for preceding; 17–18 Mar. 2018; MPUJ_ENT 0071151 • 1 ♀; same collection data as for holotype; MZSP • 1 ♀; same collection data as for preceding; 16–17 Mar. 2018; UV light trap; MPUJ_ENT 0071152.

Description of holotype male (Figs 1–4, 10, 13, 17–29)

COLORATION. Integument brown with yellowish-white setae, whiter depending on light intensity. Head and prothorax reddish brown. Antennae light brown; scape and pedicel slightly darker; antennomeres III–VI with apical margin dark brown. Ventral surface reddish brown, metaventricle darker on some areas. Base of femora light brown, remaining surface dark brown. Apical margin of tibiae dark. Body covered by long and erect, dense yellowish-white setae.

HEAD. Frons alveolate. Area between antennal tubercles densely, coarsely alveolate. Antennal tubercles elevated, smooth. Median groove distinct from frons to area between anterior margin of upper eye lobes. Area between upper eye lobes alveolate, alveoli coarser toward antennal tubercles, smooth on central area; with abundant, long, erect setae, denser centrally. Area behind upper eye lobes slightly alveolate, with central area sparsely punctate; with a few erect setae. Genae smooth; with a few long, erect setae. Distance between upper eye lobes about three times width of an upper eye lobe. Antennae 2.1 times elytral length, reaching elytral apex at antennomere IX. Scape with sparse piliferous punctures. Antennomeres with abundant, distinctly long, erect setae, shorter toward distal segments. Antennal formula (ratio) based on length of antennomere III: scape = 0.59; pedicel = 0.14; IV = 0.76; V = 0.78; VI = 0.81; VII = 0.89; VIII = 0.81; IX = 0.76; X = 0.70; XI = 0.62; XII = 0.57.

THORAX. Prothorax as long as wide. Surface of pronotum coarsely alveolate, except for small longitudinal area behind middle. Lateral sides of prothorax sparsely punctate. Prosternum slightly and transversely striated, smooth on anterior area. Prosternal process narrow, distinctly narrowed toward apex; width at narrowest point about 0.12 times width of procoxal cavity. Mesanepisternum and mesepimeron densely setose. Mesoventrite sparsely punctate. Mesoventral process microsculptured, with a few long, erect setae; emarginated at apex; width at narrowest point about 0.4 times width of mesocoxal cavity. Metanepisternum abundantly setose. Metaventricle smooth centrally, rugose on anterior sides; with



Figs 1–9. *Ysachron pilosus* gen. et sp. nov. 1–4. Holotype, ♂ (MPUJ_ENT 0071150). 1. Dorsal view. 2. Ventral view. 3. Lateral view. 4. Details of head, frontal view. 5–8. Paratype, ♀ (MPUJ_ENT 0071152). 5. Details of head, frontal view. 6. Dorsal view. 7. Ventral view. 8. Lateral view. 9. Paratype, ♀ (MZSP). Dorsal view with sides of prothorax strongly rounded. Scale bars: 1–4, 6–9 = 0.5 cm; 5 = 0.25 cm.

sparse long, erect setae, gradually sparser toward glabrous central area. Scutellum smooth, with short, decumbent setae; posterior margin rounded. Elytra with distinct piliferous punctures, with erect setae irregularly organized. Apex of elytra obliquely truncate.

LEGS. Profemora fusiform, meso- and metafemora subclavate; lateral sides of femora rugose, remaining surface smooth; setae denser toward apex. Metatarsomere I about as long as II–III together.

ABDOMEN. Ventrites sparsely, finely punctate, punctures denser centrally; with long decumbent setae, erect laterally. Apex of ventrite V truncate.

TERMINALIA. Tergite VIII (Fig. 17) with distal margin distinctly emarginated, with long and short yellow setae, abundant and longer laterally. Sternite VIII (Fig. 18) transverse, distal margin slightly emarginated, projected forward laterally, with long yellow setae laterally; apophysis short, rounded at apex, about as long as central area of sternite. Dorsal arc sinuous (Fig. 19). Ventral arc (Fig. 20) fork-shaped, with apophysis as long as arms; arms straight. Tegmen (Figs 24–26) about 0.8 times length of median lobe; distal region divided into parameres; parameres triangular-shaped, apex rounded and covered by short setae; ring piece subrounded, proximal part truncate, without projection. Median lobe (Figs 27–29) slightly curved in lateral view; dorsal lobe with apex rounded and reaching rounded apex of ventral lobe; basal apophysis about 2.3 times length of apical region.



