



Research article

urn:lsid:zoobank.org:pub:F8C5C34F-CCBB-420B-9319-1DA224A5D783

Revision of the *Merodon bombiformis* group (Diptera: Syrphidae) – rare and endemic African hoverfliesAnte VUJIĆ¹, Snežana RADENKOVIĆ², Ljiljana ŠAŠIĆ ZORIĆ³, Laura LIKOV⁴,
Tamara TOT⁵, Sanja VESELIC^{*,6} & Mihajla DJAN⁷^{1,2,4,5,6,7}University of Novi Sad, Faculty of Sciences, Department of Biology and Ecology,
Trg Dositeja Obradovića 2, 21000 Novi Sad, Serbia.³University of Novi Sad, BioSense Institute, Dr Zorana Đinđića 1, 21000 Novi Sad, Serbia.

*Corresponding author email: sanja.veselic@dbe.uns.ac.rs

¹Email: ante.vujic@dbe.uns.ac.rs²Email: snezana.radenkovic@dbe.uns.ac.rs³Email: ljsasic@biosense.rs⁴Email: laura.likov@dbe.uns.ac.rs⁵Email: tamaratot90@gmail.com⁷Email: mihajla.djan@dbe.uns.ac.rs¹ <https://orcid.org/0000-0002-8819-8079>² <https://orcid.org/0000-0002-7805-9614>³ <https://orcid.org/0000-0003-1616-2701>⁴ <https://orcid.org/0000-0002-7215-1006>⁵ <https://orcid.org/0000-0001-8776-9362>⁶ <https://orcid.org/0000-0002-7613-8052>⁷ <https://orcid.org/0000-0002-2427-0676>¹urn:lsid:zoobank.org:author:E80A5C9B-5450-4E15-ACEB-FC3B94CF2244²urn:lsid:zoobank.org:author:26DF35D9-55FA-4485-8E8C-C1F90EFE1036³urn:lsid:zoobank.org:author:A0EFB962-F816-4399-A5F2-67839DD19CE8⁴urn:lsid:zoobank.org:author:6787F9E8-3EA9-4695-B8A1-725787BCB9DD⁵urn:lsid:zoobank.org:author:E4436AC7-B3B6-4899-838D-5A6ECD8E6BFB⁶urn:lsid:zoobank.org:author:625C1317-F45B-46E8-B55E-51032B1F7C67⁷urn:lsid:zoobank.org:author:BE29D9A9-CC69-416C-8F85-4B8AA99A6C71

Abstract. In the present work, the Afrotropical species of the *bombiformis* species group, part of the *aureus* lineage, are revised. Six species are recognized, based on a combination of morphological and genetic features. Three of these species are new to science: *Merodon lotus* Vujić & Radenković sp. nov., *M. vittatus* Vujić & Likov sp. nov., and *M. zebra* Vujić & Radenković sp. nov. Redescriptions are provided for the other three species: *M. bombiformis* Hull, 1944, *M. multifasciatus* Curran, 1939, and *M. nasicus* Bezzi, 1915. The female of *M. bombiformis* is described. The name *Merodon edentulus* Macquart, 1855 is considered here as a nomen dubium. One new synonymy is proposed: *M. apimima* Hull, 1944 syn. nov. (junior synonym of *M. multifasciatus*). The distribution of the *bombiformis* species

group is discussed. The larval host plant of *M. multifasciatus* is identified as *Gladiolus*. A key to the identification of both males and females of the *bombiformis* group is provided.

Keywords. Afrotropical region, *Merodon aureus* lineage, molecular data, new species, syrphids, hoverfly, flower fly.

Vujić A., Radenković S., Šašić Zorić Lj., Likov L., Tot T., Veselić S. & Djan M. 2021. Revision of the *Merodon bombiformis* group (Diptera: Syrphidae) – rare and endemic African hoverflies. *European Journal of Taxonomy* 755: 88–135. <https://doi.org/10.5852/ejt.2021.755.1401>

Introduction

The phytophagous hoverfly genus *Merodon* Meigen, 1803 (Diptera: Syrphidae: Eristalinae: Merodontini) is distributed across the Palaearctic and Afrotropical regions and comprises more than 160 species (Ståhls *et al.* 2009; Vujić *et al.* 2012). *Merodon equestris* (Fabricius, 1794) as an introduced anthropogenic species from Europe is also widely distributed across the Nearctic (Speight 2020). The highest diversity of genus *Merodon* is recorded around the Mediterranean basin, a phenomenon which has been associated with the great variety of geophytes in this region, as those are the host plants of larvae of *Merodon* (Andrić *et al.* 2014; Ricarte *et al.* 2017). Although the immature stages of only eight species of *Merodon* have been described and the larval host plants are postulated only for eight additional species, based on still incomplete evidence, the connection of larvae of *Merodon* with underground storage organs of the family Hyacinthaceae Batsch ex Borkh. is well recognized (Ricarte *et al.* 2008, 2017; Andrić *et al.* 2014; Preradović *et al.* 2018). It might be expected that the congruence between geophyte diversity and species-richness of *Merodon*, highlighted by Ricarte *et al.* (2017) for the European Mediterranean zone, would be paralleled in South Africa, since it is there that the geophyte diversity is at its maximum (Pfosser & Speta 2004). But, surprisingly, nowhere within the Afrotropical Region is there a high diversity of species of *Merodon*.

Only 15 species consigned to the genus *Merodon* have been reported from the Afrotropical Region: *Merodon apimima* Hull, 1944, *M. bombiformis* Hull, 1944, *M. capensis* Hurkmans, 2018, *M. capi* Vujić & Radenković, 2020, *M. commutabilis* Radenković & Vujić, 2018, *M. cuthbertsoni* Curran, 1939, *M. drakonis* Vujić & Radenković, 2018, *M. edentulus* Macquart 1855, *M. flavocerus* Hurkmans, 2018, *M. melanocerus* Bezzi, 1915, *M. multifasciatus* Curran, 1939, *M. nasicus* Bezzi, 1915, *M. planifacies* Bezzi, 1915, *M. roni* Radenković & Vujić, 2020 and *M. stevensoni* Curran, 1939 (Smith & Vockeroth 1980; Radenković *et al.* 2018; Djan *et al.* 2020; Evenhuis & Pape 2020). The low species diversity of *Merodon* in South Africa is the opposite of what might be anticipated (Radenković *et al.* 2018).

Bringing together previously published data (Šašić *et al.* 2016; Radenković *et al.* 2018), Vujić *et al.* (2019a), recognised five monophyletic lineages within the genus *Merodon*: *albifrons*, *aureus*, *avidus-nigratarsis*, *desuturinus*, and *natans*. In the Afrotropical region, representatives of only two of these lineages are present, namely *aureus* and *desuturinus*.

The *desuturinus* lineage represents an important link between the Palaearctic and Afrotropical faunas. Palaearctic representatives of the group were revised by Vujić *et al.* (2018), resulting in the delimitation of four species: *Merodon cabanerensis* Marcos-García, Vujić & Mengual, 2007 from Spain and Morocco, *M. desuturinus* Vujić, Šimić & Radenković, 1995, a Balkan endemic from high mountains, *M. neolydicus* Vujić, 2018 from the Eastern Mediterranean and Iran, and *M. murorum* (Fabricius, 1794), from North-West Africa. The Afrotropical component of the *desuturinus* lineage comprises 10 taxa: *M. cuthbertsoni*, plus the *M. melanocerus* subgroup with five species (*M. capensis*, *M. commutabilis*, *M. drakonis*, *M. flavocerus* and *M. melanocerus*) (Radenković *et al.* 2018) and the

M. planifacies subgroup comprising two previously known (*M. planifacies* and *M. stevensoni*) and two recently recognized cryptic taxa: *M. capi* and *M. roni* (Djan *et al.* 2020).

The rest of the Afrotropical species of the genus *Merodon* belong to the *aureus* lineage. Šašić *et al.* (2016) defined species groups within this lineage, such as the *M. aureus*, *M. bombiformis*, *M. funestus*, *M. nanus* and *M. spinatarsis* species groups. Revision of all available material from museums and detailed analyses of newly collected specimens from our own expeditions to RSA resulted in the delimitation of the six species considered here, all belonging to the *M. bombiformis* group.

The aims of the present study were:

- 1) To review material belonging to the *bombiformis* group held in the collections of several entomological institutions and private individuals,
- 2) To define and describe the taxa within the *bombiformis* species group, including mapped geographic distributions and new species,
- 3) To examine the genetic diversity exhibited among the members of this species group and infer the phylogenetic relationship, using one mitochondrial gene (cytochrome c oxidase subunit I (COI)) and two nuclear genes (28S rRNA and 18S rRNA),
- 4) To present an illustrated key and a discussion on the different taxonomic characters used,
- 5) To discuss the larval host plant data available for the *bombiformis* species group.

Material and methods

Morphological study

The present study is based on the examination of all available material of the *Merodon bombiformis* species group deposited in the museums and universities listed below. The following abbreviations of museums and entomological collections are used in the text to indicate where the material studied is deposited:

- AMNH = American Museum of Natural History, New York, USA
BMNH = The Natural History Museum, London, United Kingdom
CWM = Michael de Coursy Williams collection, Alexandroupoli, Greece
FSUNS = Department of Biology and Ecology, Faculty of Sciences, University of Novi Sad, Novi Sad, Serbia
LSF = Museo Zoologico La Specola, Florence, Italy
MNHN = Muséum national d'histoire naturelle, Paris, France
NBCN = Naturalis Biodiversity Center, Leiden, The Netherlands
NHMW = Naturhistorisches Museum Wien, Vienna, Austria
NMSA = Natal Museum, Pietermaritzburg, Republic of South Africa
RBINS = Royal Belgian Institute of Natural Sciences, Brussels, Belgium
RMCA = Royal Museum for Central Africa, Tervuren, Belgium
TAUI = Tel Aviv University, Tel Aviv, Israel
USNM = Department of Entomology, National Museum of Natural History, Smithsonian Institution, Washington, DC, USA

Numbers given alongside the abbreviation FSUNS in the examined material refer to unique identifiers of the specimens, stored in the database of the Faculty of Sciences, University of Novi Sad. The distribution map (Fig. 1) was generated with the mapping software DIVA-GIS (Hijmans *et al.* 2012). Relationships of species to different vegetation types were based on the map of natural vegetation of Africa (Sayre *et al.* 2013).

The terminology used in the descriptions and drawings follows Thompson (1999) for non-genitalic features except for the terms “proepimeron” and “pleuron”, which follows McAlpine (1981), and those relating to male genitalia follow Marcos-García *et al.* (2007), while the term “fossette” follows Doczkal & Pape (2009).

For the study of male genitalia, dry specimens were relaxed in a humidity chamber. The genitalia were extracted with a hook-tipped entomological pin before being cleared by boiling in warm 10% potassium hydroxide (KOH) for 3–5 min. This was followed by brief immersion in acetic acid to neutralize the KOH, and then by immersion in ethanol to remove the acid. The prepared genitalia were stored in microvials containing glycerol pinned to the source specimen.

Digital photographs were taken with a Leica DFC 320 digital camera attached to a Leica MZ16 stereo microscope, while for drawings a Leica MZ16 binocular microscope was used with an FSA 25 PE drawing tube. The photographs were stacked using CombineZ software (Hadley 2006).

Body length was measured in lateral view in millimetres (mm) using an eyepiece micrometer from the lunula to the end of the abdomen. Wing length was measured from the tegula to the apex of the wing.

Abbreviations

- ams = anterior margin of surstyle
- c = cercus
- ea = ejaculatory apodeme
- p = aedeagus
- pl = posterior lobe of surstyle
- s = lateral sclerite of aedeagus

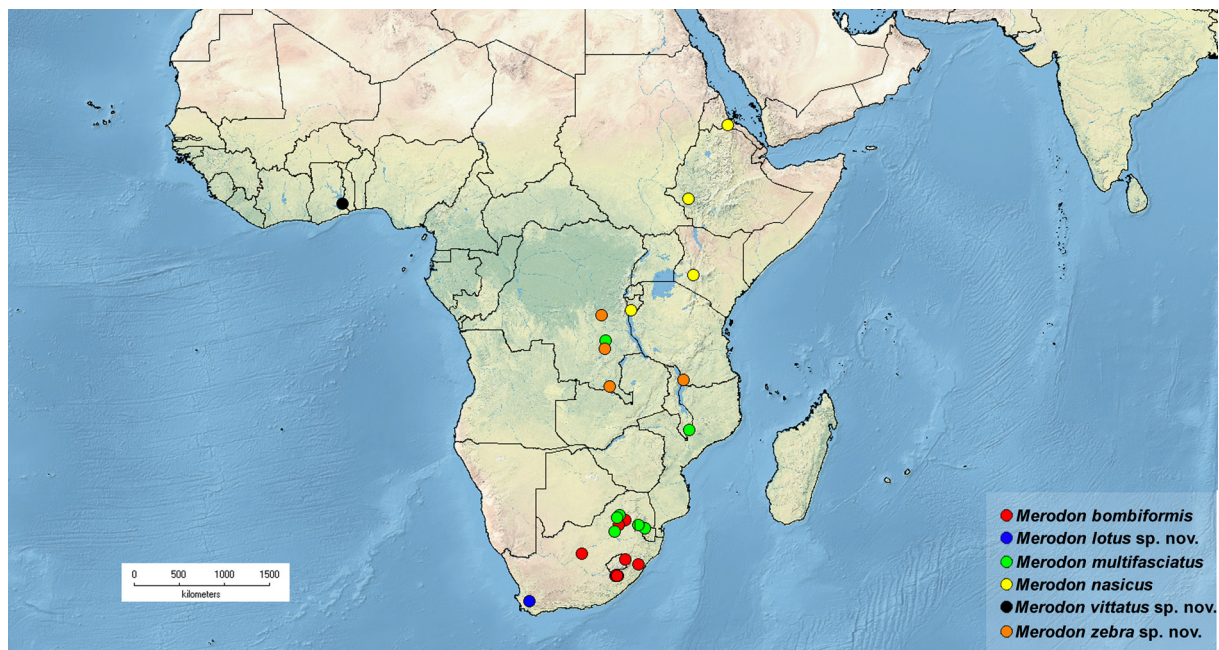


Fig. 1. Distribution map of the *Merodon bombiformis* species group.

Molecular analysis

The genomic DNA of six hoverfly specimens, belonging to three species from the *Merodon bombiformis* species group (*M. bombiformis*, *M. multifasciatus* and *M. lotus* Vujić & Radenković sp. nov.) was extracted from mid and hind legs using the sodium dodecyl sulfate (SDS) extraction protocol described by Chen *et al.* (2010). Fresh material required for high-quality DNA extraction was only available for these species. Species of the *M. bombiformis* group for which we were unable to obtain fresh material were not included. Genomic DNA vouchers are deposited at the Faculty of Sciences, Department of Biology and Ecology, University of Novi Sad (FSUNS).

We amplified one mitochondrial (COI gene) and two nuclear genes (28S rRNA and 18S rRNA genes). The Folmer fragment (DNA barcode) on 5'COI gene was amplified using LCO1490 and HCO2198 primer pair (Folmer *et al.* 1994), the 28S rRNA gene using F2 and 3DR primer pair (Belshaw *et al.* 2001), while for amplification of 18S rRNA we used 2F and b2.9 primer pair designed by The Crandall Lab (for primer sequences see Vujić *et al.* 2019b). Polymerase chain reactions (PCR) were carried out in 25 µl reaction volumes. The reaction mixture contained Dream Taq buffer (ThermoScientific, Vilnius, Lithuania), 0.1 mM of each nucleotide, 5 pmol of each primer, 1.25U Dream Taq polymerase (ThermoScientific, Vilnius, Lithuania), ~50–100 ng of template DNA and ultrapure water. PCR amplifications were performed using the following conditions: 95°C for 2 min; 29 cycles of 94°C for 30 s each, 50°C for 30 s; 72°C for 2 min; with a final extension at 72°C for 8 min. PCR products were purified using Exonuclease I and FastAP Thermosensitive Alkaline Phosphatase (Thermo Scientific, Vilnius, Lithuania) according to the manufacturer's instructions. The PCR fragments were commercially sequenced in the forward direction by Macrogen Europe (Amsterdam, Netherlands).

Sequence processing and phylogenetic analysis

The COI, 28S rRNA and 18S rRNA gene sequences were edited for base-call errors using BioEdit ver. 7.0.9.0. (Hall 1999). Additional sequences of several species representing the main *Merodon* lineages following Radenković *et al.* (2018) were downloaded from GenBank and joined to the sequence matrices of all tree genes. The COI and 18S rRNA gene sequences were aligned manually, while 28S rRNA gene sequences were aligned using MAFFT ver. 7 (Kato & Standley 2013), available from the European Bioinformatics Institute (EMBL-EBI) web services (McWilliam *et al.* 2013). The aligned COI, 28S rRNA and 18S rRNA gene sequences were concatenated and combined into a single sequence matrix. The ML tree was constructed using RAxML ver. 8.2.8 (Stamatakis 2014) using the CIPRES Science Gateway web portal (Miller *et al.* 2010) under the general time-reversible (GTR) evolutionary model with a gamma distribution (GTRGAMMA) (Rodríguez *et al.* 1990) following instructions in the manual. The applied command line was as follows: raxmlHPC-HYBRID -T 4 -n result -s infile.txt -m GTRGAMMA -p 12345 -q part.txt -f a -N 1000 -x 12345 -o E_pulchel. Nodal supports were estimated using rapid bootstrapping with 1000 replicates. The tree is rooted with *Eumerus pulchellus* Loew, 1848 as outgroup. The uncorrected pairwise (p) distance matrix for 5' COI gene sequences among specimens of the *M. bombiformis* species group was estimated using MEGA ver. 7 software (Kumar *et al.* 2016). GenBank accession numbers of all sequences are provided in Supp. file 1.

Results

Taxonomic revision

Phylum Arthropoda Latreille, 1829
Class Insecta Linnaeus, 1758
Order Diptera Linnaeus, 1758
Family Syrphidae Latreille, 1802
Subfamily Eristalinae Newman, 1834
Tribus Merodontini Edwards, 1915

Genus *Merodon* Meigen, 1803

Diagnosis of the *Merodon aureus* lineage

Mid coxa with long pile posteriorly (Fig. 2A); anterior anepisternum below postpronotum with many long pile (Fig. 2B); species with stocky abdomen (Fig. 2C); lateral sclerites of the aedeagus very small or absent (Fig. 3A: marked with arrow).

Diagnosis of the *Merodon bombiformis* group

Pedichel elongated, approximately as long as basoflagellomere (Fig. 4); abdomen broad (Fig. 5); metafemur with less serrated apicoventral triangular lamina, usually only the apical dens is distinct (Fig. 6); metatrochanter of males smooth, without calcar (Fig. 6); male genitalia with posterior surstyle lobe usually bent (as on Fig. 7A, D: pl), and hypandrium narrowed medially (as on Fig. 7C: marked with arrow); distribution: Afrotropical Region (Fig. 1).

The *Merodon bombiformis* group and *M. funestus* (Fabricius, 1794) differ from other species and groups among the *aureus* lineage by an elongated pedichel, approximately as long or even longer than basoflagellomere (Fig. 4) and small lateral sclerite of the aedeagus (as on Fig. 3B, D: s) (absent in other species and groups of the *aureus* lineage, on Fig. 3A: marked with arrow). Morphologically, the *M. bombiformis* group can be distinguished from *M. funestus* by the absence of a calcar on metatrochanter in males (present in *M. funestus*, on Fig. 8A), less dentate apicoventral triangular lamina on the metafemur, but usually with a distinct apical dens (Fig. 8B) (clearly dentate in *M. funestus*, on Fig. 8A) and by the shape of the posterior lobe of the surstylus: tip rounded in *bombiformis* group (as on Fig. 13A, D: pl), but tapering in *M. funestus* (Fig. 19A: pl).

Merodon bombiformis Hull, 1944

Figs 1, 3C, 4A, 5A, 6A, 9A, 10A, 11A, 12A, 13A–C, 14A, 15A, 16A, 21A–D, 24

Merodon bombiformis Hull, 1944: 42–43 (type locality: Waterval, Republic of South Africa).

Diagnosis

Large, golden species (10–14 mm) with reddish-yellow/golden face and frons (Figs 9A, 11A), lateral sides of scutum (Fig. 10A) and most of terga (Fig. 5A); oral margin reduced (Fig. 9A); antennal segments very short (Fig. 4A), frons inflated (Fig. 9A).

Material examined

Holotype

REPUBLIC OF SOUTH AFRICA • 1 ♂; Waterval; 25.183° S, 29.012° E; 27 Dec. 1898; Distant Collection 1911:383; BMNH.

Additional material

REPUBLIC OF SOUTH AFRICA • 1 ♂; Distant Bush; 28.586° S, 24.658° E; 1182 m a.s.l.; 1911; unknown leg.; NMSA • 1 ♂; Pretoria, Baviaanspoort; 25.664° S, 28.366° E; 1290 m a.s.l.; 15 Feb. 1920; H.K. Munro leg.; FSUNS ID 04277; NMSA • 1 ♀; Natal, Hilton Road; 29.597° S, 30.336° E; 833 m a.s.l.; 22 Dec. 1953; P. Graham leg.; FSUNS ID 04280; NMSA • 1 ♀; Pietermaritzburg; 29.597° S, 30.336° E; 833 m a.s.l.; 5 Nov. 1955; Oosthuizen leg.; reared from *Gladiolus* bulbs; FSUNS ID 04282; NBCN LML-05-5 • 1 ♂; same collection data as for preceding; FSUNS ID 04276; NMSA • 1 ♀; same collection data as for preceding; 19 Nov. 1955; FSUNS ID 04279; NMSA • 1 ♂; reared from *Gladiolus* bulbs in Wisley Horticultural Gardens (United Kingdom), which were brought from RSA; Jun. 1956;

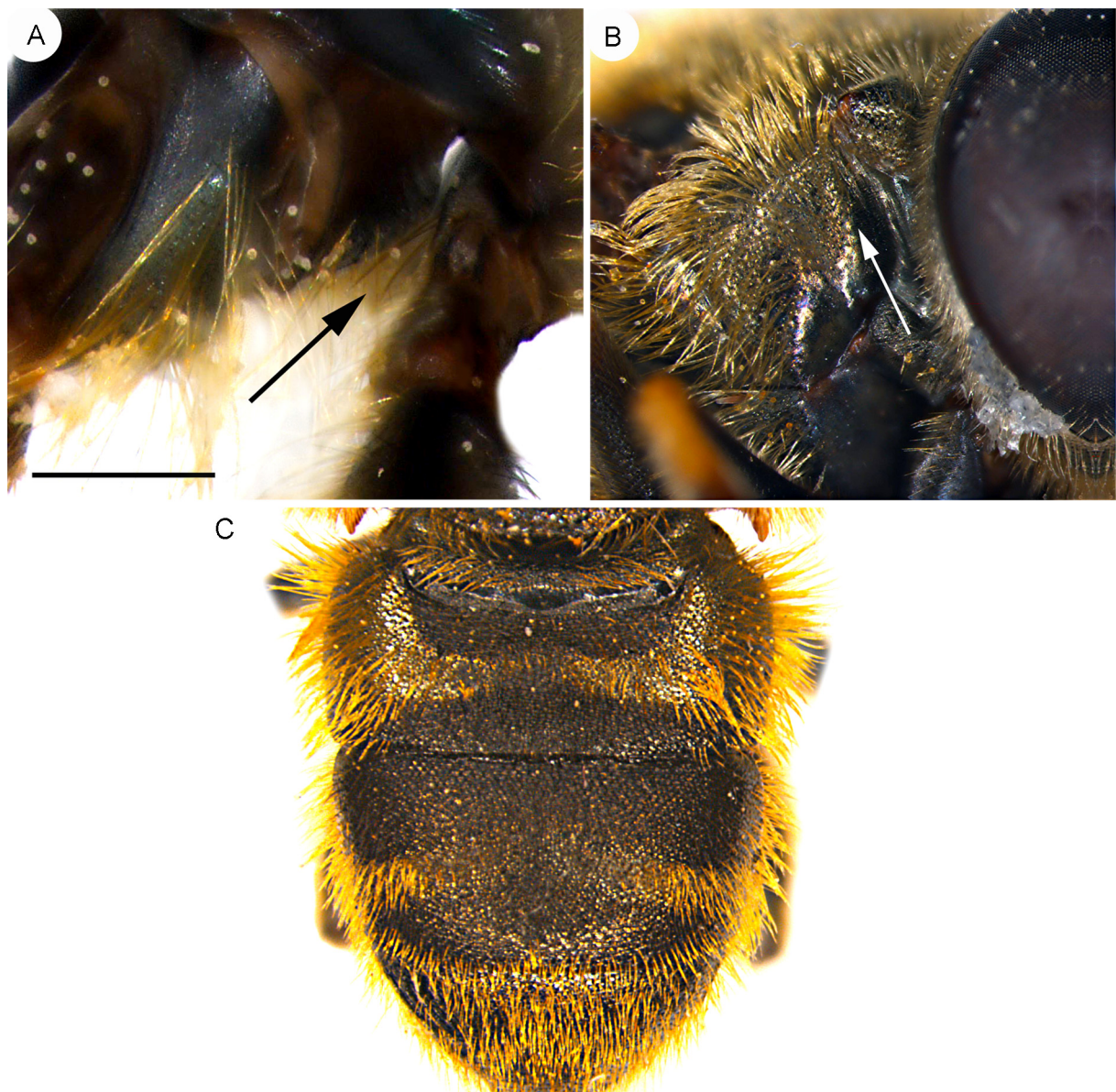


Fig. 2. A–C. *Merodon funestus* (Fabricius, 1794), ♂. **A.** Mid coxa (FSUNS; FSUNS ID F98), lateral view, posterior pile marked with arrow. **B.** Anterior anepisternum (CWM; FSUNS ID 10347), pile below postpronotum marked with arrow, anterolateral view. **C.** Abdomen, dorsal view (FSUNS; FSUNS ID F99). Scale bar = 2 mm.