Figs 10–16. 10–12. Details of pronotum of *Ysachron pilosus* gen. et sp. nov. 10. Holotype, ♂ (MPUJ_ENT 0071150). 11. Paratype, ♀ (MPUJ_ENT 0071152). 12. Paratype, ♀ (MZSP). 13–16. Procoxal cavities in Cerambycidae Latreille, 1802 tribes. 13. Achrysonini Lacordaire, 1868, *Ysachron pilosus* gen. et sp. nov. 14. Achrysonini, *Achryson* sp. 15. Hesperophanini Mulsant, 1839, *Hesperophanes* sp. 16. Oemini Lacordaire, 1868, *Oeme* sp.



Figs 17–29. *Ysachron pilosus* gen. et sp. nov., ♂ (MZSP). 17. Tergite VIII. 18. Sternite VIII. 19. Dorsal arc. 20. Ventral arc. 21–23. Tegmen + median lobe. 21. Dorsal view. 22. Lateral view. 23. Ventral view. 24–26. Tegmen. 24. Dorsal view. 25. Lateral view. 26. Ventral view. 27–29. Median lobe. 27. Dorsal view. 28. Lateral view. 29. Ventral view. Scale bars: 17, 20 = 0.5 mm; 18 = 0.3 mm; 19 = 0.1 mm; 21–29 = 1 mm.

Female (Figs 5–9, 11–12)

Antennae shorter, reaching elytral apex at antennomere XII, with scape elongate; prothorax wider, with small gibbosity medially (in one female more evident – Fig. 12); surface of lateral sides of prothorax densely alveolate; ventrite V longer than precedent, with rounded apex.

Measurements (in mm)

Holotype male, total length: 7.5, prothorax length: 1.5, anterior prothoracic width: 1.1, posterior prothoracic width: 1.1, largest prothoracic width: 1.5, elytral length: 5.1, humeral width: 1.7. Paratypes, ♂ / ♀, n = 2 / 2. Total length: 7–8.5 / 8.8–9.1, prothorax length: 1.4–1.8 / 1.6–1.6, largest prothoracic width: 1.35–1.7 / 1.4–2, elytral length: 4.6–5.7 / 6.2–6.5, humeral width: 1.6–1.9 / 2.1–2.2.

New geographical records

Tribe Cerambycini Latreille, 1802

Subtribe Sphallotrichina Martins & Monné, 2002

Amphelictus castaneus Chemsak & Linsley, 1964

Material examined

COLOMBIA • 1 ♂, 1 ♀; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4"W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, light trap; MZSP.

Geographical distribution

Venezuela, Colombia (Cundinamarca, Magdalena), Ecuador, French Guiana, Brazil (Amazonas, Goiás), Bolivia (Santa Cruz). New department record is added: Bolívar (Colombia).

Coleoxestia atrata (Gounelle, 1909)

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, light trap; UARC • 1 spec.; same collection data as for preceding; beating sheet. UARC.

Geographical distribution

Colombia (Amazonas), Brazil (Tocantins, Goiás, Paraná), Bolivia (Santa Cruz). New department record is added: Bolívar (Colombia).

Coleoxestia rubromaculata (Gounelle, 1909)

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Nicaragua, Honduras, Costa Rica, Panama, Colombia, Ecuador, French Guiana, Peru, Brazil (Rondônia, Mato Grosso, Goiás, Distrito Federal). The species is registered from Colombia but without any further locality. Herein, the species is registered from the department of Bolívar (Colombia).

Criodion cinereum (Olivier, 1795)

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Costa Rica, Panama, Colombia (Amazonas), Venezuela, Surinam, French Guiana, Guyana, Peru, Brazil (Amazonas), Bolivia (Santa Cruz), Puerto Rico (?), Paraguay (?). New department record is added: Bolívar (Colombia).

Jupoata robusta Martins & Monné, 2002

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 7 Mar. 2018; K. García leg; tropical dry forest, white light trap; UARC.

Geographical distribution

Brazil (Maranhão, Pernambuco, Bahia, Goiás, Minas Gerais, Rio de Janeiro, São Paulo, Santa Catarina), Costa Rica (Limón). A new country record from Colombia (Atlántico) is added.

Jupoata rufipennis (Gory, 1831)

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Mexico, Nicaragua, Costa Rica, Panama, Trinidad and Tobago, Colombia (Amazonas), Venezuela, Ecuador, Surinam, Guyana, French Guiana, Brazil (Roraima, Amazonas, Pará, Mato Grosso, Goiás, Maranhão, Ceará, Pernambuco, Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul), Peru, Bolivia (Cochabamba, La Paz, Santa Cruz), Paraguay, Argentina (Misiones), Uruguay, Honduras, Guatemala. New department record is added: Bolívar (Colombia).

Tribe Clytini Mulsant, 1839

Cotylytus lebasii (Chevrolat, 1862)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 13 May 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Costa Rica, Panama, Colombia (Bolívar), Venezuela, Nicaragua. New department record is added: Atlántico (Colombia).

Tribe Dichophyiini Gistel, 1848

Chrysoprasis vittata Aurivillius, 1910

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Venezuela, Colombia (Magdalena, Norte de Santander). New department record is added: Bolívar (Colombia).

Monnecles apollinarii (Gounelle, 1913)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 14 May 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Costa Rica, Colombia (Bolívar, Magdalena, Norte de Santander). New department record is added: Atlántico (Colombia).

Tribe Lissonotini Swainson, 1840

Lissonotus corallinus Dupont, 1836

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12 May 2018; K. García leg; tropical dry forest, manual capture; UARC • 1 spec.; same collection data as for preceding; 14 Feb. 2018; fruit trap; UARC • 1 spec.; same collection data as for preceding; 17 Mar. 2018; UARC.

Geographical distribution

Panama, Colombia (Antioquia, Bolívar, Boyacá, Chocó, Cundinamarca, Magdalena, Santander, Tolima, Valle del Cauca), Venezuela, Brazil. New department record is added: Atlántico (Colombia).

Tribe Oemini Lacordaire, 1868

Limernaea ochracea (Fisher, 1927)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–14 Apr. 2018; K. García leg; tropical dry forest, light trap; UARC • 3 ♀♀; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Costa Rica, Panama, Colombia. The species is registered from Colombia but without any further locality. Herein, the species is registered from the departments of Atlántico and Bolívar (Colombia).

Malacopterus tenellus (Fabricius, 1801)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–14 May 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

United States of America (Texas, California), Mexico (Durango, Sonora, Tamaulipas), Guatemala, Belize, Nicaragua, Panama, Colombia (Antioquia, Chocó, Cundinamarca, Meta, Santander, Tolima, Valle del Cauca), Brazil (Amazonas, Pará, Mato Grosso, Maranhão), Venezuela, Ecuador, Guyana, French Guiana, Surinam, Peru, Bolivia (La Paz, Santa Cruz), Jamaica, Cuba, Guadeloupe, Hispaniola, Honduras, Costa Rica, Dominican Republic. New department record is added: Atlántico (Colombia).

Tribe Rhopalophorini Blanchard, 1845

Ischionodonta colombiana Napp & Marques, 1999

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Colombia (Magdalena). New department record is added: Bolívar (Colombia).

Tribe Sydacini Martins, 2014

Sydax gibbus Joly, 1985

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–14 May 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Venezuela, Colombia (Bolívar). New department record is added: Atlántico (Colombia).

Tribe Torneutini Thomson, 1861

Diploschema mandibulare Fuchs, 1964

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC • 1 ♀; same collection data as for preceding; manual capture; MZSP.

Geographical distribution

Panama, Colombia, Venezuela, French Guiana, Guyana, Peru, Brazil (Amazonas, Pará, Mato Grosso), Ecuador, Bolivia (Santa Cruz), Paraguay. The species is registered from Colombia but without further locality. Herein, the species is registered from the department of Bolívar (Colombia).