V.W. Fowler leg.; BMNH • 1 ♀; Transkei, Pitseng Pass, banks from Luzzi River; 30.749° S, 28.299° E; 1633 m a.s.l.; 11 Jan. 1979; J. Londt and B. Stuckenberg leg.; rocky hill and grassveld; FSUNS ID 04278; NMSA • 1 ♂; Eastern Cape, 5 km ENE of Rhodes; 30.783° S, 28.000° E; 2066 m a.s.l.; 5 Feb. 1992; wet stream gully; unknown leg.; FSUNS ID 04275; NMSA • 1 ♀; Eastern Cape, Naudes Nek, near top of pass; 30.759° S, 28.137° E; 2500 m a.s.l.; 6 Feb. 2009; S.I. Morita leg.; FSUNS ID 05724; USNM SMPC_SIM_1844 • 1 ♂; Drakensberg Mountain, Maclear-Naudes Nek; 30.759° S, 28.203° E; 1800 m a.s.l.; 9 Feb. 2016; S. Radenković and N. Veličković leg.; FSUNS ID ZA2_079; FSUNS.

LESOTHO • 1 ♀; Colony of Basutoland, Orange Valley; 29.15° S, 29.03° E; 1500 m a.s.l.; R. Ellenberger leg.; FSUNS ID 03317; MNHN PM-05-46.

Redescription

Original description is insufficient, without any illustrations and based on a single male specimen from South Africa (Hull 1944).

Length: large species, body 10–14 mm, wing 8–10 mm (n = 8).

Male

HEAD (Figs 4A, 9A, 11A, 12A). Antenna (Fig. 4A) very short, reddish to dark brown; scape and pedicel covered with dense, yellow pilosity; pedicel elongated, approximately as long as basoflagellomere (relation scape:pedicel:basoflagellomere=1.0:1.8:1.8); basoflagellomere concave dorsally, with

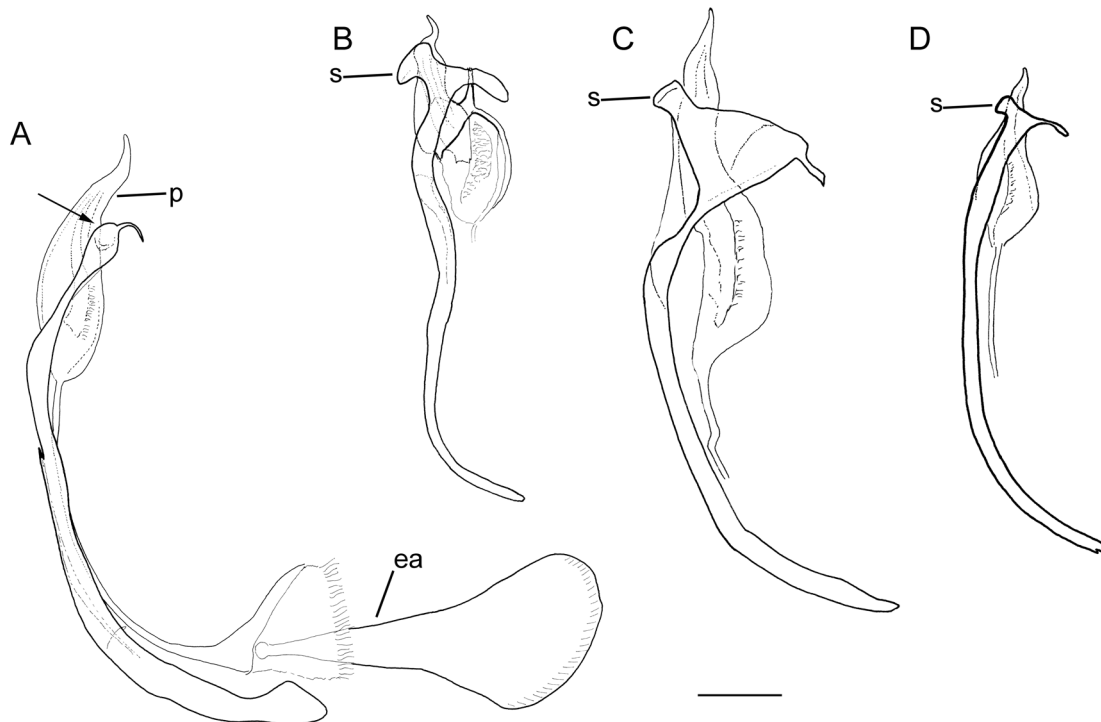


Fig. 3. Male genitalia, aedeagus, lateral view. **A.** *Merodon aureus* (Fabricius, 1805) (FSUNS; FSUNS ID 09662) the place of absence of lateral sclerite of the aedeagus marked with arrow. **B.** *M. funestus* (Fabricius, 1794) (FSUNS; FSUNS ID F99). **C.** *M. bombiformis* Hull, 1944 (FSUNS; FSUNS ID A2_079). **D.** *M. zebra* Vujić & Radenković sp. nov., paratype (BMNH; FSUNS ID 03314). Abbreviations: ea = ejaculatory apodeme; p = aedeagus; s = lateral sclerite of aedeagus. Scale bar = 0.25 mm.



Fig. 4. Male antenna, lateral view. **A.** *Merodon bombiformis* Hull, 1944 (FSUNS; FSUNS ID ZA2_079). **B.** *M. multifasciatus* Curran, 1939 (USNM; FSUNS ID 04512). **C.** *M. vittatus* Vujić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **D.** *M. zebra* Vujić & Radenković sp. nov. paratype (NBCN; FSUNS ID 05174). **E.** *M. lotus* Vujić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_066). **F.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04972). Scale bar = 2 mm.

rounded apex; arista light brown to dark brown, thickened basally, 5 times as long as basoflagellomere. Face reddish-yellow to dark brown, covered with yellow pollinosity and dense yellow pile, except on bare medial vitta that occupies $\frac{1}{4}$ width of face (Fig. 11A). Oral margin reduced (Fig. 9A). Frons from black to reddish and golden-yellow, inflated, covered with golden-yellow pollinosity and pile. Vertical triangle isosceles (Fig. 12A), usually reddish to black, predominantly covered with a long, yellow pile. Ocellar triangle equilateral. Eye pile dense, long as the scape, often pale yellow to gray. Eye contiguity about 10–15 ommatidia long. Occiput reddish, covered with yellow pile.

THORAX (Figs 6A, 10A). Mesonotum matte black, except lateral side of scutum including postpronotum and postalar callus, and posterior margin of scutellum reddish-yellow, covered with reddish pile; scutum with five pollinose longitudinal vittae (Fig. 10A). Pleuron covered with gray pollinosity and the following parts with whitish pile: anterior part of proepimeron, posterior part of anterior anepisternum, most of the posterior anepisternum except anterior end, antero-ventral and postero-dorsal part of katapisternum, anepimeron, and metasternum; in some specimens anterior anepisternum and katatergum reddish. Wing hyaline, with dense microtrichia and yellow to brown veins. Calypter pale yellow. Halter with yellow pedicel and capitulum. Legs usually dark brown to black (in some specimens partly reddish), except yellow basal third of tibiae. Metatrochanter without calcar. Metafemur moderately thickened, with less developed apicoventral triangular lamina, not dentate, only the apical dens is distinct, posteriorly oriented (Fig. 6A). Pile on legs predominantly yellow.

ABDOMEN (Fig. 5A). Predominantly reddish-yellow (Fig. 5A). Reddish parts of terga 2–4 covered with yellow pile and dense pollinosity, while dark parts with short adpressed black pile; sterna from yellow to dark brown, covered with white or yellow pile.

GENITALIA (Figs 3C, 13A–C). Posterior lobe of surstyle broad, rounded apically (Fig. 13A: pl); anterior lobe of surstyle undeveloped, virtually absent (Fig. 13A); cercus elongated (Fig. 13A: c). Hypandrium with theca slightly medially narrowed (Fig. 13C: marked with arrow). Lateral sclerite of aedeagus small, quadrilateral (Fig. 3C: s).

Female (Figs 14A, 15A, 16A)

Similar to the male except for normal sexual dimorphism: frons reddish-yellow, with broad goldish pollinose vittae along eye margins; frons completely covered with golden pile (Fig. 15A); metafemur with very small apicoventral triangular lamina; apical dens more or less distinct, slightly posteriorly oriented (Fig. 16A).

Period of flight and distribution (Fig. 1)

The species is distributed in the Republic of South Africa and Lesotho. On the basis of our data, the flight period was found to be from November to February. *Merodon bombiformis* was collected in different vegetation types, brush-grass savanna, temperate and mountain grasslands and montane forest-tundra. On Drakensberg Mountain, at the locality between Maclear and Naudes Nek, it was found near the stream, within bushes of *Leucosidea sericea* Eckl. & Zeyh. (Rosaceae) and *Gladiolus* sp. on rocky soil (Fig. 24). *Leucosidea sericea* grows in Afromontane regions of southern Africa, and is the sole species of this genus (South African National Biodiversity Institute 2004).

Larval biology

Stuckenberg (1956) described the puparia of *M. bombiformis*, reared from larvae found feeding on commercially grown *Gladiolus* sp. (Iridaceae) corms. Specimens from Pietermaritzburg (5 Nov. 1955, 19 Nov. 1955) were reared from bulbs of *Gladiolus* L. too.



Fig. 5. Male abdomen, dorsal view. **A.** *Merodon bombiformis* Hull, 1944 (FSUNS; FSUNS ZA2_079). **B.** *M. multifasciatus* Curran, 1939 (USNM 2052372; FSUNS ID 04512). **C.** *M. vittatus* Vujić & Likov sp. nov. holotype (NBCN; FSUNS ID 04085). **D.** *M. zebra* Vujić & Radenković sp. nov. holotype (RMCA; FSUNS ID 25089). **E.** *M. lotus* Vujić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_066). **F.** *M. nasicus* Bezzi, 1915 (RBINS; FSUNS ID 21976). Scale bar = 2 mm.

***Merodon lotus* Vujić & Radenković sp. nov.**

urn:lsid:zoobank.org:act:4FA6E871-8E78-418E-89EA-B4A334742811

Figs 1, 4E, 5E, 6E, 9E, 10E, 11E, 12E, 14D, 15D, 16D, 17, 18A, 22A–C, 26

Diagnosis

Black, medium sized species (8–9 mm), with shiny face, posterior half of scutum and terga without pollinosity (except a pair of indistinct pollinose fasciae on tergum 4 in female) (Fig. 14D); male genitalia with S-forming posterior lobe of surstyle, gradually narrowing toward tip (Fig. 17A, D: pl), anterior margin of surstyle rounded in lateral view (Fig. 17A, D: ams). Similar to *Merodon nasicus* from which differs in less prominent oral margin (Fig. 11E), and terga without clear pollinosity (Figs 5E, 14D), while in *M. nasicus* oral margin distinctly protruded (Fig. 11F) and terga 2–4 with distinct pollinose fasciae (Figs 5F, 14E, 20D).

Etymology

The specific epithet ‘*lotus*’ is a Latin adjective meaning ‘elegant, fine’, also ‘washed/clean’, implying clean, non pollinose terga, as the main diagnostic character of this species.

Material examined

Holotype

REPUBLIC OF SOUTH AFRICA • 1 ♂; Ceres, Ceres 3; 33.364° S, 19.289° E; 546 m a.s.l.; 10 Nov. 2018; A. Vujić and S. Radenković S. leg.; FSUNS ID ZA6_039; NMSA.

Paratypes

REPUBLIC OF SOUTH AFRICA • 1 ♂; Ceres, Ceres 2; 33.365° S, 19.289° E; 547 m a.s.l.; 9 Nov. 2018; A. Vujić and S. Radenković Sanja leg.; FSUNS ZA6_021; FSUNS • 4 ♂♂, 1 ♀; same collection data as for holotype; FSUNS ZA6_048, ZA6_049, ZA6_052, ZA6_060, ZA6_066; FSUNS • 1 ♀; same collection data as for holotype; FSUNS ID ZA6_067; NMSA.

Additional material

REPUBLIC OF SOUTH AFRICA • 1 ♂; Cape Province, Ceres; 1500 ft; Dec. 1920; R.E. Turner leg.; *Merodon appendiculatus* Hull unpublished name; BMNH.

Description

Length: medium sized species, body 8–9 mm, wing 5–6 mm (n = 8).

Male

HAD (Figs 4E, 9E, 11E, 12E). Antenna (Fig. 4E) brown; pedicel slightly shorter than basoflagellomere (relation scape:pedicel:basoflagellomere=1.0:2.1:2.5); basoflagellomere concave dorsally, with acute apex; arista thickened basally, yellowish at basal third, the rest blackish, 2.2 times as long as basoflagellomere. Face shiny black, covered with whitish or yellowish pile, except on bare medial vitta that occupies half width of face. Oral margin slightly protruded (Fig. 9E). Frons shiny black, inflated, covered with whitish-gray pile and gray pollinosity along eye margins. Vertical triangle isosceles (Fig. 12E), dark, predominantly covered with long, yellowish pile, except some black ones on ocellar triangle in some specimens. Ocellar triangle usually equilateral. Eye pile gray, except upper fourth with black pile, slightly longer than scape. Eye contiguity about 14–16 ommatidia long. Occiput blackish, mostly pollinose, except behind vertical triangle, covered with yellow pile.

THORAX (Figs 6E, 10E). Mesonotum black, covered with reddish-yellow pile; scutum without pollinose longitudinal vittae, but postpronotum and anterior part of scutum pollinose (Fig. 10E). Pleuron black, covered with sparse gray pollinosity and the following parts with whitish to yellow pile: anterior part



Fig. 6. Male metaleg, lateral view. **A.** *Merodon bombiformis* Hull, 1944 (FSUNS; FSUNS ZA2_079). **B.** *M. multifasciatus* Curran, 1939 (USNM 2052372; FSUNS ID 04512). **C.** *M. vittatus* Vujčić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **D.** *M. zebra* Vujčić & Radenković sp. nov., paratype (NBCN; FSUNS ID 05174). **E.** *M. lotus* Vujčić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_066). **F.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04972). Scale bar = 2 mm.

of proepimeron, posterior part of anterior anepisternum, most of the posterior anepisternum except anterior end, antero-ventral and postero-dorsal part of katepisternum, anepimeron, metasternum. Wing brownish, hyaline, with dense microtrichia and dark brown veins. Calypter yellowish. Halter with yellowish pedicel and capitulum. All three femora dark brown to black; tibiae mostly reddish, medially black to dark brown; tarsi reddish, except blackish apical segment (Fig. 6E). Metatrochanter without calcar. Metafemur moderately thickened, with less developed apicoventral triangular lamina, more or less denatate, the apical dens more or less distinct (Fig. 6E). Pile on legs predominantly yellowish.

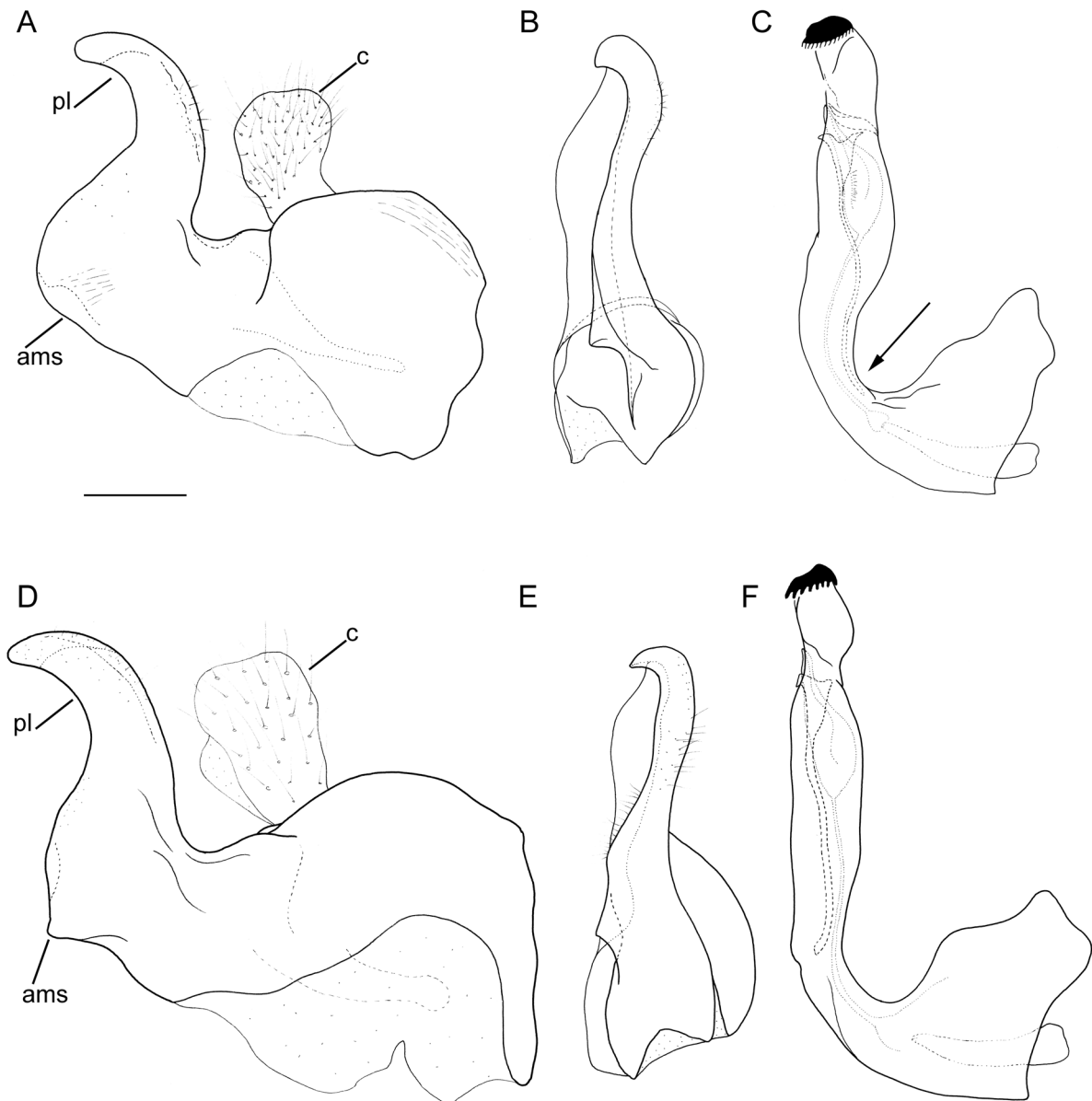


Fig. 7. Male genitalia. **A–C.** *Merodon zebra* Vujić & Radenković sp. nov., paratype (NBCN; FSUNS ID 05174). **D–F.** *M. vittatus* Vujić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **A, D.** Epandrium, lateral view. **B, E.** Epandrium, ventral view. **C, F.** Hypandrium, lateral view. Abbreviations: ams = anterior margin of surstyle; c = cercus; pl = posterior lobe of surstyle. Scale bar = 0.25 mm.

ABDOMEN (Fig. 5E). Black, covered with yellow to whitish pile on lateral sides. Tergum 1 covered with short whitish to yellow pile; terga 2 and 3 medially with shorter black pile, except two medial fasciae with whitish pile and some whitish pile at posterior margin; anterior half of tergum 4 covered with short black pile, posterior half with longer yellowish pilosity, medially with two oblique fasciae covered with whitish pile. Sterna dark brown, covered with yellowish pile.

GENITALIA (Figs 17, 18A). Posterior lobe of surstyle broad, bent, S-shaped, gradually narrowing toward tip (Fig. 17A, D: pl); anterior lobe of surstyle undeveloped (Fig. 17A, D); anterior margin of surstyle rounded in lateral view (Fig. 17A, D: ams); cercus square like (Fig. 17A, D: c). Hypandrium with theca medially distinctly narrowed (Fig. 17C, F). Lateral sclerite of aedeagus small (Fig. 18A: s).

Female (Figs 14D, 15D, 16D)

Similar to the male except for normal sexual dimorphism: Frons shiny, non pollinose, except along eye margins (Fig. 15D); tergum 4 with a pair of indistinct narrow oblique pollinose fasciae.

Period of flight and distribution (Fig. 1)

Merodon lotus sp. nov. occurs in RSA in the Ceres Mountain Fynbos Nature Reserve; the flight period is in November and December. This species appears in Mediterranean evergreen forest-hard-leaf scrub in the south of the African continent (Sayre *et al.* 2013).