Diploschemopsis howdeni (Martins & Monné, 1980)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 17–18 Mar. 2018; K. García leg; tropical dry forest, light trap; UARC • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 17–18 Feb. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Colombia (Magdalena, Antioquia), Venezuela. New department records are added: Atlántico and Bolívar (Colombia).

Tribe Trachyderini Dupont, 1836

Ceragenia insulana Fisher, 1943

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 15–16 Apr. 2018; K. García leg; tropical dry forest, beating sheet; UARC • 1 spec.; same collection data as for preceding; fruit trap; UARC • 1 spec.; same collection data as for preceding; 18 Feb. 2018; UARC • 1 spec.; same collection data as for preceding; 15 Mar. 2018; UARC.

Geographical distribution

Costa Rica, Panama, Colombia (Magdalena). New department record is added: Bolívar (Colombia).

Oxymerus aculeatus lebasii Dupont, 1838

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 15 May 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Mexico, Nicaragua, Costa Rica, Panama, Colombia (Antioquia, Bolívar, Chocó, Cundinamarca, Magdalena, Meta, Norte de Santander, Tolima, Valle del Cauca), Venezuela, French Guiana, Surinam, Guyana, Aruba, Curaçao, St. Vincent, Grenada, Guadeloupe, Cuba, Montserrat. Mustique, Jamaica, Peru, Argentina, Brazil. New department record is added: Atlántico (Colombia).

Subfamily Lamiinae Latreille, 1825

Tribe Aerenicini Lacordaire, 1872

Phaula thomsonii Lacordaire, 1872

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 15 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Brazil (Pará, Mato Grosso do Sul, Goiás, Maranhão, Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul), Bolivia (Santa Cruz), Paraguay, Argentina (Misiones). A new country record from Colombia (Bolívar) is added.

Tribe Agapanthiini Mulsant, 1839

Aphies secunda (Tippmann, 1951)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 10 Apr. 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Costa Rica, Venezuela. A new country record from Colombia (Atlántico) is added.

Hippopsis freyi Breuning, 1955

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–13 May 2018; K. García leg; tropical dry forest, manual capture, beating sheet and UV light trap; UARC • 1 spec.; same collection data as for preceding; 10 Jun. 2018; UARC • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 18 Feb. 2018; K. García leg; tropical dry forest, light trap; UARC • 1 spec.; same collection data as for preceding; 15 Jun. 2018; UV light trap; UARC.

Geographical distribution

Trinidad & Tobago, Venezuela, Colombia. The species is registered from Colombia but without further locality. Herein, the species is registered from the departments of Atlántico and Bolívar (Colombia).

Hippopsis lemniscata lemniscata (Fabricius, 1801)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–13 May 2018; K. García leg; tropical dry forest, beating sheet; UARC.

Geographical distribution

Canada, Eastern United States of America to Texas, Mexico, Guatemala, Honduras, Nicaragua, Brazil, Panama, Venezuela, Argentina, Bolivia. A new country record from Colombia (Atlántico) is added.

Hippopsis ocularis Galileo & Martins, 1995

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 10 Jun. 2018; K. García leg; tropical dry forest, beating sheet; UARC.

Geographical distribution

Bolivia (Cochabamba, Santa Cruz); French Guiana. A new country record from Colombia (Atlántico) is added.

Hippopsis septemlineata Breuning, 1940

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12 May 2018; K. García leg; tropical dry forest, beating sheet; UARC.

Geographical distribution

Panama, Colombia, Ecuador. The species is registered from Colombia but without further locality. Herein, the species is registered from the department of Atlántico (Colombia).

Hippopsis septemvittata Breuning, 1940

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 16 Jun. 2018; K. García leg; tropical dry forest, beating sheet; UARC.

Geographical distribution

Brazil. A new country record from Colombia (Bolívar) is added.

Tribe Apomecynini Thomson, 1860

Bebelis fasciata (Fisher, 1947)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 10 Jun. 2018; K. García leg; tropical dry forest, beating sheet; UARC.

Geographical distribution

Panama. A new country record from Colombia (Atlántico) is added.

Bebelis picta Pascoe, 1875

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12 May 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Mexico (Veracruz, Chiapas, Guerrero), Guatemala, Nicaragua, Honduras, Cuba, Costa Rica, Panama, Venezuela, Colombia, French Guiana, Brazil (Maranhão, Goiás, Ceará, Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Rio Grande do Sul), Bolivia (Santa Cruz), Paraguay, Argentina (Buenos Aires), Martinique, St. Vincent. New department record is added: Atlántico (Colombia).

Dorcasta dasycera (Erichson, 1849)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 9 Jun. 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Mexico (Oaxaca, Jalisco), Guatemala, Honduras, Nicaragua, Costa Rica, Panama, Aruba, Colombia (Cesar), Venezuela, Guyana, French Guiana, Brazil (Pará), Bolivia (Santa Cruz). New department record is added: Atlántico (Colombia).

Ischioloncha strandiella Breuning, 1942

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 9 Jun. 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Bolivia. A new country record from Colombia (Atlántico) is added.

Tribe Colobotheini Thomson, 1860

Colobothea fasciatipennis Linsley, 1935

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 18 Mar. 2018; K. García leg; tropical dry forest, light trap; UARC • 1 spec.; same collection data as for preceding; 9 Apr. 2018; manual capture; UARC.

Geographical distribution

Honduras, Panama, Colombia (Bolívar, Chocó, Santander). New department record is added: Atlántico (Colombia).

Tribe Desmiphorini Thomson, 1860

Desmiphora (Desmiphora) canescens Bates, 1874

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 13 May 2018; K. García leg; tropical dry forest, beating sheet; UARC.

Geographical distribution

Mexico (Chiapas), Guatemala, Honduras, Nicaragua, Costa Rica, Panama, Colombia (Bolívar), Venezuela. New department record is added: Atlántico (Colombia).

Estoloides venezuelensis Breuning, 1942

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 15 Feb. 2018; K. García leg; tropical dry forest, UV light trap; UARC • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 16 Mar. 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Venezuela. A new country record from Colombia (Atlántico and Bolívar) is added.

Mimasyngenes icuapara Galileo & Martins, 1996

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12 May 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Costa Rica, Colombia, Venezuela, Brazil (São Paulo), Argentina (Misiones). The species is registered from Colombia but without further locality. Herein, the species is registered from the department of Atlántico (Colombia).

Tribe Hemilophini Thomson, 1868

Adesmus divus (Chabrillac, 1857)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 14 May 2018; K. García leg; tropical dry forest, manual capture; UARC.

Geographical distribution

Colombia (Guajira), Brazil (Mato Grosso, Goiás, Distrito Federal, Maranhão, Minas Gerais, Espírito Santo Rio de Janeiro, São Paulo, Paraná, Rio Grande do Sul), French Guiana, Bolivia (Santa Cruz), Paraguay, Argentina (Misiones). New department record is added: Atlántico (Colombia).

Phoebe birai Galileo, 2015

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12 May 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Bolivia (Santa Cruz). A new country record from Colombia (Atlántico) is added.

Tribe Lamiini Latreille, 1825

Deliathis quadritaeniata (White, 1846)

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Costa Rica, Panama, Colombia Venezuela, Ecuador. The species is registered from Colombia but without any specific locality. Herein, the species is registered from the department of Bolívar (Colombia).

Tribe Onciderini Thomson, 1860

Cacostola colombiana Martins & Galileo, 1999

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–13 May 2018; K. García leg; tropical dry forest, beating sheet, light trap; UARC • 1 spec.; same collection data as for preceding; 10 Jun. 2018; UARC.

Geographical distribution

Colombia (Bolívar). New department record is added: Atlántico (Colombia).

Hypsioma nesiope Dillon & Dillon, 1945

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Panama, Colombia (Magdalena). New department record is added: Bolívar (Colombia).

Lochmaeocles tessellatus tessellatus (Thomson, 1868)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–14 May 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Panama, Colombia (Antioquía, Bolívar, Cundinamarca, Magdalena, Tolima), Venezuela. New department record is added: Atlántico (Colombia).