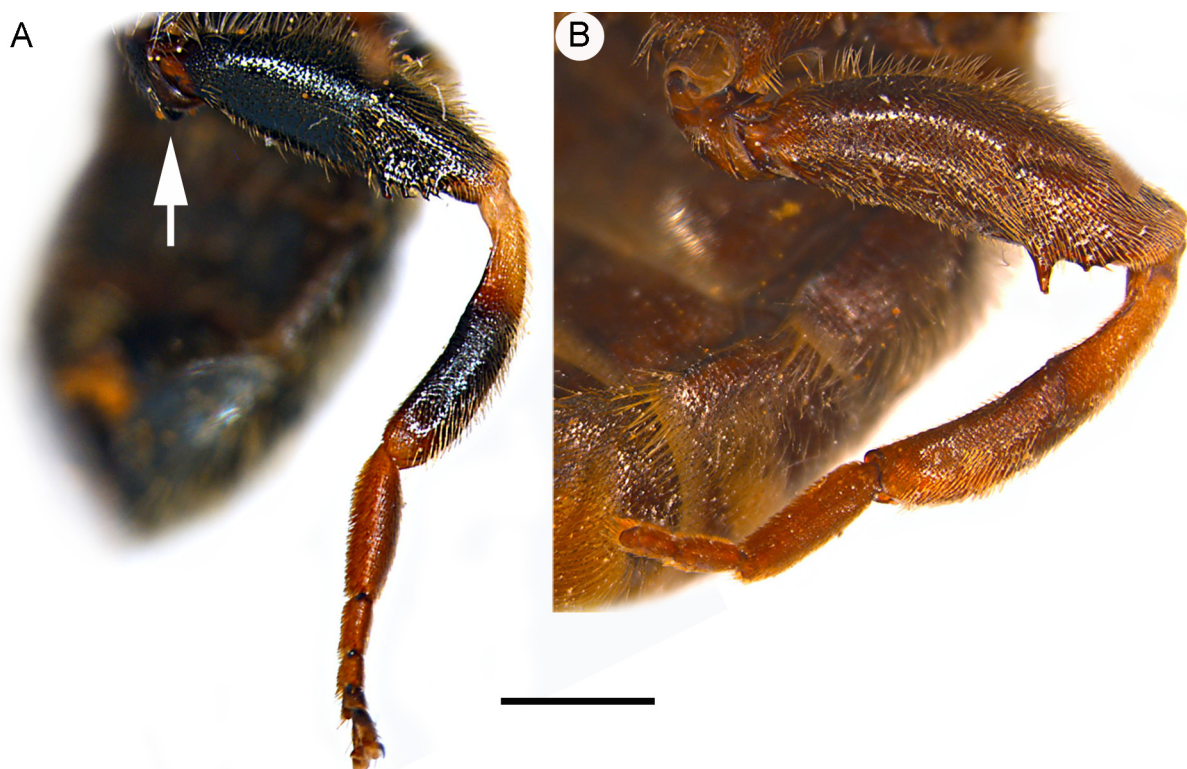


Fig. 8. Male metaleg, lateral view. **A.** *Merodon funestus* (Fabricius, 1794) (CWM; FSUNS 10347). **B.** *M. vittatus* Vujić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). Calcar on metatrochanter marked with arrow. Scale bar = 2 mm.

Flower visited

During our fieldwork, adults of *M. lotus* sp. nov. were observed as regular visitors of flowers of the genus *Coleonema* Bartl. & H.L.Wendl. (Rutaceae) in Ceres (Fig. 25).

Merodon multifasciatus Curran, 1939

Figs 1, 4B, 5B, 6B, 9B, 10B, 11B, 12B, 13D–F, 14B, 15B, 16B, 18B, 21F–H, 25

Merodon multifasciatus Curran, 1939: 8 (type locality: Johannesburg, Republic of South Africa).

Merodon apimima Hull, 1944: 40–42 (type locality: Mulanje, Nyasaland, Republic of Malawi).

Diagnosis

Large (10–12 mm) and dark, striped species, with golden-yellow pile on the tip of the abdomen (Figs 5B, 14B). Scutum with conspicuous pollinose vittae (Figs 10B, 14B); metafemur with distinct apical dens on triangular lamina (Figs 6B, 16B); terga 3 and 4 with broad medial pollinose fasciae (Figs 5B, 14B). Similar to *M. vittatus* Vujić & Likov sp. nov. from which differs in male genitalia with wider posterior lobe of surstyle, slightly curved, almost parallel sided, with rounded apex (Fig. 13D: pl), while male genitalia in *M. vittatus* sp. nov. with longer and narrower, S-shaped posterior lobe of surstyle, gradually narrowing toward tip (Fig. 7D: pl).

Material examined

Holotype

REPUBLIC OF SOUTH AFRICA • 1 ♀; Johannesburg; 26.284° S, 27.982° E; 6000 ft; Mar. 1899; J.P. Cregoe leg.; AMNH.

Additional material

REPUBLIC OF MALAWI • 1 ♂, holotype of *Merodon apimima*; Mulanje, Nyasaland; 16.015° S, 35.521° E; 22 Apr. 1913; S.A. Neave leg.; BMNH • 1 ♀, paratype of *Merodon apimima*; same collection data as for preceding; 21 Oct. 1913; labelled as allotype; BMNH • 2 ♂♂, paratypes of *Merodon apimima*; same collection data as for preceding; 18–30 Apr. 1913; BMNH • 1 ♂, paratype of *Merodon apimima*; same collection data as for preceding; FSUNS ID 04086; NBCN • 2 ♂♂, paratypes of *Merodon apimima*; same collection data as for preceding; 2 Jun. 1913; BMNH • 1 ♀, paratype of *Merodon apimima*; same collection data as for preceding; FSUNS ID 04355; BMNH.

Additional non type material

DEMOCRATIC REPUBLIC OF THE CONGO • 1 ♂; Lualaba River; 7.023° S, 27.046° E; 2500–4000 ft; 21 Apr. 1907; S.A. Neave leg.; FSUNS ID 03909; NBCN LML-05-3.

REPUBLIC OF SOUTH AFRICA • 1 ♂; Modimolle (former Nylstroom); 24.683° S, 28.416° E; 17 Feb. 1908; unknown leg.; reared from *Gladiolus* bulb; USNM • 1 ♂; Transvaal, 5 mi (8 km) W of Warmbad; 24–25 Feb. 1968; K.V. Krombein leg.; FSUNS ID 04512; USNM ENT 00036564, USNM 2052372 • 1 ♂; Transvaal, Fortuna Trail, S of Barberton, N slope; 26.009° S, 31.048° E; 8 Apr. 1985; J.G.H. Londt leg.; bushveld; FSUNS ID 04284; NMSA • 1 ♀; Mpumalanga, Emgwenya (Waterval Boven), near Elandsrivier; 25.634° S, 30.326° E; 6 Apr. 2018; Sanja Vujić leg.; FSUNS ZA5_242; FSUNS • 1 ♀; same collection data as for preceding; L.S. Forcada leg.; FSUNS ZA5_243; FSUNS.

Redescription

Original descriptions are insufficient, without any illustrations and based on a single female from South Africa (Curran 1939) and male and female types (Hull 1944).

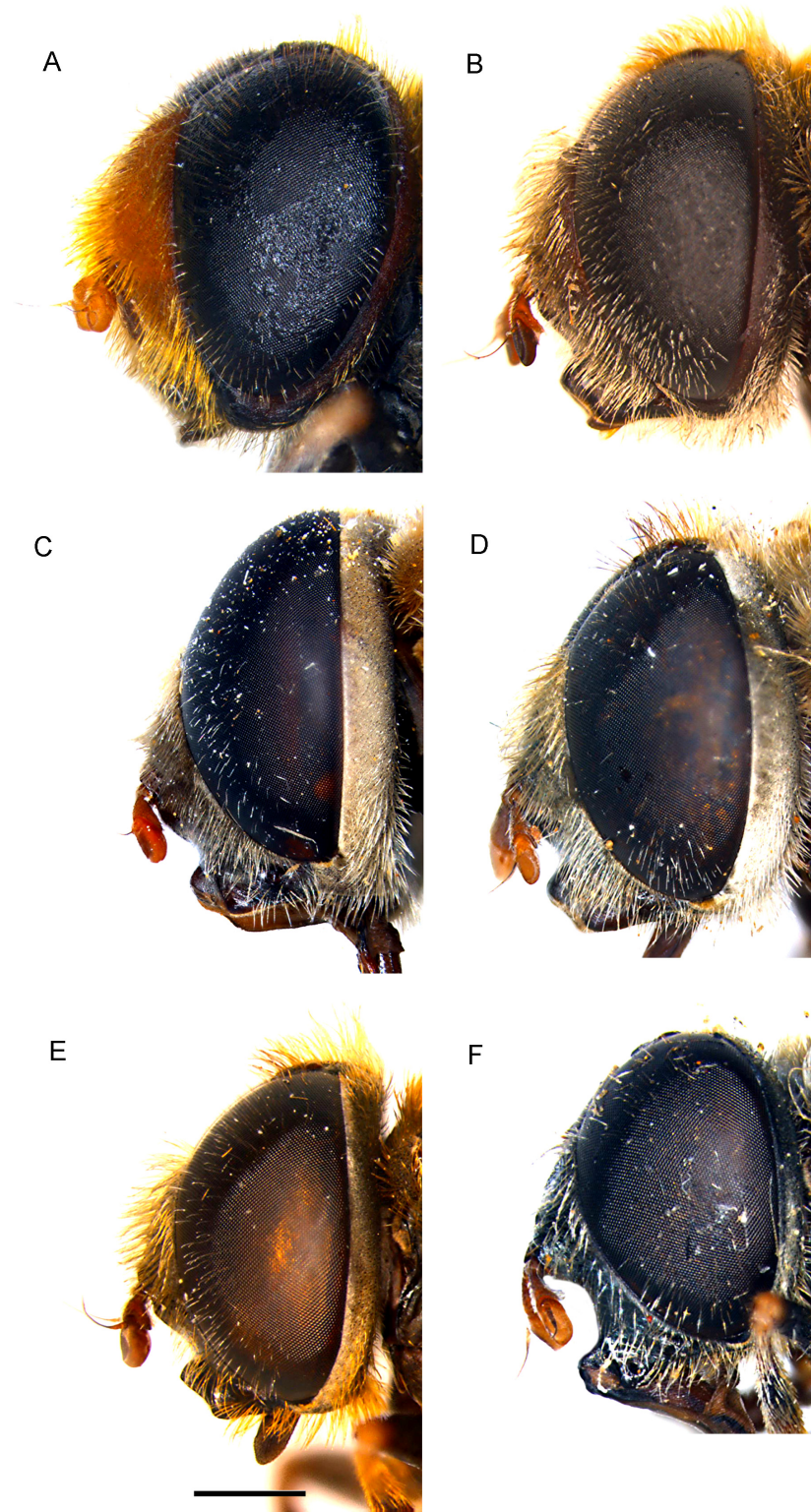


Fig. 9. Male head, lateral view. **A.** *Merodon bombiformis* Hull, 1944 (FSUNS; FSUNS ZA2_079). **B.** *M. multifasciatus* Curran, 1939 (USNM 2052372; FSUNS ID 04512). **C.** *M. vittatus* Vujčić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **D.** *M. zebra* Vujčić & Radenković sp. nov., holotype (RMCA; FSUNS ID 25089). **E.** *M. lotus* Vujčić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_066). **F.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04972). Scale bar = 2 mm.

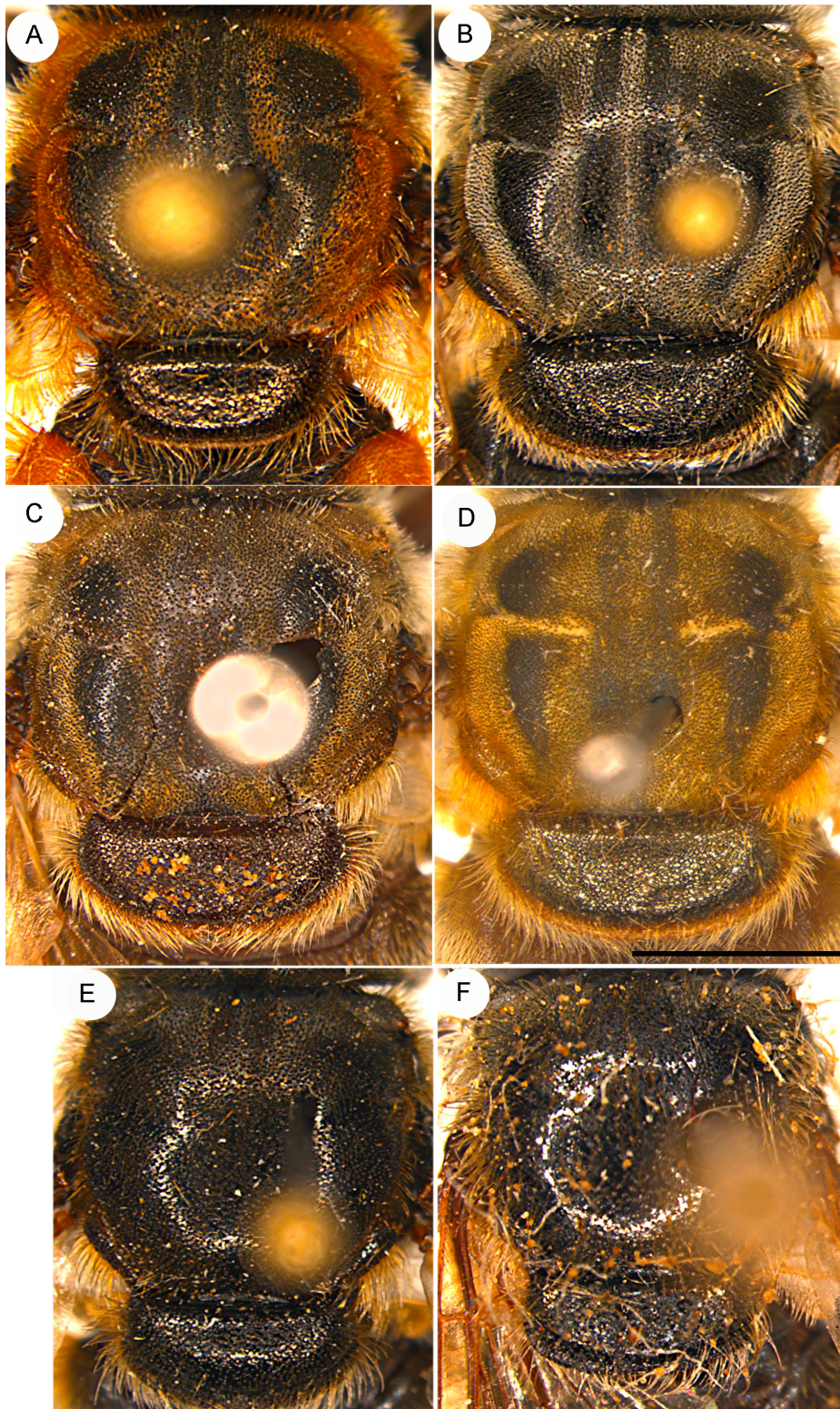


Fig. 10. Mesonotum, dorsal view. **A.** *Merodon bombiformis* Hull, 1944 (FSUNS; FSUNS ZA2_079). **B.** *M. multifasciatus* Curran, 1939 (FSUNS; FSUNS ID ZA5_243). **C.** *M. vittatus* Vujić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **D.** *M. zebra* Vujić & Radenković sp. nov., paratype (RMCA; FSUNS ID 25090). **E.** *M. lotus* Vujić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_066). **F.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04972). **A, C, E-F.** Male. **B, D.** Female. Scale bar = 2 mm.

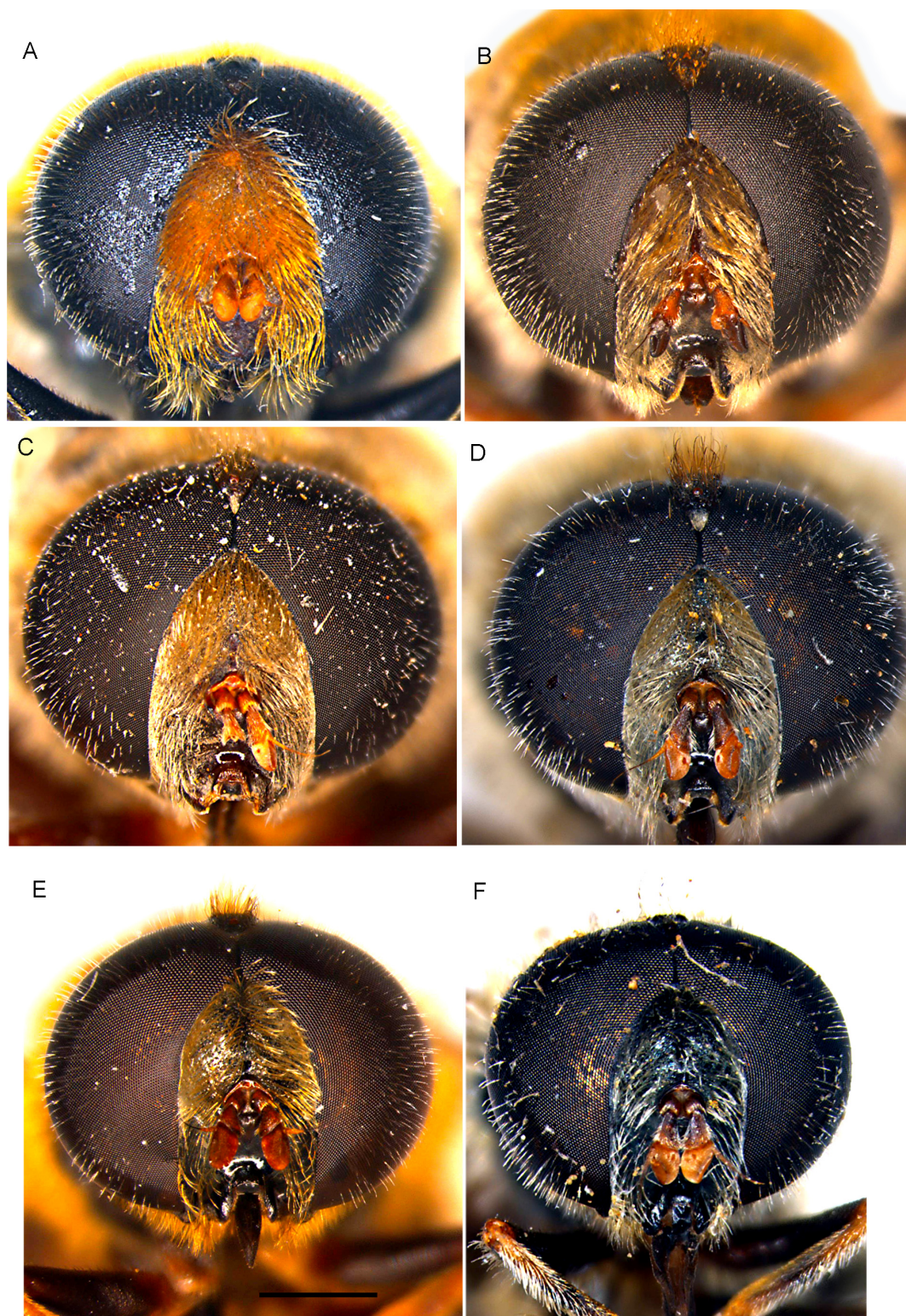


Fig. 11. Male head, frontal view. **A.** *Merodon bombiformis* Hull, 1944 (FSUNS; FSUNS ZA2_079). **B.** *M. multifasciatus* Curran, 1939 (USNM 2052372; FSUNS ID 04512). **C.** *M. vittatus* Vujčić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **D.** *M. zebra* Vujčić & Radenković sp. nov., holotype (RMCA; FSUNS ID 25089). **E.** *M. lotus* Vujčić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_066). **F.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04972). Scale bar = 2 mm.

Length: large species, body 10–12 mm, wing 8–9 mm (n = 5).

Male

HEAD (Figs 4B, 9B, 11B, 12B). Antenna (Fig. 4B) reddish-brown; pedicel elongated, approximately as long as basoflagellomere (relation scape : pedicel : basoflagellomere = 1.0 : 2.5 : 2.5); basoflagellomere concave dorsally, with acute apex; arista light brown to dark brown, thickened basally, 1.6 times as long as basoflagellomere. Face from black to dark brown, covered with gray pollinosity and whitish pile, except on bare medial vitta that occupies $\frac{1}{4}$ width of face. Oral margin protruded (Fig. 9B). Frons from black to reddish, inflated, covered with gray pollinosity and whitish pile. Vertical triangle and isosceles (Fig. 12B), usually dark brown, predominantly covered with a long, yellow pile. Ocellar triangle equilateral. Eye pile dense, gray, slightly longer than scape. Eye contiguity about 15–18 ommatidia long. Occiput reddish, pollinose, covered with yellow pile.

THORAX (Figs 6B, 10B). Mesonotum black, except postpronotum and posterior margin of scutellum brown-reddish, covered with reddish-yellow pile; scutum with five pollinose longitudinal vittae (as on Fig. 10B). Pleuron black to dark brown, covered with gray pollinosity and the following parts with whitish to yellow pile: anterior part of proepimeron, posterior part of anterior anepisternum, most of the posterior anepisternum except anterior end, antero-ventral and postero-dorsal part of katepisternum, anepimeron, and metasternum. Wing hyaline, with dense microtrichia and yellow to brown veins. Calypter pale yellow. Halter with yellow pedicel and capitulum. All three femora dark brown to black; tibiae mostly reddish, medially black to dark brown; tarsi reddish (Fig. 6B). Metatrochanter without calcar. Metafemur moderately thickened, with less developed apicoventral triangular lamina, more or less dentate, the apical dens is distinct (Fig. 6B). Pile on legs predominantly whitish to yellow.

ABDOMEN (Fig. 5B). Black to dark brown. Tergum 1 black, usually pollinose, covered with short whitish to yellow pile; tergum 2 with pollinose posterior margin and narrow medial fascia (in some specimens interrupted in the middle) covered with whitish pile except posterior half of tergum (between pollinose fasciae) covered with short black pilosity; terga 3 and 4 with pollinose posterior margin and broad medial pollinose fasciae (Fig. 5B); anterior half of tergum 3 covered with short black pile; tergum 4 covered with long golden-yellow pilosity (Fig. 5B). Sterna reddish-brown, covered with white or yellow pile.

GENITALIA (Figs 13D–F, 18B). Posterior lobe of surstyle broad, slightly curved, almost parallel sided, with rounded apex (Fig. 13D: pl); anterior lobe of surstyle undeveloped (Fig. 13D); anterior margin of surstyle angular in lateral view (Fig. 13D: ams); cercus square like (Fig. 13D: c). Hypandrium with theca medially distinctly narrowed (Fig. 13F). Lateral sclerite of aedeagus small (Fig. 18B: s).

Female (Figs 10B, 14B, 15B, 16B)

Similar to the male except for normal sexual dimorphism: metafemur usually with small apical dens on triangular lamina (Fig. 16B); frons with non pollinose medial vitta (Fig. 15B); vertex predominantly covered with black pile.

Remarks

Hull (1944) described *Merodon apimima* “related to *multifasciatus* Curran in type of femoral armament and abdominal fascia; distinct in the black front, yellow pilose vertex and, brown humeri, absence of red on second abdominal segment, beside other details of pattern” based on six males and one female. All these characters mentioned in original description can be regarded as intraspecific variability of *M. multifasciatus*. Based on our study of the type material of both species, we regard *M. apimima* as a junior synonym of *M. multifasciatus*.

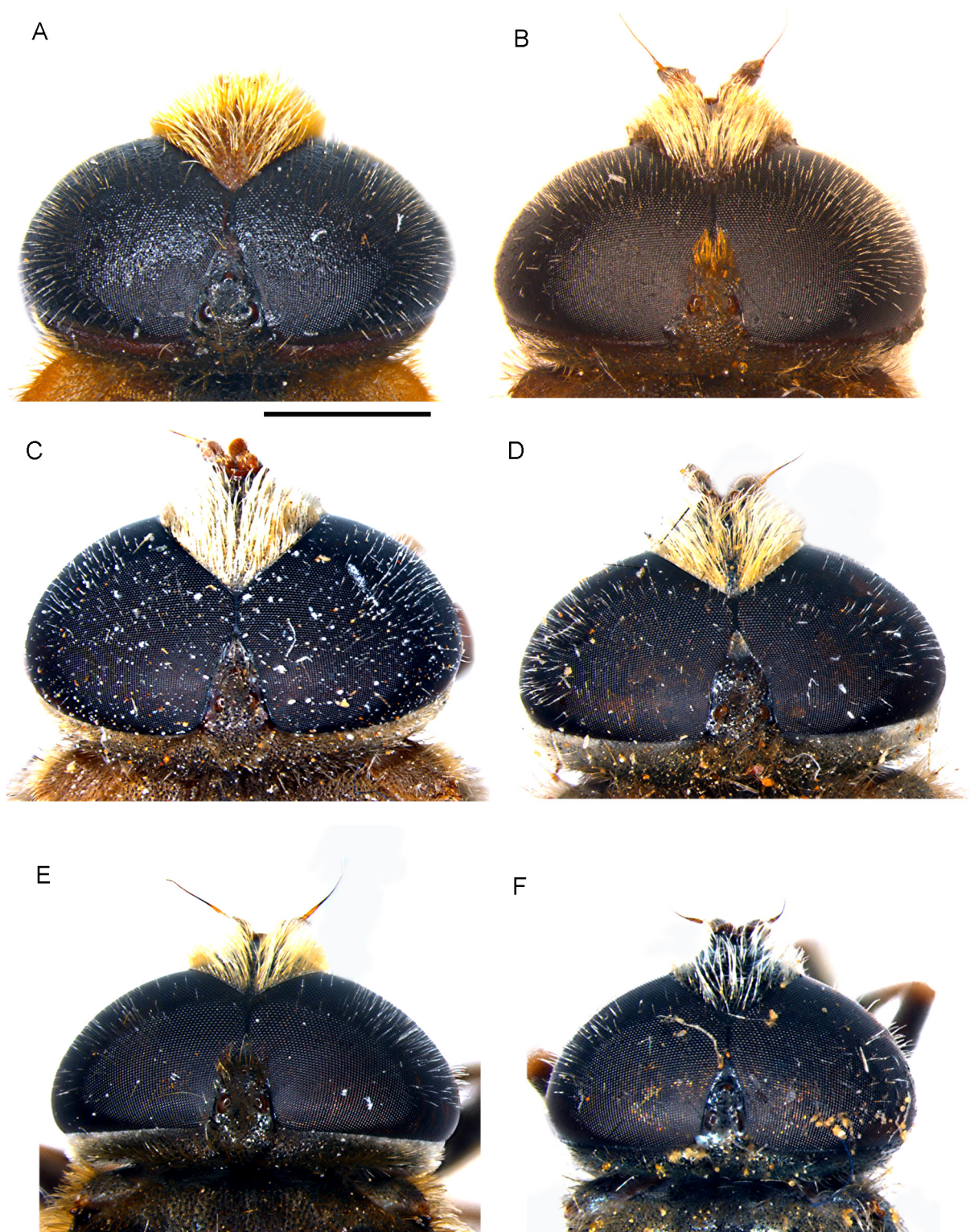


Fig. 12. Male head, dorsal view. **A.** *Merodon bombiformis* Hull, 1944 (FSUNS; FSUNS ZA2_079). **B.** *M. multifasciatus* Curran, 1939 (USNM 2052372; FSUNS ID 04512). **C.** *M. vittatus* Vujčić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **D.** *M. zebra* Vujčić & Radenković sp. nov., holotype (RMCA; FSUNS ID 25089). **E.** *M. lotus* Vujčić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_066). **F.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04972). Scale bar = 2 mm.

In the original description of species, Hull (1944) did not state the etymology of the species name, so we cannot conclude whether he treated “*apimima*” as a noun or an adjective. According to the International Code of Zoological Nomenclature (Article 31.2.2) (International Commission on Zoological Nomenclature 1999) if the author does not indicate whether the species name is a noun or an adjective, as in this case, it has to be treated as a noun. When the species name is a noun simple or compound, in apposition, it does not need to agree in gender with the generic name, therefore the original spelling has to be retained (Article 34.2.1). In some references (Smith & Vockeroth 1980; Dirickx 1988) *M. apimimus* is used because authors considered “*apimima*” to be an adjective. But in Systema Diptorum (Evenhuis & Pape 2020), which is the most comprehensive database related to the Syrphidae, the original spelling of *M. apimima* is applied.

Period of flight and distribution (Fig. 1)

This species occurs in Malawi, the Republic of South Africa and the Democratic Republic of Congo. On the basis of our data, the flight period was found to be throughout the whole year. *Merodon multifasciatus* was found in deciduous forest-woodland savanna and brush-grass savanna.

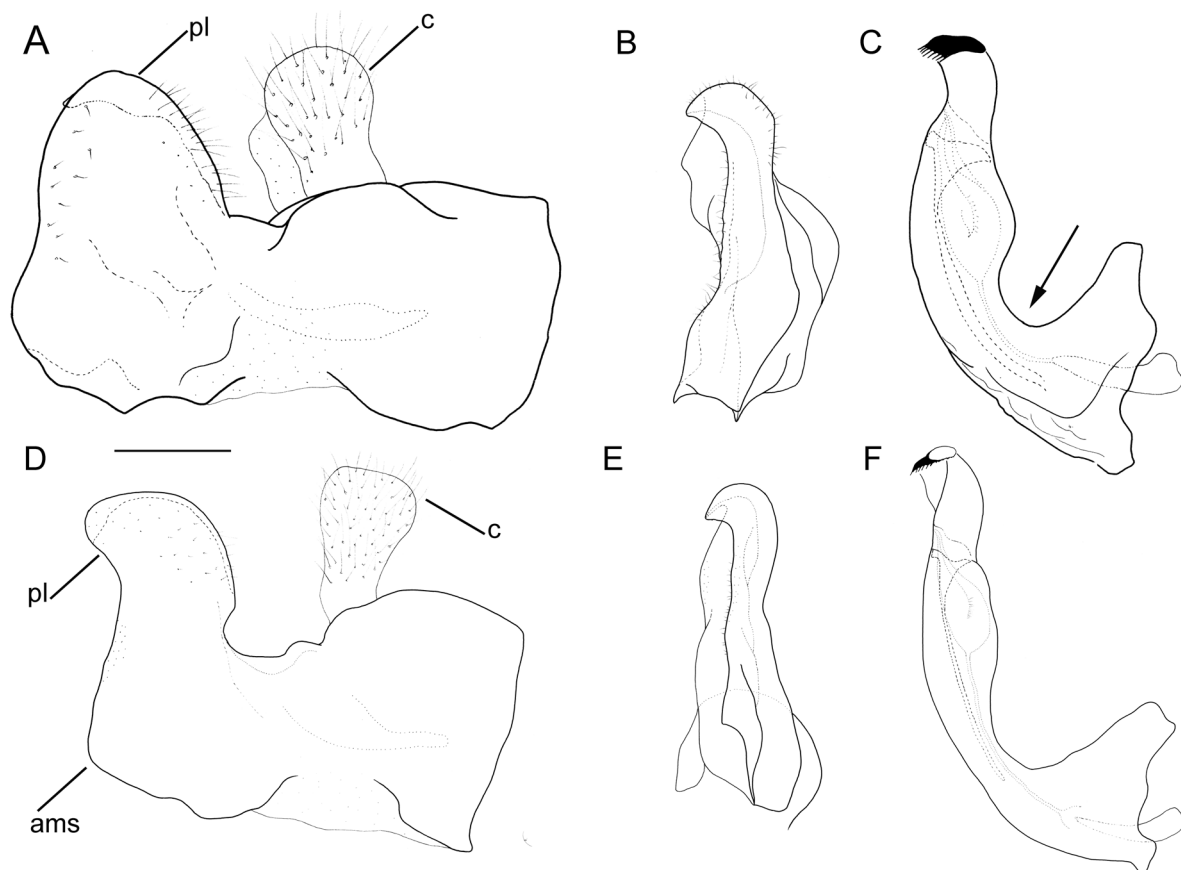


Fig. 13. Male genitalia. **A–C.** *Merodon bombiformis* Hull, 1944 (NMSA; FSUNS ID 04277). **D–F.** *M. multifasciatus* Curran, 1939 (NMSA; FSUNS ID 04284). **A, D.** Epandrium, lateral view. **B, E.** Epandrium, ventral view. **C, F.** Hypandrium, lateral view. Abbreviations: ams = anterior margin of surstyle; c = cercus; pl = posterior lobe of surstyle. Medially narrowed hypandrium marked with arrow. Scale bar = 0.5 mm.

Larval biology

The material from Nylstroom (17 Feb. 1908) was bred from bulbs of *Gladiolus* (immature stages undescribed). Specimens collected in Emgwenya (Waternal Boven) (6 Apr. 2018) were found on flowers of different plant species inside a small spot (20 × 40 m) with the natural population of *Gladiolus sericeovillosus* Hook., along a small periodical spring. It is a strong indication that this plant is the primary host for larvae of *M. multifasciatus*.

Flower visited

During our fieldwork, adults of *M. multifasciatus* were observed visiting flowers of *Nidorella auriculata* DC. in Emgwenya (Waternal Boven) (Fig. 26).

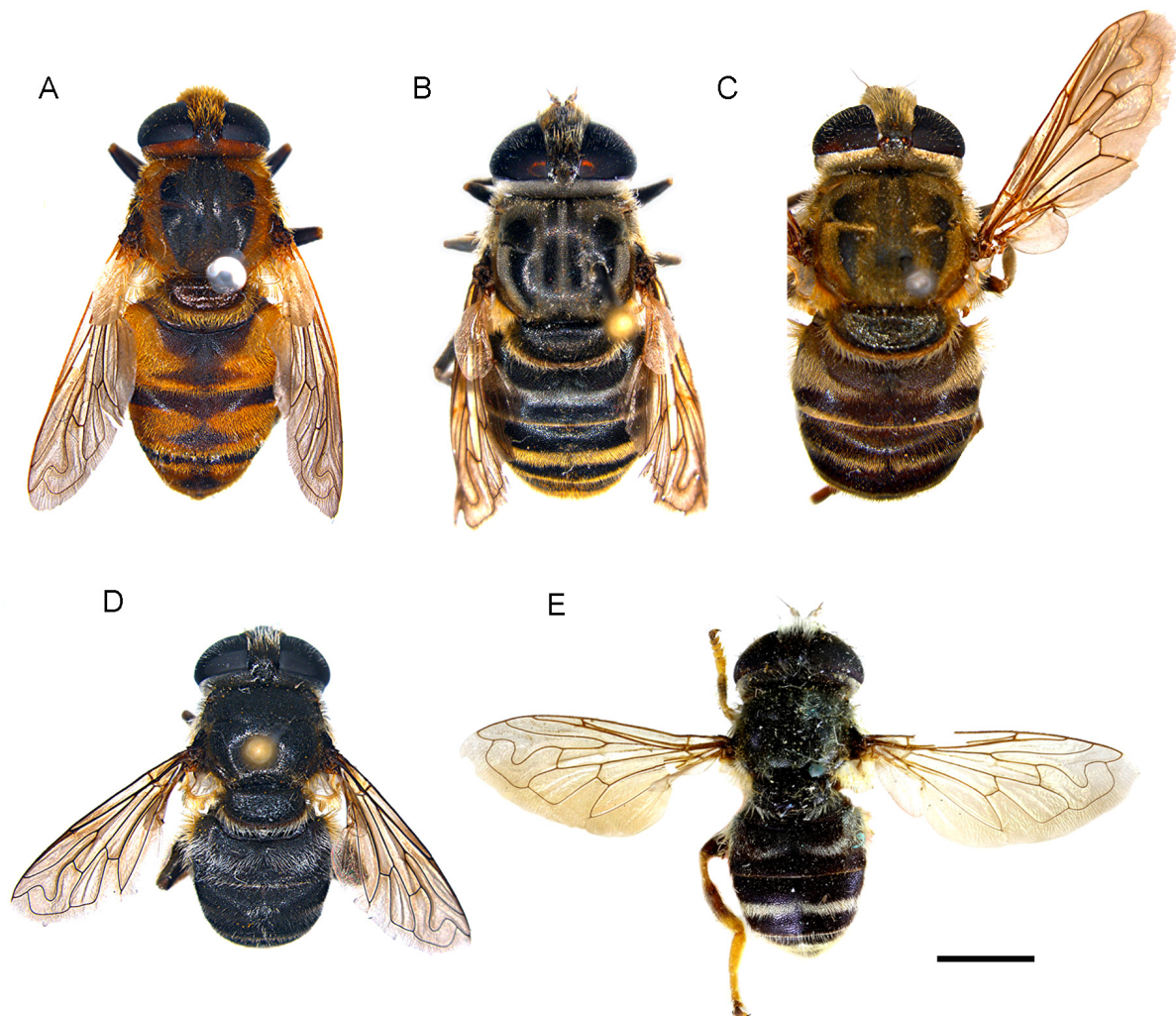


Fig. 14. Habitus of the female, dorsal view. **A.** *Merodon bombiformis* Hull, 1944 (NMSA; FSUNS ID 04278). **B.** *M. multifasciatus* Curran, 1939 (FSUNS; FSUNS ID ZA5_243). **C.** *M. zebra* Vujić & Radenković sp. nov., paratype (RMCA; FSUNS ID 25090). **D.** *M. lotus* Vujić & Radenković sp. nov., paratype (NMSA; FSUNS ID ZA6_067). **E.** *M. nasicus* Bezzi, 1915, lectotype (BMNH NHMUK010369943). Scale bar = 2 mm.

***Merodon nasicus* Bezzi, 1915**

Figs 1, 4F, 5F, 6F, 9F, 10F, 11F, 12F, 14E, 15E, 16E, 18D, 19D–F, 20, 23A–B

Merodon nasicus Bezzi, 1915: 102 (type locality: Kenya, Njoro).

Diagnosis

Small (6–7 mm), black species with narrow, white pollinose, tergal fasciae. Oral margin distinctly protruded (Fig. 9F); face and frons without pollinosity, except narrow vitta along eye margins; terga 2–4 with medial pollinose fasciae (Figs 5F, 14E); tergum 4 covered with whitish to yellowish pile; male genitalia with S-shaped posterior lobe of surstyle, gradually narrowing toward tip (Fig. 19D: pl), anterior margin of surstyle rounded in lateral view (Fig. 19D: ams). Differs from *Merodon bombiformis*, *M. vittatus* sp. nov., *M. multifasciatus* and *M. zebra* Vujić & Radenković sp. nov. by absence of distinct pollinose vittae on scutum (conspicuous in these four species); narrow medial pollinose fasciae on terga 3 and 4 (broad in *M. vittatus* sp. nov., *M. multifasciatus* and *M. zebra* sp. nov.) and male genitalia with moderately broad posterior lobe of surstylus, narrower than in *M. multifasciatus*, but broader than in *M. vittatus* sp. nov. and *M. zebra* sp. nov. (Figs 7A, D: pl, 19D: pl). Similar to *M. lotus* sp. nov. from which differs in presence of distinct medial pollinose fasciae on terga 2 and 3 (absent in males and indistinct in female of *M. lotus* sp. nov.) and more protruded oral margin (Fig. 9E–F).

Material examined

Lectotype (designated here by A. Vujić)

KENYA • 1 ♀; Njoro, about 140 km NW of Nairobi; 0.37° S, 35.917° E; 2322 m a.s.l.; date unknown; A.J. Cholmley leg.; BMNH NHMUK010369943.

Bezzi published the Syrphidae of the Ethiopian Region with the descriptions of two new species, *Merodon planifacies* and *M. melanocerus* (Bezzi 1915). In the same work, he presented diagnostic characters for, and notes on, the third Ethiopian species, *M. nasicus*, referring to 1914 as the year of its description (Bezzi 1915: 102), based on material from Eritrea in the collections of the Hungarian Museum. Bezzi (1915) also stated that there was a single female specimen of this species from Njoro, in British East Africa. Subsequently, in the *Catalogue of the Diptera of the Afrotropical Region* (Smith & Vockeroth 1980), and in the *Diptera Database* (Evenhuis & Pape 2020), this Njoro specimen was erroneously considered as the holotype. Shortly after his 1915 monograph, Bezzi published a description of *Lampetia (Merodon) nasica* (Bezzi 1921), based on a male and female from Eritrea in the collection of the Hungarian Museum. This is actually the unpublished description from 1914 (Bezzi 1915). However, based on *International Code of Zoological Nomenclature* (International Commission on Zoological Nomenclature 1999), the oldest available name for a taxon is its valid name, therefore the publication with the first appearance of the name of the taxon (Bezzi 1915) should be considered as valid. Following from this conclusion, in addition to specimens from Eritrea, cited in both publications (Bezzi 1915, 1921), one female specimen from Kenya (Bezzi 1915), deposited in BMNH, also belongs to the syntype series. The Diptera collection in the Hungarian National Museum in Budapest, including the syntypes from Eritrea, was destroyed by a fire in 1956. However, in order to stabilize nomenclature should additional specimens of the type series be found, we designate the specimen from BMNH as lectotype.

Additional material

BURUNDI • 1 ♂; Bururi; 3.951° S, 29.619° E; 1950 m a.s.l.; 20 Mar. 1949; F.J. François leg.; FSUNS ID 21976; RBINS.

ETHIOPIA • 1 ♂; Welo Gondi, 60 km SW Lalibela; 7.779° N, 35.458° E; 2485 m a.s.l.; 9 Oct. 2005; A. Freidberg leg.; FSUNS ID 04972; TAUJ.

ERITREA • 1 ♀; Adi Keyh (on eastern slope of Ethiopian highlands); 14.816° N, 39.383° E; 2442 m a.s.l.; Sep. 1902; Dr A. Andreini leg.; LSF.

Redescription

Original description without any illustrations was based on single male and female (Bezzi 1921).

Length: small sized species, body 6–7 mm, wing 5–6 mm (n =2).

Male

HEAD (Figs 4F, 9F, 11F, 12F). Antenna (Fig. 4F) reddish-brown; pedicel elongated, approximately as long as basoflagellomere (relation scape:pedicel:basoflagellomere=1.0:3.0:3.0); basoflagellomere concave dorsally, with acute apex; arista reddish-brown, thickened basally, 1.3 times as long as basoflagellomere. Face black, without pollinosity, except along eye margins and covered with whitish to yellowish pile, except on bare medial vitta that occupies $\frac{1}{2}$ width of face. Oral margin strongly protruded (Fig. 9F). Frons black, without gray pollinosity, except eye margins, covered with whitish to yellowish pile. Vertical triangle isosceles (Fig. 12F), usually dark brown, predominantly covered with a long, yellow pile. Ocellar triangle equilateral. Eye pile black in upper half and gray in lower, as long as scape. Eye contiguity about 15–18 ommatidia long. Occiput black, partly pollinose, covered with yellowish to whitish pile.

THORAX (Fig. 6F). Mesonotum dark brown, covered with light yellowish to yellowish pile; scutum without or with indistinct pollinose longitudinal vittae; postpronotum and anterior margin of scutum with sparse gray pollinosity. Pleuron dark brown, covered with gray pollinosity and the following parts with whitish to yellow pile: anterior part of proepimeron, posterior part of anterior anepisternum, most of the posterior anepisternum except anterior end, antero-ventral and postero-dorsal part of katapisternum, anepimeron, metasternum. Wing hyaline, mostly covered with microtrichia; veins yellow to brown. Calypter pale yellow. Halter with yellow pedicel and capitulum. All three femora dark brown to black, except reddish apex; tibiae mostly reddish, medially dark brown; basal three tarsomeres yellowish to reddish, the rest blackish (Fig. 6F). Metatrochanter without calcar. Metafemur moderately thickened, with less developed apicoventral triangular lamina, more or less dentate, the apical dens is distinct (Fig. 6F). Pile on legs predominantly whitish to yellow.

ABDOMEN (Fig. 5F). Dark brown with yellow to whitish-yellow pile on lateral margins. Tergum 1 usually with pollinosity at least laterally, covered with short pile; tergum 2 with narrow medial fascia interrupted in the middle, covered with whitish pile; tergum 2 medially mostly covered with short black pilosity; terga 3 and 4 with or without narrow pollinose posterior margin and distinct medial fascia interrupted in the middle; tergum 3 covered with short black pile medially; tergum 4 covered with longer whitish to yellowish pilosity, except few black pile in anterior half. Brown sterna pollinose, covered with white or yellow pile.

GENITALIA (Figs 18D, 19D–F). Posterior lobe of surstyle moderately broad, bent, S-shaped, gradually narrowing toward tip (Fig. 19D: pl); anterior lobe of surstyle undeveloped (Fig. 19D); anterior margin of surstyle rounded in lateral view (Fig. 19D: ams); cercus square like (Fig. 19D: c). Hypandrium with theca medially distinctly narrowed (Fig. 19F). Lateral sclerite of aedeagus small, pointed (Fig. 18D: s).

Female (Figs 14E, 15E, 16E, 20)

Similar to the male except for normal sexual dimorphism: metafemur usually with very small apical dens on triangular lamina (Figs 16E, 20B); frons with non pollinose medial vitta (Fig. 15E); vertex shiny, non pollinose, mostly covered with black pile.