Oncideres sobrina Dillon & Dillon, 1946

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 18 Feb. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Panama, Colombia (Magdalena, Santander). New department record is added: Bolívar (Colombia).

Tulcus fulvofasciatus (Dillon & Dillon, 1945)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–14 May 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Costa Rica, Panama, Colombia. The species is registered from Colombia but without any specific locality. Herein, the species is registered from the department of Atlántico (Colombia).

Tribe Phacellini Lacordaire, 1872

Piola colombica Martins & Galileo, 1999

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–13 May 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Colombia (Bolívar). New department record is added: Atlántico (Colombia).

Subfamily Prioninae Latreille, 1802
Tribe Macrotomini Lameere, 1912

Mallodon dasystemus (Say, 1824)

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

Southern United States of America, Mexico (Jalisco, Nayarit, Sinaloa, Tamaulipas, Veracruz), Honduras, Tobago, Colombia. The species is registered from Colombia but without any specific locality. Herein, the species is registered from the department of Bolívar (Colombia).

Physopleurus erikae Santos-Silva & Martins, 2009

Material examined

COLOMBIA • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13 Apr. 2018; K. García leg; tropical dry forest, light trap; UARC.

Geographical distribution

Colombia (Valle del Cauca). New department record is added: Bolívar (Colombia).

Strongylaspis corticaria (Erichson, 1849)

Material examined

COLOMBIA • 1 spec.; Atlántico, Usiacurí, Reserva Campesina La Montaña; 10°46'2.6" N, 75°0.2'34" W; 260 m a.s.l.; 12–13 May 2018; K. García leg; tropical dry forest, UV light trap; UARC • 1 spec.; Bolívar, San Jacinto, Reserva La Flecha; 09°51'12.4" N, 75°10'41.4" W; 324 m a.s.l.; 13–16 Apr. 2018; K. García leg; tropical dry forest, UV light trap; UARC.

Geographical distribution

United States of America (Florida), Mexico (Veracruz), Guatemala, Belize, Honduras, El Salvador, Nicaragua, Costa Rica, Panama, Colombia, Guyana, French Guiana, Venezuela, Brazil (Roraima), Cuba, Jamaica, Dominica, Martinique, St. Lucia, Dominican Republic. The species is registered from Colombia but without any specific locality. Herein, the species is registered from the departments of Atlántico and Bolívar (Colombia).

Discussion

Colombia is considered one of the 12 countries with the greatest biological biodiversity in the world, it has a prodigious natural richness and, although it only represents 0.7% of the world's land surface, concentrates in its territory about 10% of the planet's biodiversity (Mittermeier & Goettsch 1997; Colciencias 2016).

Currently, in Colombia there are about 920 known species of Cerambycidae (Botero 2018). Considering the richness of the country in terms of biodiversity and when comparing to the Cerambycidae fauna of other better-studied countries in the region, there is no doubt that the number of Cerambycidae species and the knowledge of their distribution in Colombia should continue to increase in upcoming years.

With the description of the new genus herein, Achrysonini now comprises ten genera and 32 species in South America.

Acknowledgments

We are deeply thankful to Sônia Casari for her kindness and help in the MZSP during the visit of the first author in 2018; to the members of Neoptera, especially to Neis Martínez, Jeniffer Meriño and José Sarmiento (UARC), for their help in collecting the specimens used in this study; and to all the curators of the collections visited. The first author thanks “Fundação de Amparo à Pesquisa do Estado de São Paulo” (FAPESP) for a magister fellowship (process number 2019/13603-1). The second author also thanks “Fundação de Amparo à Pesquisa do Estado de São Paulo” (FAPESP) for a postdoctoral fellowship (process number 2017/17898-0).