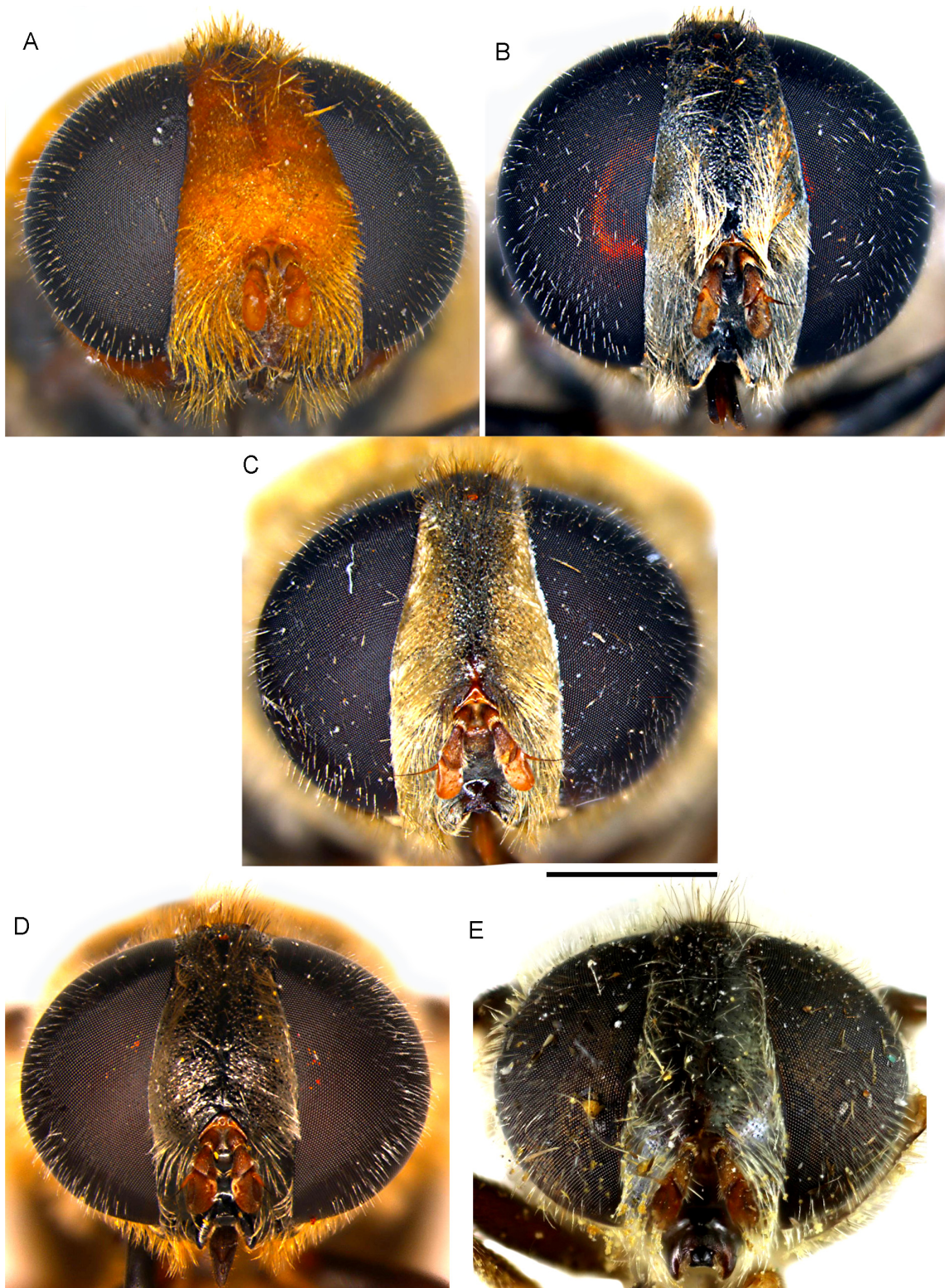


Fig. 15. Female head, frontal view. **A.** *Merodon bombiformis* Hull, 1944 (NMSA; FSUNS ID 04278). **B.** *M. multifasciatus* Curran, 1939 (FSUNS; FSUNS ID ZA5_243). **C.** *M. zebra* Vujić & Radenković sp. nov., paratype (RMCA; FSUNS ID 25090). **D.** *M. lotus* Vujić & Radenković sp. nov., paratype (NMSA; FSUNS ID ZA6_067). **E.** *M. nasicus* Bezzi, 1915, lectotype (BMNH; NHMUK010369943). Scale bar = 2 mm.



Fig. 16. Female metalleg, lateral view. **A.** *Merodon bombiformis* Hull, 1944 (NMSA; FSUNS ID 04278). **B.** *M. multifasciatus* Curran, 1939 (FSUNS; FSUNS ID ZA5_243). **C.** *M. zebra* Vujčić & Radenković sp. nov., paratype (NHMW; FSUNS ID 05725). **D.** *M. lotus* Vujčić & Radenković sp. nov., paratype (NMSA; FSUNS ID ZA6_067). **E.** *M. nasicus* Bezzi, 1915, lectotype (BMNH NHMUK010369943). Scale bar = 2 mm.

Period of flight and distribution (Fig. 1)

Merodon nasicus is known from Kenya, Ethiopia, Eritrea and Burundi. On the basis of our data, the flight period was found to be in March and from September to October. This species occurs only on high mountains at elevation between 2000 and 2500 m a.s.l. in vegetation of montane forest-tundra (Sayre *et al.* 2013).

***Merodon vittatus* Vujić & Likov sp. nov.**

urn:lsid:zoobank.org:act:591CEDD4-80E4-4978-B13E-3F66511C9026

Figs 1, 4C, 5C, 6C, 7D–F, 8B, 9C, 10C, 11C, 12C, 18C, 21E

Diagnosis

Brown, large (12 mm), striped species, with golden yellow pile on the tip of abdomen (Fig. 5C); frons and postpronotum dark brown; scutum with five pollinose vittae (Fig. 10C); metafemur with distinct apical thorn on triangular process (Figs 6C, 8B); terga 3 and 4 with broad medial pollinose fasciae (Fig. 5C). Similar to *Merodon multifasciatus* from which differs only in male genitalia with narrow, S-shaped posterior lobe of surstyle, gradually narrowing toward tip (Fig. 7D: pl) (in *M. multifasciatus* broad, slightly curved, with almost parallel sides and rounded apex (Fig. 13D: pl).

Etymology

The specific epithet '*vittatus*' 'wearing or carrying a vitta', an adjective, referring to the broad golden gray fasciae on the terga 2–4.

Material examined

Holotype

GHANA • 1 ♂; Transvolta Togoland, Amedzofe; 6.853° N, 0.433° E; 2 Nov. 1959; N.D. Jago leg.; FSUNS ID 04085; NBCN.

Description

Length: large species, body 12 mm, wing 8.5 mm (n = 1).

Male

HEAD (Figs 4C, 9C, 11C, 12C). Antenna (Fig. 4C) reddish-yellow; pedicel elongated, approximately as long as basoflagellomere (relation scape : pedicel : basoflagellomere = 1.0 : 2.5 : 2.5); basoflagellomere concave dorsally, with acute apex; arista yellow to reddish, thickened basally, 1.7 times as long as basoflagellomere. Face dark brown, covered with gray pollinosity and whitish pile, except on bare medial vitta that occupies ¼ width of face. Oral margin protruded (Fig. 9C). Frons dark brown, inflated, covered with gray pollinosity and whitish pile. Vertical triangle isosceles (Fig. 12C), reddish, covered with a long, whitish pile. Ocellar triangle equilateral. Eye pile dense, gray, slightly longer than scape. Eye contiguity about 15 ommatidia long. Occiput reddish-brown, pollinose, covered with a whitish pile.

THORAX (Figs 6C, 8B, 10C). Mesonotum black, except lateral side of scutum including postpronotum, postalar callus, and posterior margin of scutellum red-brown; covered with grayish-yellow pile; scutum with five pollinose longitudinal vittae (Fig. 10C). Pleuron black to dark brown, covered with gray pollinosity and the following parts with whitish to yellow pile: anterior part of proepimeron, posterior part of anterior anepisternum, most of the posterior anepisternum except anterior end, antero-ventral and postero-dorsal part of katepisternum and anepimeron. Wing hyaline, with dense microtrichia and yellow to brown veins. Calypter pale yellow. Halter with pale yellow pedicel and capitulum. All three femora dark brown; tibiae mostly reddish, medially dark brown; tarsi reddish (Figs 6C, 8B). Metatrochanter without calcar. Metafemur moderately thickened, with less developed apicoventral triangular lamina,

more or less dentate, the apical dens is distinct (Figs 6C, 8B). Pile on legs predominantly whitish to yellow.

ABDOMEN (Fig. 5C). Black to dark brown. Tergum 1 with gray pollinosity, covered with black basal sockets of pile and with short pale yellow pile; tergum 2 with pollinose posterior margin and narrow medial fascia, interrupted in the middle, covered with pale yellow pile; terga 3 and 4 with pollinose posterior margin and broad medial fasciae (Fig. 5C); tergum 3 covered with pale yellow pile except anterior half with few short black pile; tergum 4 covered with long golden-yellow pilosity (Fig. 5C). Sterna reddish-brown, covered with pale yellow pile.

GENITALIA (Figs 7D–F, 18C). Posterior lobe of surstyle S-shaped, gradually narrowing toward tip (Fig. 7D: pl); anterior margin of surstyle angular (Fig. 7D: ams); cercus elongated (Fig. 7D: c). Hypandrium with theca medially distinctly narrowed (Fig. 7F). Lateral sclerite of aedeagus small (Fig. 18C: s).

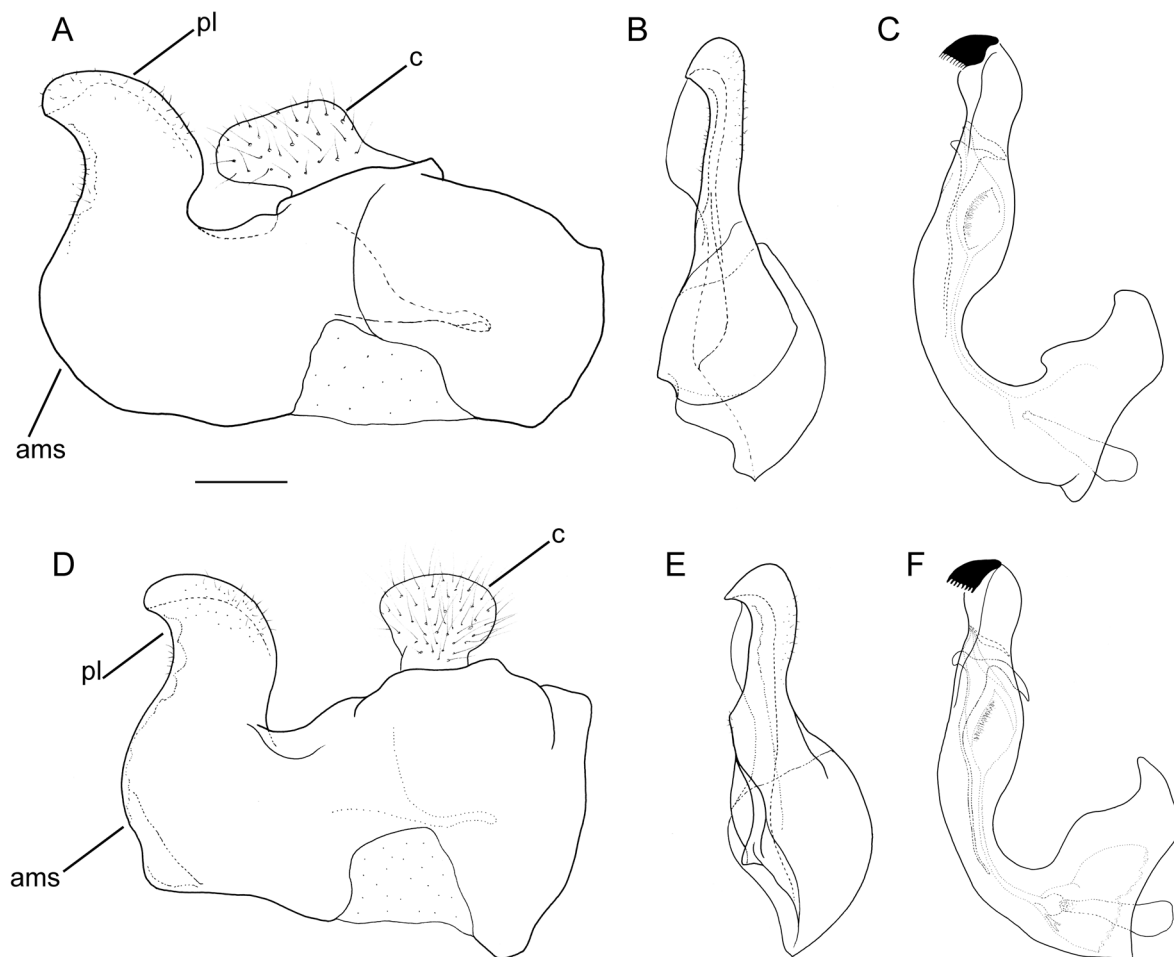


Fig. 17. *Merodon lotus* Vujić & Radenković sp. nov., male genitalia. Paratypes. **A–C.** Specimen FSUNS (FSUNS ID ZA6_066). **D–F.** Specimen FSUNS (FSUNS ID ZA6_060). **A, D.** Epandrium, lateral view. **B, E.** Epandrium, ventral view. **C, F.** Hypandrium, lateral view. Abbreviations: ams = anterior margin of surstyle; c = cercus; pl = posterior lobe of surstyle. Scale bar = 0.25 mm.

Female

Unknown.

Period of flight and distribution (Fig. 1)

Only the holotype is known, which originated from Ghana and was collected in November. It was found in a deciduous forest-woodland savanna type of vegetation.

***Merodon zebra* Vujić & Radenković sp. nov.**

urn:lsid:zoobank.org:act:9B6ABB0D-7FF4-4217-9C23-FBD06F8D80F1

Figs 1, 3D, 4D, 5D, 6D, 7A–C, 9D, 10D, 11D, 12D, 14C, 15C, 16C, 22D–F

Diagnosis

Large (10–12 mm), dark, striped species, with golden-yellow pile on the tip of abdomen (Fig. 5C). Scutum with conspicuous pollinose vittae (Fig. 10D); metafemur with medium sized apical dens on triangular lamina (Figs 6D, 16D); terga 3 and 4 with broad medial pollinose fasciae (Fig. 5D); male genitalia with S-shaped posterior lobe of surstyle, gradually narrowing toward tip (Fig. 7A: pl); anterior margin of surstyle rounded in lateral view (Fig. 7A: ams). Similar to *Merodon vittatus* sp. nov. and *M. multifasciatus* from which differs in less distinct dens on apicoventral triangular lamina of metafemur; shorter eye contiguity, about 10 ommatidia long, while in *M. vittatus* sp. nov. is about 15, and in *M. multifasciatus* about 15–20; and shape of male genitalia (rounded anterior margin of surstyle (Fig. 7A: ams), in *M. vittatus* sp. nov. angular in lateral view (Fig. 7D: ams); narrow, S-shaped posterior lobe of surstyle, gradually narrowing toward tip (Fig. 7A: pl), in *M. multifasciatus* broad, slightly curved, with almost parallel and rounded apex (Fig. 13D: pl).

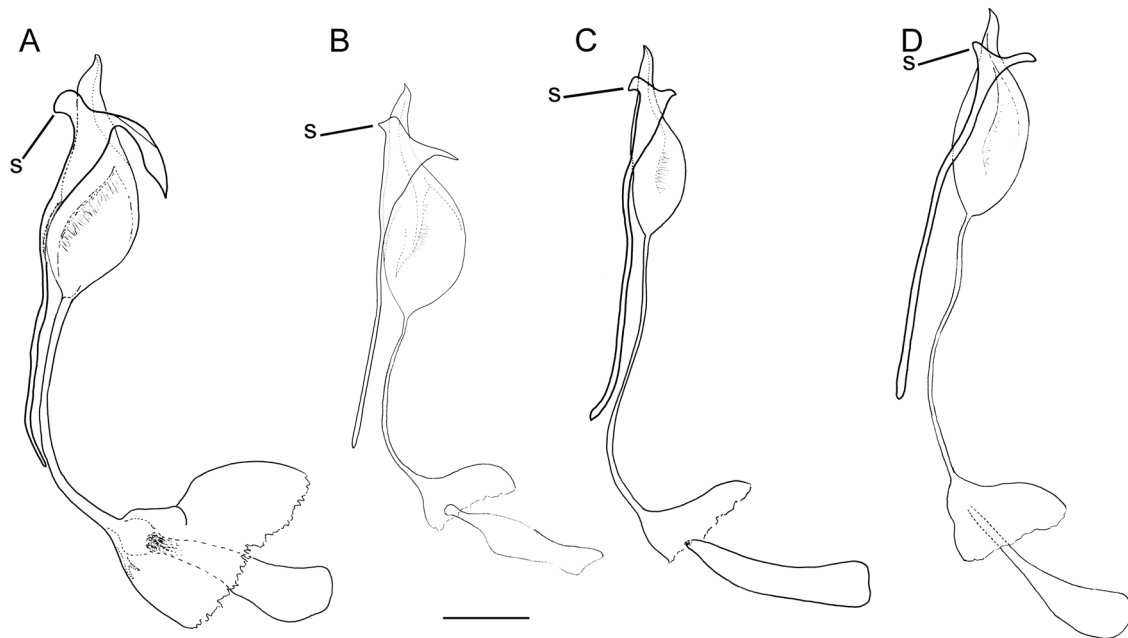


Fig. 18. Male genitalia, aedeagus, lateral view. **A.** *Merodon lotus* Vujić & Radenković sp. nov., paratype (FSUNS; FSUNS ID ZA6_060). **B.** *M. multifasciatus* Curran, 1939 (NBCN LML-05-3; FSUNS ID 03909). **C.** *M. vittatus* Vujić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **D.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04972). Abbreviation: s = lateral sclerite of aedeagus. Scale bar = 0.25 mm.

Etymology

The specific epithet ‘zebra’ was given as a reference to the African equine, zebra with a distinctive black and white striped coat, pattern like in this *Merodon* species. It is to be treated as a noun in apposition. The name ‘zebra’ was proposed by W. Hurkmans in an unpublished manuscript, but unfortunately the designated holotype belongs to *M. multifasciatus*. We decided to keep this descriptive name for this African endemic.

Material examined

Holotype

DEMOCRATIC REPUBLIC OF THE CONGO • 1 ♂; Lubumbashi (former Elisabethville); 11.589° S, 27.427° E; 4 Feb. 1923; Dr M. Bequaert leg.; FSUNS ID 25089; RMCA.

Paratypes

DEMOCRATIC REPUBLIC OF THE CONGO • 1 ♂, 1 ♀; Lualaba River; 7.809° S, 26.919° E; 562 m a.s.l.; 2500–4000 ft; 17 May 1907; S.A. Neave leg.; FSUNS ID 03314, 03911; BMNH • 1 ♂; same collection data as for preceding; 30 May 1907; FSUNS ID 03912; NBCN AM-05-073 • 1 ♂, 1 ♀; same collection data as for preceding; FSUNS ID 03316, 03910; BMNH • 1 ♂; same collection data as for preceding; 31 May 1907; FSUNS ID 03315; BMNH • 1 ♀; Kasongo; 4.45° S, 26.666° E, 661 m a.s.l.; 7 Nov. 1912; Dr M. Bequaert leg.; FSUNS ID 25090; RMCA • 1 ♀; same collection data as for holotype; 16 Jan. 1921; FSUNS ID 25091; RMCA • 1 ♂; Lubumbashi (former Elisabethville), R. Kimilol; 11.589° S, 27.427° E; Dec. 1928; Dr M. Bequaert leg.; FSUNS ID 05174; NBCN.

TANZANIA • 1 ♀; Tanganyika Territory (former), Ugano, Matengo Highland WSW of Songea; 10.949° S, 34.934° E; 1500–1700 m a.s.l.; 11–20 Jan. 1936; H. Zerny leg.; FSUNS ID 05725; NHMW.

Description

Length: large species, body 10–12 mm, wing 7–8 mm (n = 4).

Male

HEAD (Figs 4D, 9D, 11D, 12D). Antenna (Fig. 4D) reddish-brown; pedicel elongated, approximately as long as basoflagellomere (relation scape : pedicel : basoflagellomere = 1.0 : 2.5 : 2.5); basoflagellomere concave dorsally, with acute apex; arista light brown to dark brown, thickened basally, 1.6 times as long as basoflagellomere. Face from black to dark brown, covered with gray pollinosity and whitish pile, except on bare medial vitta that occupies $\frac{1}{4}$ width of face. Oral margin protruded (Fig. 9D). Frons from black to reddish, inflated, covered with gray pollinosity and whitish pile. Vertical triangle isosceles (Fig. 12D), usually dark brown, predominantly covered with a long, yellow pile. Ocellar triangle equilateral. Eye pile dense, gray, slightly longer than scape. Eye contiguity about 10 ommatidia long. Occiput reddish, pollinose, covered with yellow pile.

THORAX (Figs 6D, 10D). Mesonotum black, except postpronotum and posterior margin of scutellum brown-reddish, covered with reddish-yellow pile; scutum with five pollinose longitudinal vittae (as on Fig. 10D). Pleuron black to dark brown, covered with gray pollinosity and the following parts with whitish to yellow pile: anterior part of proepimeron, posterior part of anterior anepisternum, most of the posterior anepisternum except anterior end, antero-ventral and postero-dorsal part of katepisternum, anepimeron and metasternum. Wing hyaline, with dense microtrichia and yellow to brown veins. Calypter pale yellow. Halter with yellow pedicel and capitulum. All three femora dark brown to black; tibiae mostly reddish, medially black to dark brown; tarsi reddish, except darker apical two tarsomeres (Fig. 6D). Metatrochanter without calcar. Metafemur moderately thickened, with less developed apicoventral triangular lamina, more or less dentate, the apical dens is distinct (Fig. 6D). Pile on legs predominantly whitish to yellow.

ABDOMEN (Fig. 5D). Black to dark brown. Tergum 1 black, usually with pollinosity, covered with short whitish to yellow pile; tergum 2 with pollinose posterior margin and narrow medial fascia, in some specimens interrupted in the middle, with whitish pile except posterior half between pollinose fasciae covered with short black pilosity; terga 3 and 4 with pollinose posterior margin and broad medial fasciae; anterior half of tergum 3 covered with short black pile; tergum 4 covered with long golden yellow pilosity. Sterna reddish-brown, covered with white or yellow pile.

GENITALIA (Figs 3D, 7A–C). Posterior lobe of surstyle narrow, bent, S-shaped, gradually narrowing toward tip (Fig. 7A: pl); anterior lobe of surstyle undeveloped (Fig. 7A); anterior margin of surstyle rounded in lateral view (Fig. 7A: ams); cercus square like (Fig. 7A: c). Hypandrium with theca medially distinctly narrowed (Fig. 7C: marked with arrow). Lateral sclerite of aedeagus small (Fig. 3D: s).

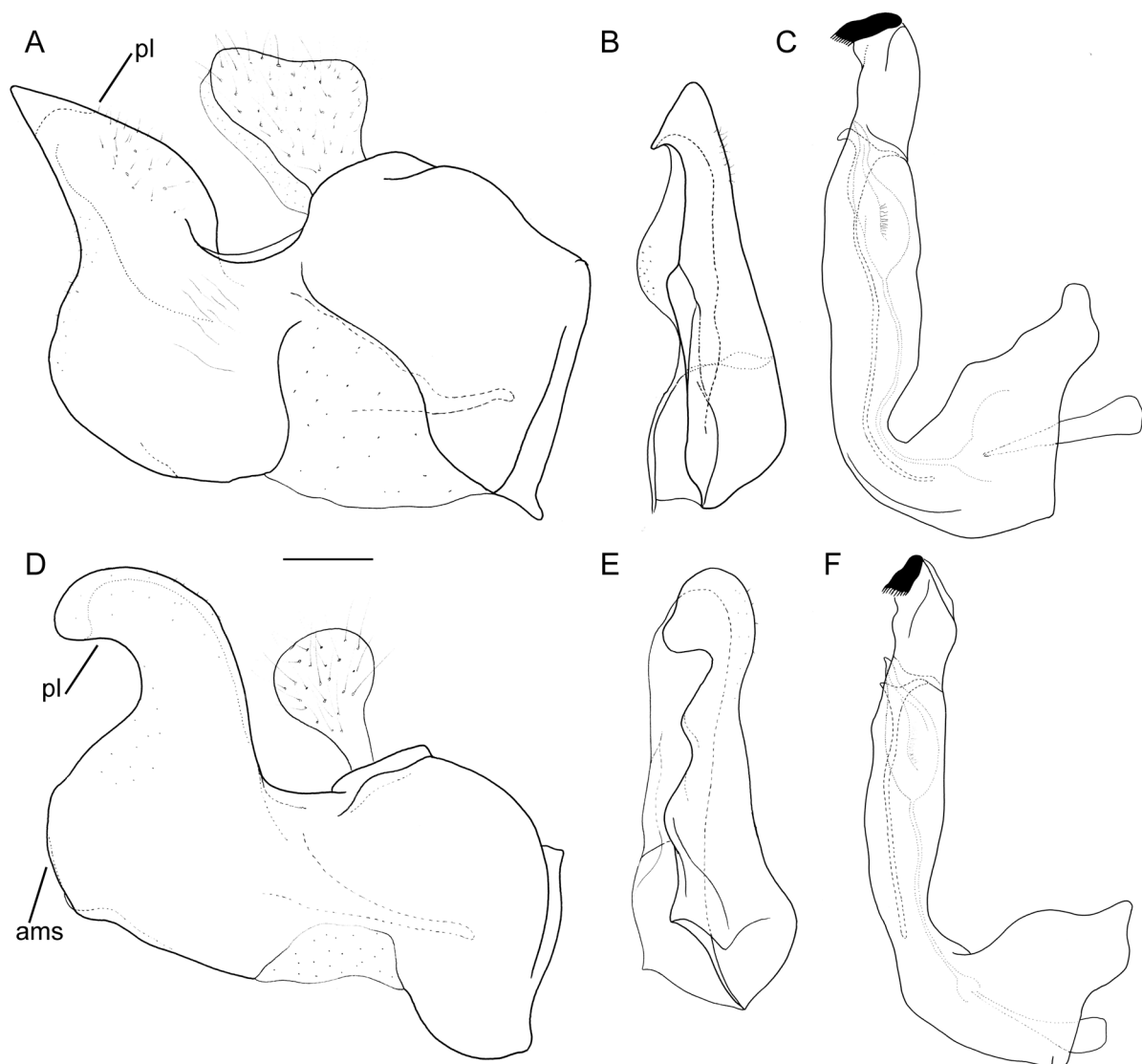


Fig. 19. Male genitalia. **A–C.** *Merodon funestus* (Fabricius, 1794) (FSUNS; FSUNS ID F99). **D–F.** *M. nasicus* Bezzi, 1915 (TAUI; FSUNS ID 04792). **A, D.** Epandrium, lateral view. **B, E.** Epandrium, ventral view. **C, F.** Hypandrium, lateral view. Abbreviations: ams = anterior margin of surstyle; pl = posterior lobe of surstyle. Scale bar = 0.25 mm.

Female (Figs 14C, 15C, 16C)

Similar to the male except for normal sexual dimorphism: metafemur usually with small apical dens on triangular lamina (Fig. 16C); frons with non pollinose medial vitta (Fig. 15C); vertex shiny, non pollinose, mostly covered with black pile.

Period of flight and distribution (Fig. 1)

Merodon zebra sp. nov. occurs in central parts of Africa (Democratic Republic of the Congo and Tanzania); the flight period may extend over the whole year and probably depends on local circumstances. It appears in a deciduous forest-woodland savanna type of vegetation (Sayre *et al.* 2013).

Names of *Merodon* in the Afrotropical fauna with unresolved status

Merodon edentulus Macquart, 1855 nom. dub., stat. rev.

Merodon edentulus Macquart, 1855: 90.

This name was introduced by Macquart (1855) based on a single specimen from South Africa. The type material is lost. Curran (1939) stated that this can scarcely be a *Merodon* because it is said to

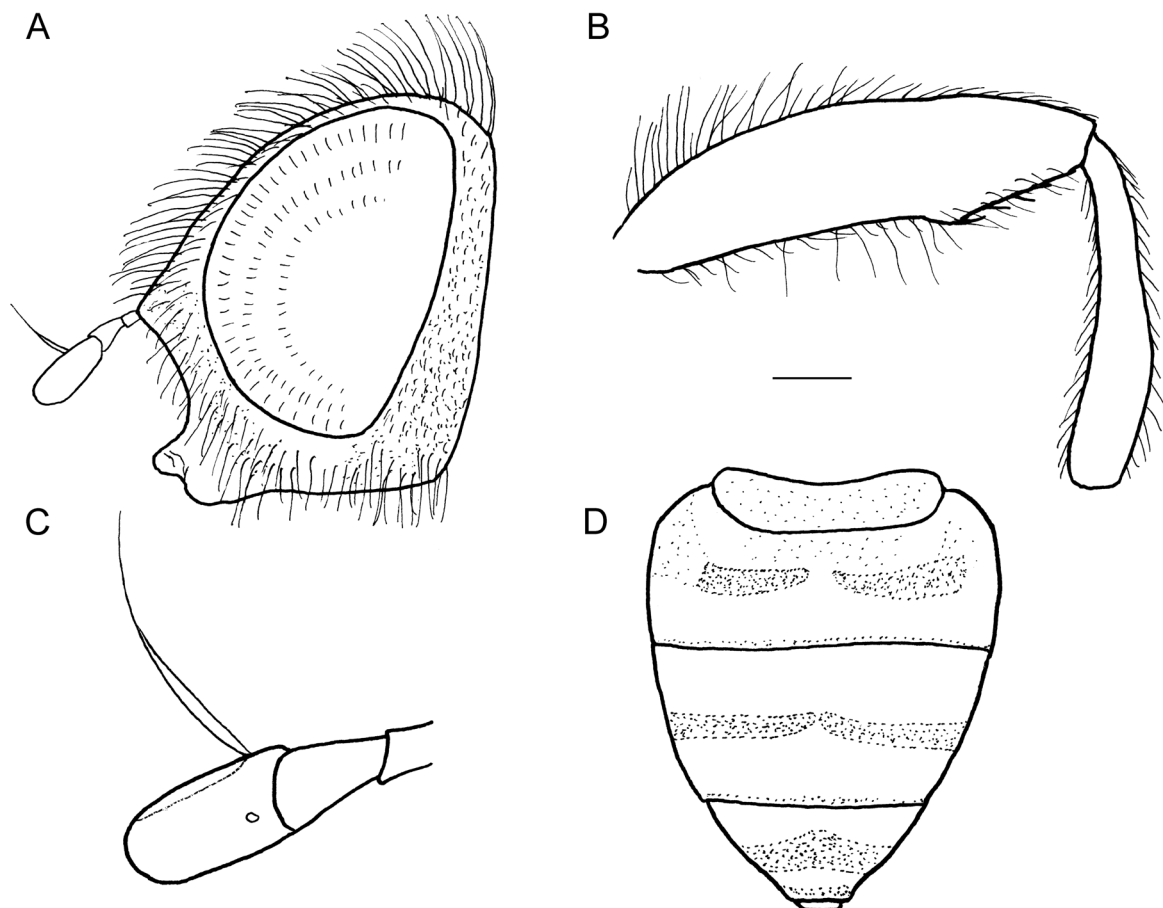


Fig. 20. *Merodon nasicus* Bezzi, 1915, lectotype, ♀ (BMNH NHMUK010369943). **A.** Head, lateral view. **B.** Metafemur and metatibia, lateral view. **C.** Antenna, lateral view. **D.** Abdomen, dorsal view. Scale bar = 1 mm.

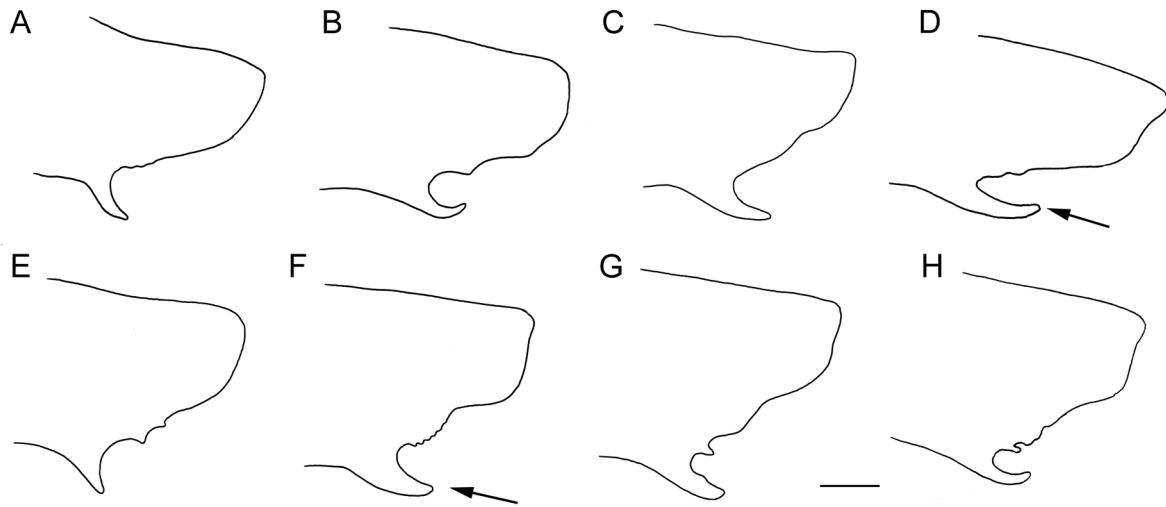


Fig. 21. Apical part of male metafemur, lateral view. **A–D.** *Merodon bombiformis* Hull, 1944 **A.** Specimen FSUNS (FSUNS ID ZA2_079). **B.** Specimen NMSA (FSUNS ID 04276). **C.** Specimen NMSA (FSUNS ID 04277). **D.** Specimen NBCN LML-05-5 (FSUNS ID 04282). **E.** *M. vittatus* Vujić & Likov sp. nov., holotype (NBCN; FSUNS ID 04085). **F–G.** *M. multifasciatus* Curran, 1939. **F.** Specimen NMSA (FSUNS ID 04284). **G.** Specimen NBCN LML-05-3 (FSUNS ID 03909). **H.** *M. apimina* Hull, 1944, paratype (NBCN; FSUNS ID 04086). **D, F.** Distinct apical dent on triangular process marked with arrow. Scale bar = 1 mm.

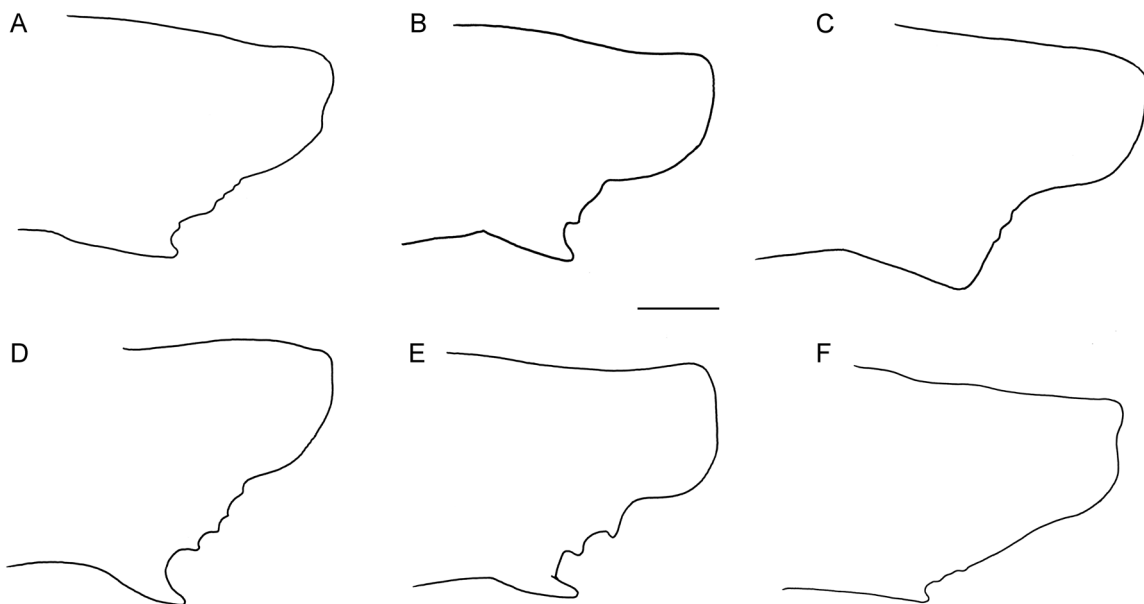


Fig. 22. Apical part of male metafemur, lateral view. **A–C.** *Merodon lotus* Vujić & Radenković sp. nov. Paratypes. **A.** Specimen FSUNS (FSUNS ID ZA6_052). **B.** Specimen FSUNS (FSUNS ID ZA6_060). **C.** Specimen FSUNS (FSUNS ID ZA6_066). **D–F.** *M. zebra* Vujić & Radenković sp. nov. **D.** Holotype (RMCA; FSUNS ID 25089). **E.** Paratype (NBCN; FSUNS ID 05174). **F.** Paratype (RMCA; FSUNS ID 25090). Scale bar = 1 mm.

have no thorn (lamina) on the metafemur. Type locality is Cape of Good Hope (RSA) from where no specimen of *Merodon* was collected. The description is insufficient, even for an identification to the genus level. In the same publication, Macquart (1855) described one other exotic *Merodon* from South America, *Merodon angustiventris* Macquart, 1855 which was re-identified as a member of the genus *Quichuana* Knab, 1913 (Thompson 1988). Therefore, we regard the name *Merodon edentulus* to be a nomen dubium.

Variability of species in the *Merodon bombiformis* group

A variability in shape of the triangular lamina of the metafemur is present in most of the species of the *Merodon bombiformis* group, as seen in figures of *M. bombiformis* (Fig. 21A–D), *M. lotus* sp. nov. (Fig. 22A–C) *M. multifasciatus* (Fig. 21F–H), *M. nasicus* (Fig. 23A–B) and *M. zebra* sp. nov. (Fig. 22D–F). The triangular lamina is more or less dentate, and the apical dens is distinct (*M. bombiformis*, *M. multifasciatus*, *M. vittatus* sp. nov.) or less distinct, as in *M. lotus* sp. nov. and *M. nasicus*. A scutum with conspicuous pollinose vittae, and terga with pollinose posterior margin and medial fasciae, are characters present in most species, except *M. lotus* sp. nov., characterized by the absence of pollinosity on thorax and abdomen (in females a pair of indistinct pollinose fasciae on tergum 4). Width and shape of vittae and fasciae on scutum and terga vary, especially in *M. multifasciatus*, *M. nasicus* and *M. zebra* sp. nov. The colour of the postpronotum, lateral sides of scutum, frons and face in *M. multifasciatus* vary from reddish-brown to black. Specimens from the south of the range (RSA) are darker than individuals from northern parts (Central Africa).

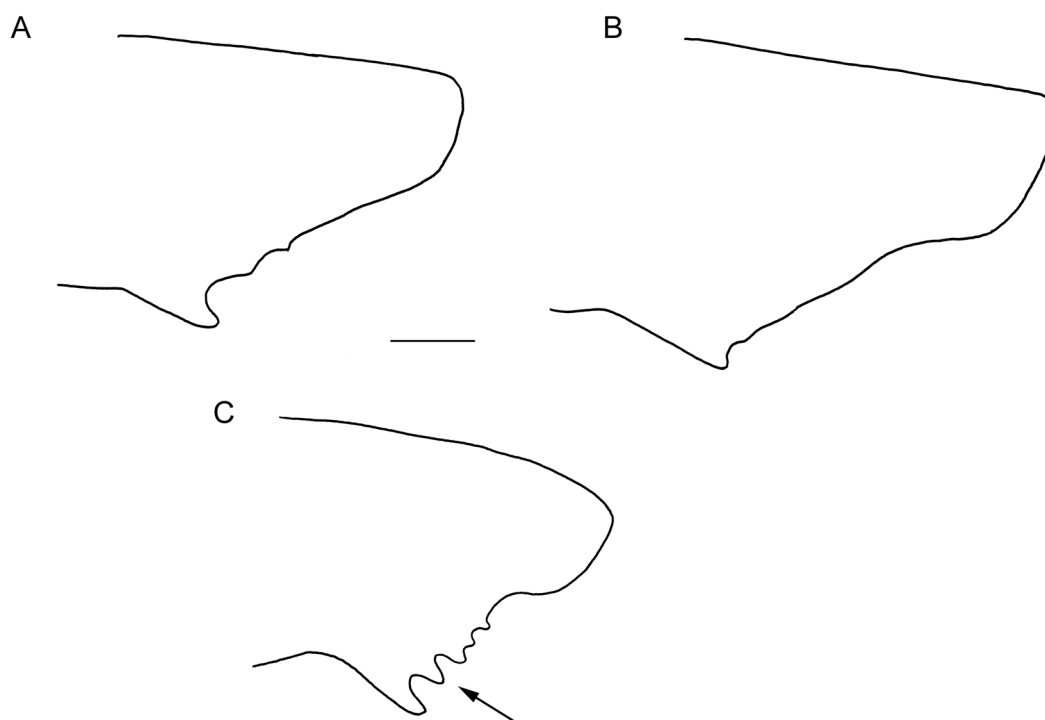


Fig. 23. Apical part of male metafemur, lateral view. **A–B.** *Merodon nasicus* Bezzi, 1915. **A.** Specimen RBINS (FSUNS ID 29176). **B.** Specimen TAUI (FSUNS ID 04972). **C.** *M. funestus* (Fabricius, 1794) (FSUNS; FSUNS ID F98), dentate apicoventral triangular lamina on metafemur marked with arrow. Scale bar = 1 mm.

Key to the species of the *Merodon bombiformis* species group

Female of *M. vittatus* sp. nov. is unknown.

1. Species with reddish-yellow/golden face, frons (Figs 9A, 15A), lateral sides of scutum (Figs 10A, 14A) and most of terga (Figs 5A, 14A); oral margin reduced (Fig. 9A); antennal segments very short (Fig. 4A) *Merodon bombiformis* Hull, 1944
 - Species with mostly dark face, frons, lateral sides of scutum and most of terga; oral margin protruded (Fig. 9B–F); antennal segments longer (Fig. 4B–F) 2
2. Large, dark/brownish species (body length 10–14 mm) with distinct pollinose fasciae on terga 2–4 covered with whitish to yellow pile (Figs 5B–D, 14B–C); tergum 4 with broad medial pollinose fasciae; scutum with five longitudinal pollinose vittae; face dark, mostly covered with gray pollinosity 3
 - Smaller, black species (body length 6–9 mm) without or with less distinct pollinose fasciae on terga 2–4 (Fig. 5E–F); tergum 4 with less distinct medial pollinose fasciae; scutum without or with indistinct pollinose vittae; face shiny black without pollinosity 5
3. Metafemur with shorter and less dense pilosity, especially ventro-laterally (Figs 6B–C, 16B); eye contiguity longer, more than 15 ommatidia long; anterior margin of surstyle angular in lateral view (Figs 7D, 13D) 4
 - Metafemur with long and dense pilosity, especially ventro-laterally (Figs 6D, 16C); eye contiguity shorter, about 10 ommatidia long; anterior margin of surstyle rounded in lateral view (Fig. 7A: ams) *Merodon zebra* Vujić & Radenković sp. nov.
4. Male genitalia with narrow, S-shaped posterior lobe of surstyle, gradually narrowing toward tip (Fig. 7D) (female unknown) *Merodon vittatus* Vujić & Likov sp. nov.
 - Male genitalia with broad posterior lobe of surstyle, slightly curved, with almost parallel sides and rounded apex (Fig. 13D) *Merodon multifasciatus* Curran, 1939
5. Terga 2–4 without pollinosity, only with medial fasciae of whitish pile (Figs 5E, 14D) (in female a pair of indistinct pollinose fasciae on tergum 4) *Merodon lotus* Vujić & Radenković sp. nov.
 - Terga 2–4 with pairs of medial pollinose fasciae, covered with whitish to yellowish pile (Figs 5F, 14E) *Merodon nasicus* Bezzi, 1915

Collecting effort

In the period 2011–2018, eight expeditions to RSA were organized with 235 field working days. During these days, 187 specimens of *Merodon* were collected, among them 11 specimens belonging to the *M. bombiformis* group, identified as three species. A single specimen of *M. bombiformis* was collected on field day 108 (in 2016) on the Drakensberg Mountains (Fig. 24). Subsequently, we focused on two other taxa of the *M. bombiformis* group, previously detected in RSA. Namely, *Merodon multifasciatus* was recorded on day 215 (in 2018) in the province Mpumalanga, near Emgwenya (Waterval Boven) (Fig. 25). A third species was known from a single specimen deposited in the Natural History Museum, London (previously unpublished), with Ceres as locality on the label. We revisited the Ceres area 18 times in the period 2015–2018 and found a population of *M. lotus* sp. nov. on day 230 (in 2018) of our fieldwork (Fig. 26).

Molecular analysis

The specimens of the three collected *Merodon bombiformis* group species (*M. bombiformis*, *M. multifasciatus* and *M. lotus* sp. nov.) were used to extract DNA for molecular analysis. COI sequences

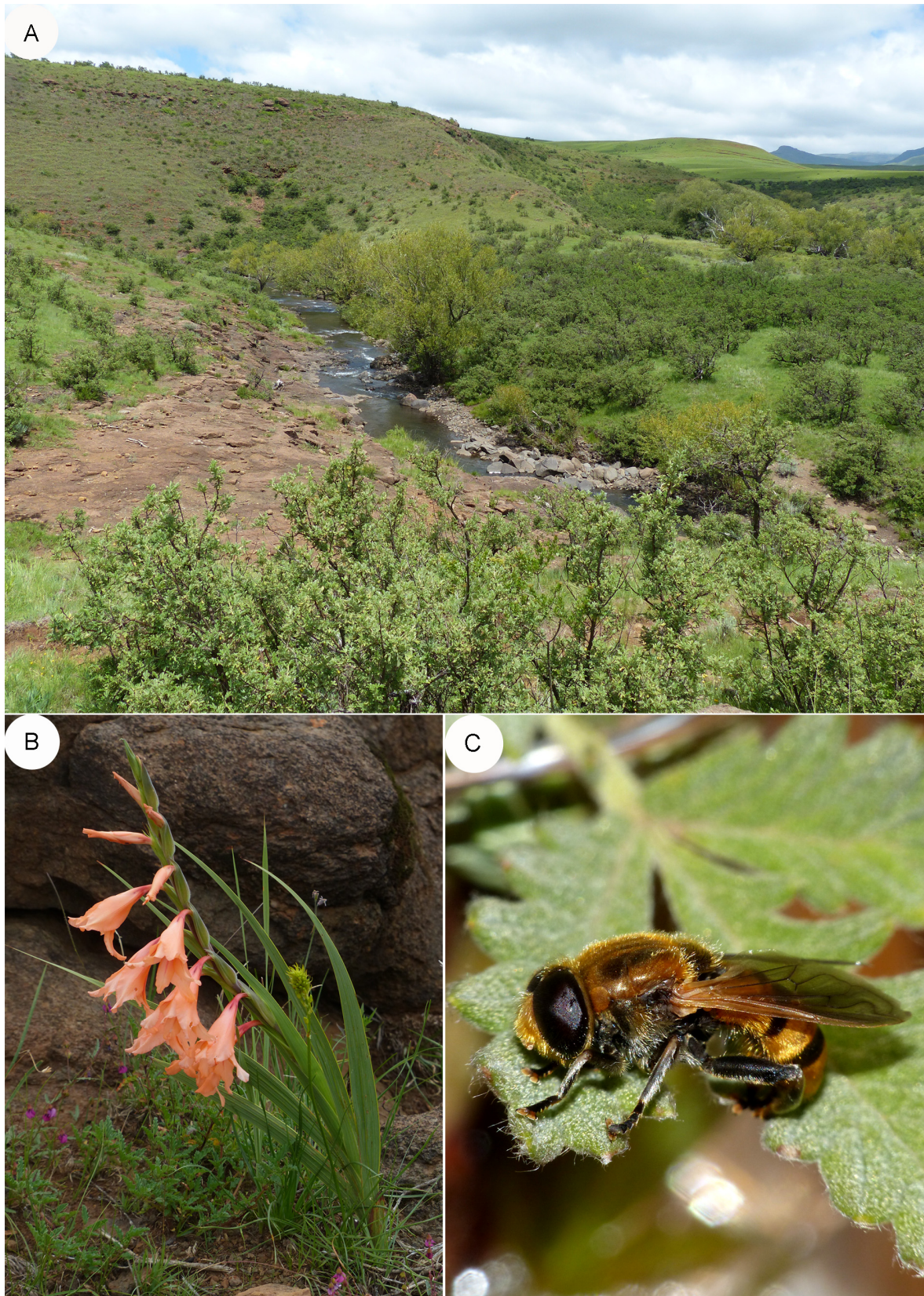


Fig. 24. Habitat of *Merodon bombiformis* Hull, 1944. **A.** Drakensberg Mountains, Maclear-Naudes Nek. **B.** Likely host plant *Gladiolus* sp. **C.** *Merodon bombiformis* male habitus. Photos by Axel Ssymank.

were produced for one specimen of *M. bombiformis*, two *M. multifasciatus* and three specimens of *M. lotus* sp. nov. The obtained COI sequences were 637bp in length and show high uncorrected pairwise genetic distances between species (Table 1). The mean interspecific divergence between species are ~8.4% between *M. multifasciatus* and *M. lotus* sp. nov., ~14.9% between *M. bombiformis* and *M. multifasciatus*, and ~15.5% between *M. bombiformis* and *M. lotus* sp. nov.