References

- Bates H.W. 1870. Contributions to an insect fauna of the Amazon Valley (Coleoptera: Cerambycidae). *The Transactions of the Entomological Society of London* 18 (3): 243–335.
<https://doi.org/10.1111/j.1365-2311.1870.tb01876.x>
- Botero J.P. 2018. La familia Cerambycidae (Coleoptera: Chrysomeloidea) en Colombia. In: Deloya C. & Gasca H.J. (eds) *Escarabajos del Neotrópico (Insecta: Coleoptera)*: 153–169. S y G Editores, Ciudad de México.
- Colciencias. 2016. Colombia, el segundo país más biodiverso del mundo. Available from https://minciencias.gov.co/sala_de_prensa/colombia-el-segundo-pais-mas-biodiverso-del-mundo [accessed 14 Aug. 2020].
- Ehara S. 1954. Comparative anatomy of male genitalia in some Cerambycid beetles. *Journal of the Faculty of Science, Hokkaido University (Zoology)* 12: 61–115.
Available from <http://hdl.handle.net/2115/27139> [accessed 4 Feb. 2021].
- Lacordaire J.T. 1868. *Histoire Naturelle des Insectes. Genera des Coléoptères, ou exposé méthodique et critique de tous les genres proposés jusqu'ici dans cet ordre d'insectes*. Vol. 8. Librairie encyclopédique de Roret, Paris. <https://doi.org/10.5962/bhl.title.8864>
- Lawrence J.F., Beutel R.G., Leschen R.A.B. & Ślipiński A. 2010. Glossary of morphological terms. In: Leschen R.A.B, Beutel R.G. & Lawrence J.F. (eds) *Handbook of Zoology, Arthropoda Insecta. Coleoptera, Beetles, Morphology and Systematics* Vol. 2: 9–20. De Gruyter, Berlin and New York.
- Martínez C. 2000. Escarabajos longicornios (Coleoptera, Cerambycidae) de Colombia. *Biota Colombiana* 1 (1): 76–105.
- Martins U.R. 1997. Tribo Oemini. In: Martins U.R. (ed.) *Cerambycidae Sul-Americanos (Coleoptera)*. *Taxonomia* Vol. 1: 3–155. Sociedade Brasileira de Entomologia, São Paulo.
- Martins U.R. 1998. Tribo Ectenessini. In: Martins U.R. (ed.) *Cerambycidae Sul-Americanos (Coleoptera)*. *Taxonomia* Vol 2: 82–183. Sociedade Brasileira de Entomologia, São Paulo.
- Martins U.R. 2002. Tribo Achrysonini. In: Martins U.R. (ed.) *Cerambycidae Sul-Americanos (Coleoptera)*. *Taxonomia* Vol. 5: 37–144. Sociedade Brasileira de Entomologia, São Paulo.
- Martins U.R. & Galileo M.H.M. 1999. Tribo Hesperophanini. In: Martins U.R. (ed.) *Cerambycidae Sul-Americanos (Coleoptera)*. *Taxonomia* Vol. 3: 1–117. Sociedade Brasileira de Entomologia, São Paulo.
- Mittermeier R.A. & Goettsch, C. 1997. *Megadiversidad. Los Países Biológicamente más Ricos del Mundo*. Cemex, Ciudad de México.

Monné M.A. 2020. Catalogue of the Cerambycidae (Coleoptera) of the Neotropical region. Part I. Subfamily Cerambycinae. Available from <https://cerambycids.com/catalog> [accessed 3 Aug. 2020].

Monné M.L. & Monné M.A. 2004. *Neoachryson*, novo gênero de Achrysonini (Coleoptera, Cerambycidae, Cerambycinae). *Iheringia, Série Zoologia* 94 (2): 133–134.

<https://doi.org/10.1590/S0073-47212004000200003>

Švácha P. & Lawrence J.F. 2014. 2.4 Cerambycidae Latreille, 1802. In: Leschen R.A.B. & Beutel R.G. (eds) *Handbook of Zoology, Arthropoda: Insecta; Coleoptera, beetles, Volume. 3: Morphology and systematics (Phytophaga)*: 77–177. Walter de Gruyter, Berlin.

Tavakilian G.L. & Chevillotte H. 2020. Titan: base de données internationales sur les Cerambycidae ou Longicornes. Version 3.0. Available from <http://titan.gbif.fr> [accessed 3 Aug. 2020].

Manuscript received: 25 August 2020

Manuscript accepted: 26 November 2020

Published on: 19 February 2021

Topic editor: Nesrine Akkari

Section editor: Max Barclay

Desk editor: Pepe Fernández

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