The ML phylogenetic analysis of the concatenated dataset (1858 bp long) consisting of one mitochondrial gene (COI) and two nuclear ribosomal genes (28S and 18S) produced the tree presented in Fig. 27. *Merodon multifasciatus* and *M. lotus* sp. nov. are resolved in a clade with high bootstrap support value

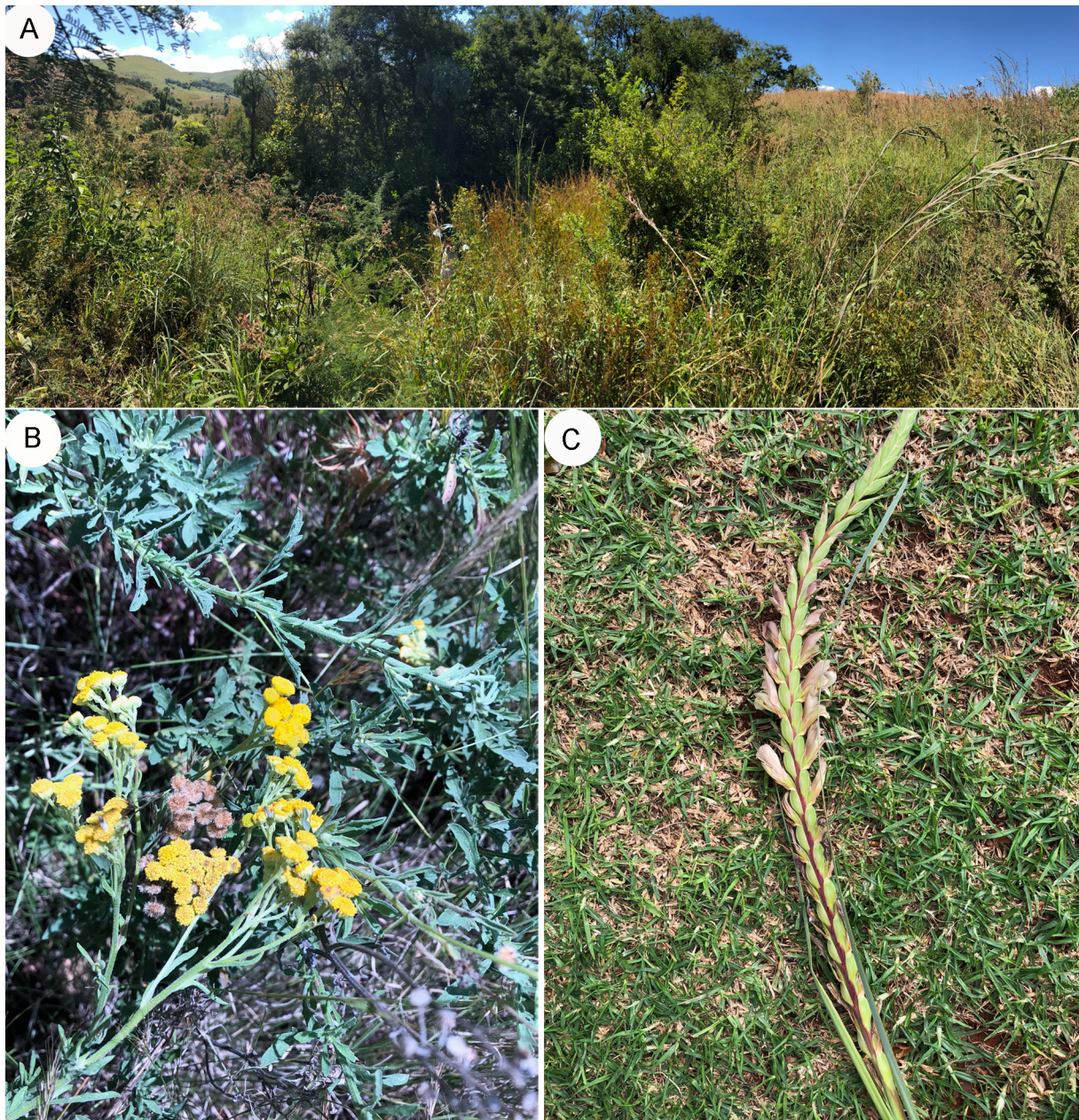


Fig. 25. Habitat of *Merodon multifasciatus* Curran, 1939. **A.** Mpumalanga, near Emgwenya (Waterval Boven). **B.** *Nidorella auriculata* DC. **C.** *Gladiolus sericeovillosus* Hook. Photos by Branka Božičić Lothrop.

Table 1. Uncorrected pairwise distances of COI gene sequences among specimens of species in the *M. bombiformis* species group.

	1.	2.	3.	4.	5.
1. <i>M. bombiformis</i> AF68					
2. <i>M. multifasciatus</i> ZA5_242	0.149				
3. <i>M. multifasciatus</i> ZA5_243	0.149	0.000			
4. <i>M. lotus</i> sp. nov. AU1710	0.157	0.086	0.086		
5. <i>M. lotus</i> sp. nov. AU1712	0.154	0.082	0.082	0.005	
6. <i>M. lotus</i> sp. nov. AU1714	0.154	0.082	0.082	0.005	0.000

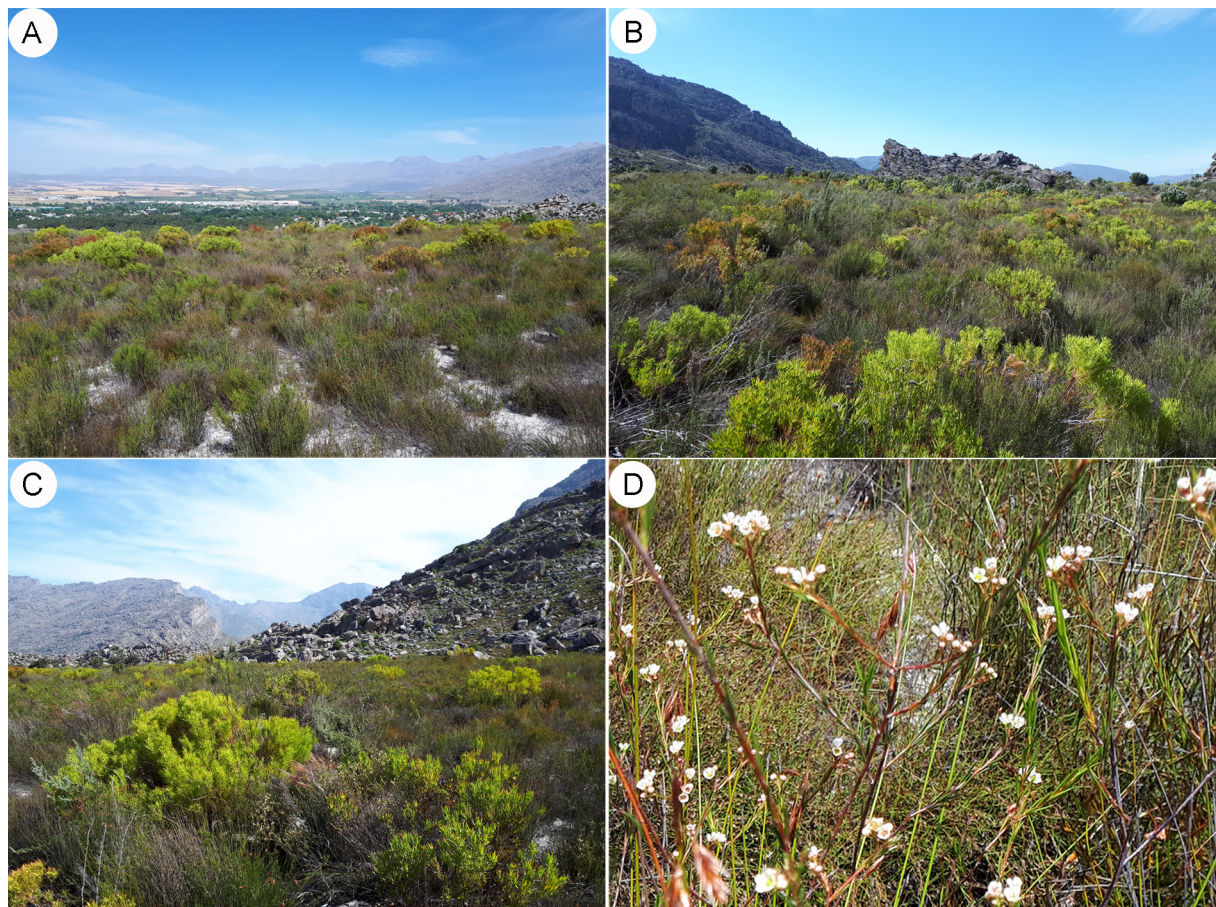


Fig. 26. Habitat of *Merodon lotus* Vujić & Radenković sp. nov. **A–C.** Ceres. **D.** *Coleonema* sp. Photos by Ante Vujić.

(100). *Merodon bombiformis* represents a separate branch within the *aureus* lineage, but this placement on the tree has low support (bootstrap value <50).

Discussion

The majority of species of *Merodon* are present in Southern Europe and Asia Minor, which coincides with the high diversity of geophytes (their larval host plants) in these regions. Out of a total of ca 160 species of *Merodon*, based on recent literature and the present work, only 16 are known from the Afrotropical region. Of the five monophyletic lineages of *Merodon* (Vujić *et al.* 2019a), only two, *aureus* and *desuturinus*, have representatives in the Afrotropical region, as well as in the Palaearctic. The *Merodon desuturinus* lineage consists of two clearly separate groups of species based on both adult morphological and molecular data: the Palaearctic group with four species (Vujić *et al.* 2018) and the Afrotropical group with 10 (Radenković *et al.* 2018; Djan *et al.* 2020).

The *Merodon bombiformis* group consists of six related, though clearly morphologically separated, species distributed in central and southern Africa. The most divergent species are *M. bombiformis*, with

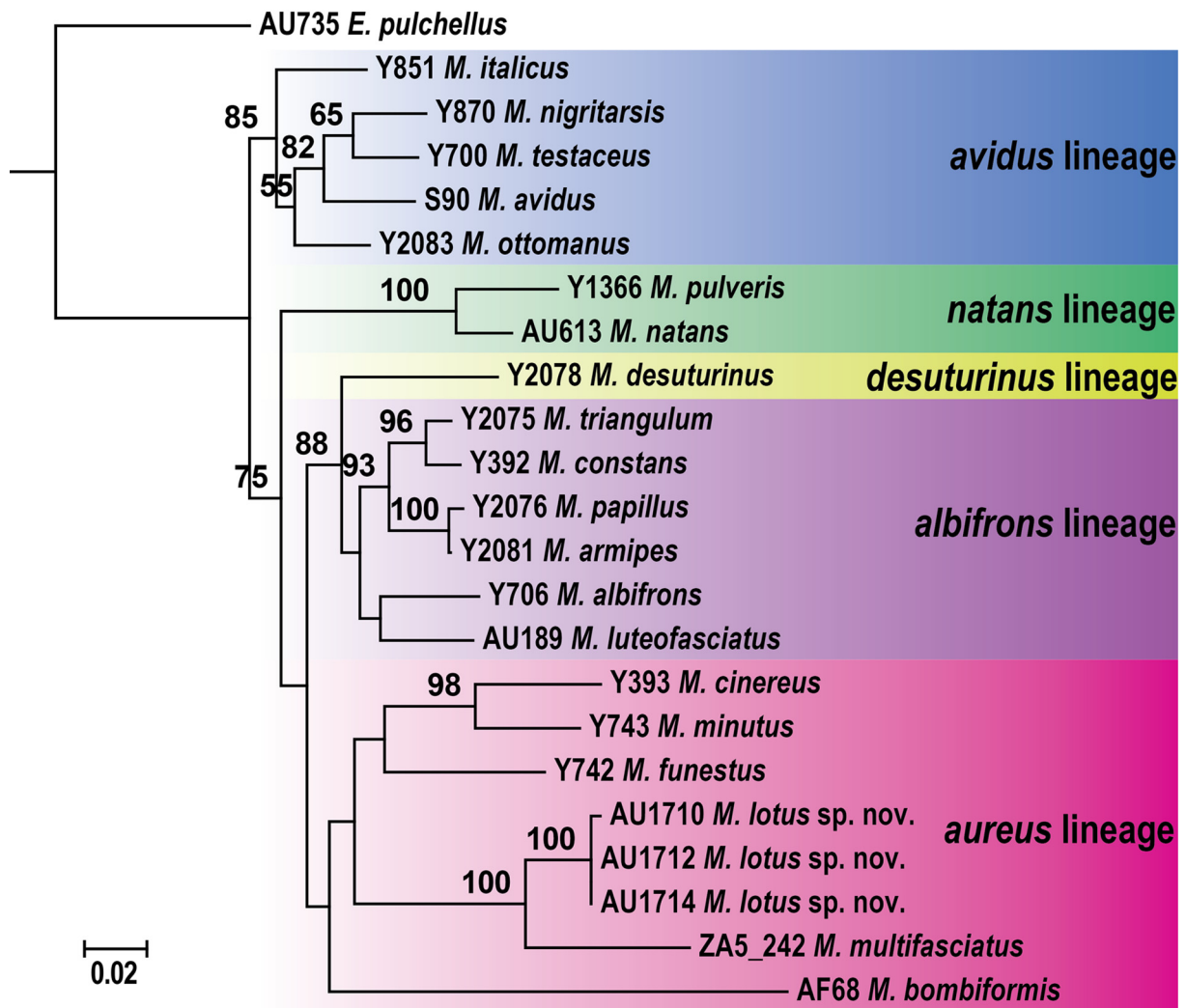


Fig. 27. Maximum likelihood tree from the concatenated data set with COI, 28S and 18S rRNA gene sequences.

several distinct characteristics (reddish-yellow face and frons; reduced mouth parts; antennal segments very short) and *M. lotus* sp. nov., with the posterior half of the scutum and the terga lacking pollinosity. Three species, *M. multifasciatus*, *M. vittatus* sp. nov. and *M. zebra* sp. nov., are very similar in external morphology with small differences between their male genitalia and in the size of the distal tooth on the apicoventral triangular lamina of the metafemur, the pilosity of the metafemur, and the length of the eye contiguity. For three species of the *M. bombiformis* group, *M. bombiformis*, *M. lotus* sp. nov. and *M. multifasciatus*, we have molecular data showing they are clearly genetically separated with a high COI divergence. *Merodon bombiformis* is morphologically similar to *M. multifasciatus*, but genetically highly divergent.

The *Merodon bombiformis* group and *M. funestus* differ from other species and groups among the *aureus* lineage in two apomorphic characters, an elongate pedicel and small lateral sclerite of the aedeagus. Lateral sclerite of the aedeagus are absent in other groups and species belonging to the *M. aureus* lineage. Morphologically, the *M. bombiformis* group can be distinguished from *M. funestus* by the absence of a projection on the metatrochanter in the males and a less dentate apicoventral triangular lamina on the metafemur. Based on the general external morphology, two species of the *M. bombiformis* group, *Merodon lotus* sp. nov. and *M. nasicus*, show a clear resemblance to *M. funestus*. *Merodon lotus* sp. nov. and *M. multifasciatus* are resolved in one highly supported clade, while *M. funestus* has an uncertain position on the phylogenetic tree. This is an indication of the monophyly of the *M. bombiformis* group, supported by morphological data. But, additional species, representatives of different groups of the *aureus* lineage, require to be included in order to resolve the phylogeny of the whole lineage.

Merodon bombiformis possesses some distinct apomorphic characters, such as a reduced mouthpart, reddish-yellow face and frons, convex frons, pedicel with strong setulae, basoflagellomere short, rounded, with a medial fossette. Reduced mouthparts (oral margin) covered by microtrichia are also present in the *M. planifacies* subgroup (Vujić *et al.* 2018; Djan *et al.* 2020). During 32 fieldwork days in the Drakensberg Mountains, spread over seven years of research conducted in RSA, members of our team collected 51, and observed twice as many, specimens of the *M. planifacies* subgroup (Djan *et al.* 2020).

There were no observed flower visits by any of the adult specimens during all of these fieldwork days. Adults do not visit flowers because they do not feed due to an adult life strategy adapted to fasting, and therefore they do not need well-developed mouthparts (Vujić pers. obs.). We can suppose that the same situation exists in *M. bombiformis*. We collected 11 specimens from the *M. bombiformis* group during our fieldwork. Two specimens of *M. multifasciatus* were recorded on flowers of *Nidorella auriculata*; all of the eight specimens of *M. lotus* sp. nov. were on flowers of *Coleonema* sp. By contrast, a single specimen of *M. bombiformis* was recorded sitting on the leaves of bushes.

Placement of the *Merodon bombiformis* group in the *Merodon* phylogeny

As stated, previous studies on the phylogeny of *Merodon* resolve five evolutionary lineages: *albifrons*, *aureus*, *avidus-nigratarsis*, *desuturinus*, and *natans* (Vujić *et al.* 2019a). The results of all of these studies were based on analysis of COI and 28S rRNA gene sequences (Mengual *et al.* 2006; Vujić *et al.* 2012; Radenković *et al.* 2018), while a new study on the phylogeny of *Merodon* based on a combined analysis of molecular and morphological data is currently under preparation (Veselić in prep). In this paper, we present a preliminary phylogeny of *Merodon* based on combined COI, 28S rRNA and 18S rRNA gene sequences with the main goal to resolve the *M. bombiformis* group relationship in the *aureus* lineage. We confirmed the monophyly of the five *Merodon* lineages, although the *aureus* and *albifrons* lineages both have low bootstrap support (<50), possibly as a result of a limited number of species per lineage included in the analysis. The *M. bombiformis* group is resolved within the *aureus* lineage as paraphyletic. However, due to a limited number of specimens included in the analysis, our results must be interpreted

with caution. Thus, more comprehensive analysis which will include additional specimens to resolve the phylogeny of the *M. bombiformis* group with more certainty.

Distribution and diversity

The *Merodon bombiformis* group comprises only six species with an extremely low number of collected specimens. Before we started our research, only 42 specimens of the *M. bombiformis* group had been detected in all available collections (26 males and 16 females). The initial aim of our research in the Republic of South Africa (RSA) was to collect fresh specimens of *Merodon bombiformis* for molecular analysis in order to understand the position of this taxon in the phylogenetic tree of the *M. aureus* lineage. During eight expeditions, together providing a total of 235 field days (2011–2018), 187 specimens of *Merodon* were collected, only 11 of them belonging to the *M. bombiformis* group. The other 176 specimens belong to the *M. desuturinus* lineage, and have partly been reported on in recent publications (Radenković *et al.* 2018; Djan *et al.* 2020). We found specimens of all three species of the *M. bombiformis* group known from RSA and these were used in the molecular analysis presented here. The single specimen of *M. bombiformis* was collected in 2016 after 108 collecting days, on Naudes Nek in the Drakensberg Mountains. We decided to continue research and focus on two other taxa of the *M. bombiformis* group previously detected in RSA, to obtain further molecular data. The other previously described species, *M. multifasciatus*, was recorded on day 215 in Mpumalanga Province, near Emgwenya (Waterval Boven). The third species was, until our research, undescribed and represented by a single specimen in the BMNH collections. This specimen was collected in 1920, from Ceres. We revisited Ceres area 18 times and at last found a population of the species, described here as *M. lotus* sp. nov., on day 230 of our fieldwork. Molecular data obtained for *M. multifasciatus* and *M. lotus* sp. nov. were resolved in a separate clade within the *aureus* lineage. The divergence between them supported the proposition that there has been a high genetic divergence among species in the *M. bombiformis* group, even though our analysis included only 2 and 3 specimens, respectively. We found that it required huge effort to observe, find and collect specimens of *Merodon* during our fieldwork in RSA. Comparing collection of specimens of *Merodon* in the Afrotropical region and the Mediterranean, an almost incredible disproportion is observed. The total number of 176 *Merodon* specimens from 10 species, collected during 235 days of fieldwork in RSA, can be reached in only one day's fieldwork during peak flying season, in species rich Mediterranean localities like Agiassos Mountain on Lesbos Island, Greece (Vujić *et al.* 2020).

Radenković *et al.* (2018) discussed the link between Afrotropical and Mediterranean species of *Merodon* and their host plants. Larvae of *Merodon* develop in the underground bulbs, corms, or rhizomes of geophytes (Amaryllidaceae, Iridaceae and Hyacinthaceae) or in the surrounding soil, based on the data for species with known immature stages (Hurkmans 1993; Andrić *et al.* 2014; Ricarte *et al.* 2017; Preradović *et al.* 2018). In the Afrotropical region, larvae of the *M. planifacies* subgroup species have been recorded in bulbs of *Merwillia Speta* (family Hyacinthaceae) (Radenković *et al.* 2018) and larvae of *M. bombiformis* group species in corms of *Gladiolus* (Iridaceae). The family Hyacinthaceae comprises between 700 and 900 species, mainly distributed in Africa, Europe and SW Asia (Martínez-Azorín *et al.* 2011) and in the family Iridaceae Juss. there are more than 2200 species worldwide (Christenhusz & Byng 2016). The subfamily Crocoideae Burnett, to which *Gladiolus* belongs, comprises about 995 species, with most of the species in Africa, Madagascar and Eurasia, but with the highest diversity in South Africa (Goldblatt *et al.* 2006). These distributional patterns suggest that diversification of these taxa began when North America was already clearly separated from Eurasia. The absence of the genus *Merodon* on the American continent can thus be linked to historical events in the Hyacinthaceae and Iridaceae. However, the greatest diversity of Hyacinthaceae and Crocoideae is found in South Africa (Pfosser & Speta 2004; Goldblatt *et al.* 2006), which is not in congruence with the exceptionally high diversity of *Merodon* in the Eastern Mediterranean (more than 100 species) as opposed to South Africa (11 species) (Radenković *et al.* 2018; Djan *et al.* 2020). Migration of bulb species from southern Africa

to northern Africa and Eurasia was possible during the late Early Miocene, giving rise to free exchange of flora and fauna between Africa and Eurasia (Rögl 1998; Pfosser & Speta 2004). Radenković *et al.* (2018) postulated that diversification in the *M. desuturinus* group most probably happened much later, during fundamental shifts in the African climate, likewise the diversification in the *M. bombiformis* group. Generally, the genus *Merodon* prefers warm, dry, open habitats with numerous geophytes. Radenković *et al.* (2018) supposed that during the Pliocene-Pleistocene epoch, favorable conditions for species of *Merodon* (increased aridity and open grasslands) in Africa (De Menocal 2004) most probably allowed faunal transitions. In the case of the *M. desuturinus* and *M. bombiformis* groups, this occurred in the opposite direction to the northward trajectory of the Hyacinthaceae; from the eastern Mediterranean (including SW Asia), one lineage migrated to South Africa and another to the western Palaearctic.

Except for *Merodon vittatus* sp. nov., reported from Ghana in Western Africa, all other species of the *M. bombiformis* group are known from the southern or eastern parts of Africa. *Merodon vittatus* sp. nov. occurs in a deciduous forest-woodland savanna. *Merodon nasicus* is recorded in Eritrea, Ethiopia, Burundi and Kenya, at high altitudes (2000–2500 m a.s.l.). All of these records originate from localities belonging to the Eastern Afrotropical Biodiversity Hotspot, which includes several widely scattered, but biogeographically similar mountain ranges in Eastern Africa (Critical Ecosystem Partnership Fund 1996), with vegetation of montane forest-tundra (Sayre *et al.* 2013). This chain of localities in East Africa supporting representatives of the *M. bombiformis* group continues toward the south with three species which alternate and partly overlap in their ranges. *Merodon zebra* sp. nov. is recorded from Congo and Tanzania at moderate elevations (500–1700 m a.s.l.), in deciduous forest-woodland savanna. *Merodon multifasciatus* occurs in RSA, Malawi and Congo, at the same range of elevations as *M. zebra* sp. nov. (500–1800 m a.s.l.), in both deciduous forest-woodland savanna and brush-grass savanna. The range of *M. zebra* sp. nov. and *M. multifasciatus* overlap in Congo. *Merodon bombiformis* occurs in RSA and Lesotho over a range of moderate to high altitudes (800–2500 m a.s.l.), in different types of vegetation, including brush-grass savanna, temperate and mountain grasslands and montane forest-tundra (Sayre *et al.* 2013). *Merodon bombiformis* and *M. multifasciatus* have a common range in brush-grass savanna in the north of RSA. The localised endemic *M. lotus* sp. nov. appears in Mediterranean evergreen forest-sclerophyllous shrub in the extreme south of Africa.

All habitats in RSA where we collected specimens from the *Merodon bombiformis* group are either outside protected areas, or in protected areas with high pressure of human impact, or at sites both unprotected and heavily impacted by human activity. *Merodon bombiformis* was found on a small patch of natural habitat (0.7 ha) on the slopes of the Drakensberg Mountains far from protected areas, on the east and north of the mountain range. *Merodon multifasciatus* was recorded on the edge of the city (Emgwenya (Waterwal Boven)) with high human impact and a very small remaining population of the assumed host plant. This situation is similar to that of the habitat of *M. lotus* sp. nov. (0.5 ha), which belongs to Ceres Bergfynbos Reserve (BIOPAMA 2011), but subject to everyday human influence, being on the edge of city infrastructure. Despite our extremely intense collecting efforts in more than 50 localities in the RSA, most of them in protected areas, all specimens found were outside well-preserved National Parks and Reserves. Conservation efforts to protect these small populations of the extremely rare species of the *M. bombiformis* group thus need to be concentrated on the remaining small patches of natural habitats supporting their host plants. Specimens of the genus *Gladiolus* are clearly visible during the flowering season and very often collected by the local communities.

Host plants

The data gathered from both the literature (Stuckenberg 1956) and our collecting experiences point to *Gladiolus* L. (Iridaceae) as the larval host plant for species of *Merodon* of the *bombiformis* group (see under larval biology in *M. bombiformis* and *M. multifasciatus*). Our field observations from Emgwenya (Waterwal Boven), where we collected adults of *M. multifasciatus* within a habitat containing a natural

population of *Gladiolus sericeovillosus* suggests this plant species is the primary natural host plant. Data about distribution of two subspecies of *G. sericeovillosus* in South Africa show a broad correspondence with the distribution of these two species of *Merodon*. *Gladiolus sericeovillosus* subsp. *sericeovillosus*, the southern subspecies, is especially common in KwaZulu-Natal (Goldblatt & Manning 1998), from where most known specimens *M. bombiformis* originated. The range of this *Gladiolus* extends from Baziya in the northern Eastern Cape to southern Mpumalanga, along a line extending from Ermelo to Barberton (Goldblatt & Manning 1998). The second subspecies *G. sericeovillosus* subsp. *calvatus* (Baker) Goldblatt, extends within the RSA from Mpumalanga through Northern Province into Zimbabwe, where it occurs widely across the eastern half of the country (Goldblatt & Manning 1998). This subspecies was also detected in Emgwenya (Waterval Boven), where we collected *M. multifasciatus*. If we conclude from the distribution data that *M. bombiformis* is connected with *G. sericeovillosus* subsp. *sericeovillosus* and *M. multifasciatus* with *G. sericeovillosus* subsp. *calvatus*, then we can predict the occurrence of the latter plant subspecies in Malawi and Congo. A second inference can be drawn from the line from Ermelo to Barberton, which divides these two subspecies. Along this line there are three records of *M. bombiformis* (Pretoria, Pietermaritzburg and Waterval) and three of *M. multifasciatus* (Barberton, Nylstroom and Emgwenya (Waterval Boven)). It would be interesting if botanists were to check the taxonomic status of these subspecies: *G. sericeovillosus* subsp. *sericeovillosus* and *G. sericeovillosus* subsp. *calvatus*. The entomological data provide reasons to believe that they are distinct species-level taxa.

There are a lot of endemic species of the genus *Gladiolus* in the Ceres area, from where *Merodon lotus* sp. nov. was described. Examples are *G. carneus* de la Roche, *G. ceresianus* Bolus, *G. cylindraceus* Lewis, all of them Western Cape endemics (Biodiversity Explorer 2000). Further research is needed to determine more precisely the host plant(s) of this localised endemic syrphid species.

Acknowledgements

We thank the curators of the museums listed in the Materials and methods for facilitating visits and loans for the study of specimens in their care. We are also indebted to Martin Speight, native English speaker for the linguistic revision, valuable comments and editing of the manuscript. We are sincerely grateful to Dr Nigel Wyatt (NHM) for the photos of the type specimen of *Merodon nasicus*. We wish to thank Neil Crouch and Troos Van Der Merwe for their help with the identification of plant material. The study was supported by the European Commission, Horizon 2020 Marie Skłodowska-Curie action, Research and Innovation Staff Exchange (RISE) Programme: FlyHigh-Insect-plant relationships: insight into biodiversity and new applications (Grant No. 645636), the Ministry of Education, Science and Technological Development, Republic of Serbia, Grant No. 173002, III43002, 451–03–9/2021–14/200358 and 451–03–9/2021–14/200125, the Provincial Secretariat for Science and Technological Development of the Republic of Serbia (Grant No. 114–451–1125/2014–03 and 114–451–1702/2014–03) and European Commission, Horizon 2020 project ‘ANTARES’ (Grant No. 739570).

References

- Andrić A., Šikoparija B., Obreht D., Đan M., Preradović J., Radenković S., Pérez-Bañón C. & Vujić A. 2014. DNA barcoding applied: identification of the larva of *Merodon avidus* (Diptera: Syrphidae). *Acta Entomologica Musei Nationalis Pragae* 54: 741–757.
- Belshaw R., Lopez-Vaamonde C., Degerli N. & Quicke D.L. 2001. Paraphyletic taxa and taxonomic chaining: evaluating the classification of braconine wasps (Hymenoptera: Braconidae) using 28S D2-3 rDNA sequences and morphological characters. *Biological Journal of the Linnean Society* 73: 411–424. <https://doi.org/10.1111/j.1095-8312.2001.tb01370.x>

- Bezzi M. 1915. *The Syrphidae of the Ethiopian Region based on Material in the Collection of the British Museum (Natural History) with Descriptions of new Genera and Species*. British Museum (Natural History), London. <https://doi.org/10.5962/bhl.title.10351>
- Bezzi M. 1921. Syrphidae aethiopicae Musei nationalis hungarici [concl.]. *Brotéria: Série Zoológica* 19: 5–22.
- Biodiversity Explorer. 2000. Genus: *Gladiolus*. Available from <https://www.biodiversityexplorer.info/plants/iridaceae/gladiolus.htm> [accessed 20 Jul. 2020].
- BIOPAMA. 2011. Ceres Bergfynbos Reserve. Available from <https://tris.biopama.org/pa/300356> [accessed 10 Jul. 2020].
- Chen H., Rangasamy M., Tan S.Y., Wang H. & Siegfried B.D. 2010. Evaluation of five methods for total DNA extraction from western corn rootworm beetles. *PLoS ONE* 5: e11963. <https://doi.org/10.1371/journal.pone.0011963>
- Christenhusz M.J.M. & Byng J.W. 2016. The number of known plants species in the world and its annual increase. *Phytotaxa* 261 (3): 201–217. <https://doi.org/10.11646/phytotaxa.261.3>
- Critical Ecosystem Partnership Fund. 1996. Eastern Afromontane. Available from <https://www.cepf.net/our-work/biodiversity-hotspots/eastern-afromontane> [accessed 1 Jul. 2020].
- Curran C.H. 1939. Records and descriptions of African Syrphidae (Diptera). *III American Museum Novitates* 1025: 1–11.
- De Menocal P. 2004. African climate change and faunal evolution during the Pliocene-Pleistocene. *Earth and Planetary Science Letters* 220: 3–24. [https://doi.org/10.1016/S0012-821X\(04\)00003-2](https://doi.org/10.1016/S0012-821X(04)00003-2)
- Dirickx H.G. 1998. *Catalogue synonymique et géographique des Syrphidae (Diptera) de la Région Afrotropicale*. Vol. 2. Muséum d'histoire naturelle, Ville de Genève, Suisse.
- Djan M., Ståhls G., Veličković N., Ačanski J., Vidaković D.O., Rojo S., Pérez-Bañón S., Radenković S. & Vujić A. 2020. The *Merodon planifacies* subgroup (Diptera: Syrphidae): congruence of molecular and morphometric evidences reveal new taxa in Drakensberg mountains valleys (Republic of South Africa). *Zoologischer Anzeiger* 287: 105–120. <https://doi.org/10.1016/j.jcz.2020.05.010>
- Doczkal D. & Pape T. 2009. *Lyneborgimyia magnifica* gen. et sp. n. (Diptera: Syrphidae) from Tanzania, with a phylogenetic analysis of the Eumerini using new morphological characters. *Systematic Entomology* 34 (3): 559–573. <https://doi.org/10.1111/j.1365-3113.2009.00478.x>
- Evenhuis N.L. & Pape T. 2020. Systema Dipteriorum, Version 2.6. Available from <http://diptera.org/> [accessed 2 Jun. 2020].
- Folmer O., Black M., Hoeh W., Lutz R. & Vrijenhoek R. 1994 DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3: 294–299.
- Goldblatt P. & Manning J. 1998. *Gladiolus in Southern Africa*. Fernwood Press, Cape Town.
- Goldblatt P., Davies J., Manning J., van der Bank M. & Savolainen V. 2006. Phylogeny of Iridaceae Subfamily Crocoideae based on a combined multigene plastid DNA analysis. *Aliso* 22 (1): 399–411. <https://doi.org/10.5642/aliso.20062201.32>
- Hadley A. 2006. CombineZ, Ver. 5. Published by the author.
- Hijmans R.J., Guarino L. & Mathur P. 2012. A geographic information system for the analysis of species distribution data. Ver. 7.5. LizardTech, Inc., University of California.

- Hall T.A. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95–98.
- Hull F.M. 1944. Some flies of the Family Syrphidae in the British Museum (Natural History). *Annals and Magazine of Natural History Series 11* 11 (73): 21–66. <https://doi.org/10.1080/00222934408527401>
- Hurkmans W. 1993. A monograph of *Merodon* (Diptera: Syrphidae). Part 1. *Tijdschrift voor Entomologie* 136: 147–234.
- International Commission on Zoological Nomenclature. 1999. *International Code of Zoological Nomenclature* (4th edition) [Incorporating Declaration 44, amendments of Article 74.7.3, with effect from 31 December 1999 and the Amendment on e-publication, amendments to Articles 8, 9, 10, 21 and 78, with effect from 1 January 2012]. International Trust for Zoological Nomenclature, London. Available from <https://www.iczn.org/the-code-online/> [accessed 17 Jul. 2020].
- Katoh K. & Standley D.M. 2013. MAFFT multiple sequence alignment software. Ver.7: improvements in performance and usability. *Molecular Biology and Evolution* 30: 772–780. <https://doi.org/10.1093/molbev/mst010>
- Kumar S., Stecher G. & Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis version 7.0 for biggest datasets. *Molecular Biology and Evolution* 33: 1870–1874. <https://doi.org/10.1093/molbev/msw054>
- Macquart J. 1855. Diptères exotiques nouveaux ou peu connus. 5^e supplément. *Memoires de la Société (royale) des Sciences, de l'Agriculture et des Arts à Lille* 1 (2): 25–156. Also published separately as his “*Diptères exotiques nouveaux ou peu connus. Supplément V*”: 5–136.
- Marcos-García M.Á., Vujić A. & Mengual X. 2007. Revision of Iberian species of the genus *Merodon* (Diptera: Syrphidae). *European Journal of Entomology* 104: 531–572. <https://doi.org/10.14411/eje.2007.073>
- Martínez-Azorín M., Crespo M.B., Juan A. & Fay M.F. 2011. Molecular phylogenetics of subfamily Ornithogaloidae (Hyacinthaceae) based on nuclear and plastid DNA regions, including a new taxonomic arrangement. *Annals of Botany* 107 (1): 1–37. <https://doi.org/10.1093/aob/mcq207>
- McAlpine J.F. 1981. Morphology and Terminology – Adults. In: McAlpine J.F., Peterson B.V., Shewell G.E., Teskey H.J., Vockeroth J.R. & Wood D.M. (eds) *Manual of Nearctic Diptera. Vol. 1*: 9–63. Monograph No. 27, Research Branch, Agriculture Canada, Ottawa.
- McWilliam H., Li W., Uludag M., Squizzato S., Mi Park Y., Buso N., Cowley A.P. & Lopez R. 2013. Analysis tool web services from the EMBL-EBI. *Nucleic Acids Research* 41: W597–W600. <https://doi.org/10.1093/nar/gkt376>
- Mengual X., Ståhls G., Vujić A. & Marcos-García M.A. 2006. Integrative taxonomy of Iberian *Merodon* species (Diptera, Syrphidae). *Zootaxa* 1377 (1): 1–26. <https://doi.org/10.11646/zootaxa.1377.1.1>
- Miller M.A., Pfeiffer W. & Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. *2010 Gateway Computing Environments Workshop (GCE)*: 1–8. New Orleans, LA. <https://doi.org/10.1109/GCE.2010.5676129>
- Pfossor M.F. & Speta F. 2004. From *Scilla* to *Charibdis*-is our voyage safer now? *Plant Systematics and Evolution* 246: 245–263.
- Preradović J., Andrić A., Radenković S., Šašić Zorić Lj., Pérez-Bañón C., Campoy A. & Vujić A. 2018. Pupal stages of three species of the phytophagous genus *Merodon* Meigen (Diptera: Syrphidae). *Zootaxa* 4420 (2): 229–242. <https://doi.org/10.11646/zootaxa.4420.2.5>

- Radenković S., Veličković N., Ssymank A., Obreht Vidaković D., Djan M., Ståhls G., Veselić S. & Vujić A. 2018. Close relatives of Mediterranean endemo-relict hoverflies (Diptera, Syrphidae) in South Africa: Morphological and molecular evidence in the *Merodon melanocerus* subgroup. *PLoS ONE* 13 (7): e0200805. <https://doi.org/10.1371/journal.pone.0200805>
- Ricarte A., Marcos-García M.Á. & Rotheray G.E. 2008. The early stages and life histories of three *Eumerus* and two *Merodon* species (Diptera: Syrphidae) from the Mediterranean region. *Entomologica Fennica* 19: 19–141. <https://doi.org/10.33338/ef.84424>
- Ricarte A., Souba-Dols G.J., Hauser M. & Marcos-García M.Á. 2017. A review of the early stages and host plants of the genera *Eumerus* and *Merodon* (Diptera: Syrphidae), with new data on four species. *PLoS ONE* 12 (12): e0189852. <https://doi.org/10.1371/journal.pone.0189852>
- Rodríguez F., Oliver J.L., Marín A. & Medina J.R. 1990. The general stochastic model of nucleotide substitution. *Journal of Theoretical Biology* 142: 485–501. [https://doi.org/10.1016/S0022-5193\(05\)80104-3](https://doi.org/10.1016/S0022-5193(05)80104-3)
- Rögl F. 1998. Paleogeographic consideration for Mediterranean and Paratethys seaways (Oligocene to Miocene). *Annalen des Naturhistorischen Museums in Wien* 99A: 279–310.
- Šašić Lj., Ačanski J., Vujić A., Ståhls G., Radenković S., Milić D., Obreht-Vidaković D. & Đan, M. 2016. Molecular and morphological inference of three cryptic species within the *Merodon aureus* species group (Diptera: Syrphidae). *PLoS ONE* 11: e0160001. <https://doi.org/10.1371/journal.pone.0160001>
- Sayre R., Comer P., Hak J., Josse C., Bow J., Warner H., Larwanou M., Kelbessa E., Bekele T., Kehl H., Amena R., Andriamasimanana R., Ba T., Benson L., Boucher T., Brown M., Cress J., Dassering O., Friesen B., Gachathi F., Houcine S., Keita M., Khamala E., Marangu D., Mokuia F., Morou B., Mucina L., Mugisha S., Mwavu E., Rutherford M., Sanou P., Syampungani S., Tomor B., Vall A., Vande Weghe J., Wangui E., & Waruingi L. 2013. *A New Map of Standardized Terrestrial Ecosystems of Africa*. Washington, DC: Association of American Geographers. Available from http://www.aag.org/cs/publications/special/map_african_ecosystems [accessed 24 May 2021].
- Smith K.G.V. & Vockeroth J.R. 1980. Family Syrphidae. In: Crosskey R.W. (ed.) *Catalogue of the Diptera of the Afrotropical Region*: 488–510. British Museum, London.
- South African National Biodiversity Institute. 2004. *Leucosidea sericea*. Available from <http://pza.sanbi.org/leucosidea-sericea> [accessed 21 Jun. 2020].
- Speight M.C.D. 2020. Species accounts of European Syrphidae, 2020. *Syrph the Net, the database of European Syrphidae (Diptera)* vol. 104. Syrph the Net publications, Dublin.
- Ståhls G., Vujić A., Pérez-Bañón C., Radenković S., Rojo S. & Petanidou T. 2009. COI barcodes for identification of *Merodon* hoverflies (Diptera, Syrphidae) of Lesvos Island, Greece. *Molecular Ecology Resources* 9 (6): 1431–1438. <https://doi.org/10.1111/j.1755-0998.2009.02592.x>
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30: 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- Stuckenberg B.R. 1956. The immature stages of *Merodon bombiformis* Hull, a potential pest of bulbs in South Africa. (Diptera: Syrphidae). *Journal of the Entomological Society of Southern Africa* 19 (2): 219–224.
- Thompson F.C. 1988. Syrphidae (Diptera) described from unknown localities. *Journal of the New York Entomological Society* 96 (2): 200–226.
- Thompson F.C. 1999. A key to the genera of the flower flies (Diptera: Syrphidae) of the Neotropical Region including descriptions of new genera and species and a glossary of taxonomic terms. *Contributions on Entomology, International* 3: 321–378.

Vujić A., Radenković S., Ståhls G., Ačanski J., Stefanović A., Veselić S., Andrić A. & Hayat R. 2012. Systematics and taxonomy of the *ruficornis* group of genus *Merodon* Meigen (Diptera: Syrphidae). *Systematic Entomology* 37: 578–602. <https://doi.org/10.1111/j.1365-3113.2012.00631.x>

Vujić A., Radenković S. & Likov L. 2018. Revision of the Palaearctic species of the *Merodon desuturinus* group (Diptera, Syrphidae). *ZooKeys* 771: 105–138. <https://doi.org/10.3897/zookeys.771.20481>

Vujić A., Radenković S., Likov L., Andrić A., Gilasian E. & Barkalov A. 2019a. Two new enigmatic species of the genus *Merodon* Meigen (Diptera: Syrphidae) from the north-eastern Middle East. *Zootaxa* 4555 (2): 187–208. <https://doi.org/10.11646/zootaxa.4555.2.2>

Vujić A., Ståhls G. & Radenković S. 2019b. Hidden European diversity: a new monotypic hoverfly genus (Diptera: Syrphidae: Eristalinae: Rhingiini). *Zoological Journal of the Linnean Society* 185 (4): 1188–1211. <https://doi.org/10.1093/zoolinlean/zly066>

Vujić A., Speight M., de Courcy Williams M., Rojo S., Ståhls G., Radenković S., Likov L., Miličić M., Pérez-Bañón C., Falk S. & Petanidou T. 2020. *Atlas of the Hoverflies of Greece*. Brill, Leiden, The Netherlands. <https://doi.org/10.1163/9789004334670>

Manuscript received: 6 November 2020

Manuscript accepted: 12 April 2021

Published on: 21 June 2021

Topic editor: Nesrine Akkari

Section editor: Torbjørn Ekrem

Desk editor: Pepe Fernández

Printed versions of all papers are also deposited in the libraries of the institutes that are members of the *EJT* consortium: Muséum national d'histoire naturelle, Paris, France; Meise Botanic Garden, Belgium; Royal Museum for Central Africa, Tervuren, Belgium; Royal Belgian Institute of Natural Sciences, Brussels, Belgium; Natural History Museum of Denmark, Copenhagen, Denmark; Naturalis Biodiversity Center, Leiden, the Netherlands; Museo Nacional de Ciencias Naturales-CSIC, Madrid, Spain; Real Jardín Botánico de Madrid CSIC, Spain; Zoological Research Museum Alexander Koenig, Bonn, Germany; National Museum, Prague, Czech Republic.

Supplementary material

Supp. file 1: Data related to the studied specimens and the corresponding GenBank accession numbers. <https://doi.org/10.5852/ejt.2021.755.1401.4469>