

Research article

urn:lsid:zoobank.org:pub:C4F57584-0AD9-47C9-BA95-02133C434941

**Integrative description of a new Tunisian tardigrade species,
Macrobiotus azzunae sp. nov. (Eutardigrada,
Macrobiotidae, *hufelandi* group)**Jamila BEN MARNISSI ¹, Michele CESARI ²,
Lorena REBECCHI ^{3,*} & Roberto BERTOLANI ^{4,5}¹Department of Biological Sciences, Faculty of Mathematical, Physical and Natural Sciences of Tunis,
University of Tunis El Manar, 2092 El Manar, Tunis, Tunisia.^{2,3}Department of Life Sciences, University of Modena and Reggio Emilia,
Via G. Campi 213/D, 41125, Modena, Italy.⁴Department of Education and Humanities, University of Modena and Reggio Emilia,
Via A. Allegri 9, 42121 Reggio Emilia, Italy.⁵Museo Civico di Storia Naturale, Lungadige Porta Vittoria, 9, 37129 Verona, Italy.

* Corresponding author: lorena.rebecchi@unimore.it

¹ Email: jamila.marnissi@fst.utm.tn² Email: michele.cesari@unimore.it⁴ Email: roberto.bertolani@unimore.it¹urn:lsid:zoobank.org:author:F10DD853-D540-4984-A16A-5D5EC377E4C0²urn:lsid:zoobank.org:author:87D45951-C71A-48C1-92A1-C07387A80284³urn:lsid:zoobank.org:author:5A02A091-C92C-44A3-B7C4-672B9D27B590⁴urn:lsid:zoobank.org:author:A7381EA2-93C7-4127-9460-2640581E48B6

Abstract. In this paper a new tardigrade species, *Macrobiotus azzunae* sp. nov., from Tunisia, is described. An integrative taxonomic approach was applied by combining morphological, morphometric and molecular data. In particular, light and scanning electron microscopy observations, and four genetic markers, three nuclear (18S rRNA, 28S rRNA and ITS-2) and one mitochondrial (COI) were used. The analysis showed that *M. azzunae* sp. nov. belongs to the *Macrobiotus hufelandi* group and is most similar to *Macrobiotus sandrae* Bertolani & Rebecchi, 1993. It differs from *M. sandrae* by a more pronounced constriction of the first macroplocoid (hardly visible in *M. sandrae*) and for the eggshell shape, with thinner wires of the reticulum and meshes around the processes larger than the inter-process meshes in *M. azzunae* sp. nov., while all meshes are similar in size in *M. sandrae*. The species is gonochoristic. With this discovery, there are 33 species of tardigrades identified in Tunisia, all non-marine. This result, compared with nearby Sicily, where more research has been conducted, indicates that there is a considerable potential for identification of new species. Further research will be most informative if multiple habitats are explored and if carried out with an integrated approach as done in this present work.

Keywords. Tardigrada, Ain Soltan Forest, Tunisia, *Macrobiotus azzunae*, new species.

Ben Marnissi J., Cesari M., Rebecchi L. & Bertolani R. 2021. Integrative description of a new Tunisian tardigrade species, *Macrobotus azzunae* sp. nov. (Eutardigrada, Macrobotidae, *hufelandi* group). *European Journal of Taxonomy* 758: 122–146. <https://doi.org/10.5852/ejt.2021.758.1429>

Introduction

Tardigrades are hygrophilous micrometazoans whose outstanding resistance enables most of them to inhabit a large variety of habitats from the greatest depths of oceans to the highest mountain peaks, as well as extreme environments such as cryoconite holes. In terrestrial environments they colonize mosses, lichens, leaf litter and soil which receive stochastic hydration (Nelson *et al.* 2015; Schill 2018). Much research on the diversity and distribution of tardigrade fauna has been carried out in recent years in various parts of the world, especially considering the terrestrial environment. This has led to a significant increase in the number of known species: 531 in 1983 (Ramazzotti & Maucci 1983), about 960 in 2005 (Guidetti & Bertolani 2005) and more than 1300 species in 2020 (Degma *et al.* 2020). Nevertheless, knowledge of the diversity and distribution of North African tardigrades is still very limited and in particular in Tunisia, for which only three studies have been conducted. The first report on Tunisian tardigrades comes from a survey by Iharos (1978) in Northern Tunisian regions in which eleven species of tardigrades were detected. Then, Binda & Pilato (1987) studied the tardigrade fauna of Salambo (Tunis), Tabarka and Ain Drahem (Jendouba), and recently Gąsiorek *et al.* (2017) studied the tardigrade fauna of Bni Mtir, Jendouba. Currently, the known Tunisian tardigrade fauna includes 32 species, of which one, *Bryodelphax maculatus* Gąsiorek, Stec, Morek, Marnissi, Michalczyk, 2017, was originally discovered from the same region considered here, namely the Kroumirie Mountains. The Kroumirie Mountains are located in Northern Tunisia near the Algerian boundary and are characterized by a climate which varies from sub-humid to humid in winter. The Kroumirie forests are covered especially by cork oaks (*Quercus suber* L.) and Algerian oaks (*Quercus canariensis* Willd.) (Stambouli-Essassi *et al.* 2007). The analysis of the tardigrade fauna of a moss collected at the Kroumirie Mountains led us to find a new eutardigrade species (*Macrobotus azzunae* sp. nov.) belonging to the *Macrobotus hufelandi* group, considered as one of the most common groups of limnoterrestrial tardigrades on the planet (McInnes 1994; Kaczmarek & Michalczyk 2017a; McInnes *et al.* 2017). In this study we combined classical morphological and morphometric methods with modern molecular techniques in an integrative approach as suggested by Cesari *et al.* (2009, 2011, 2020), Stec *et al.* (2018b) and Kayastha *et al.* (2020). Using phase contrast (PhC) light microscopy (LM), differential interference contrast (DIC) and scanning electron microscopy (SEM), we were able to describe the phenotypic characteristics of the new species whereas the amplification of DNA markers (three nuclear, 18S rRNA, 28S rRNA and ITS-2, and one mitochondrial, COI) provided barcodes for the genetic identification of this new species.

Material and methods

Tardigrade collection

The moss containing the new species was collected in April 2017 by the first author from Ain Soltan forest in the North West of Tunisia (Fig. 1) on a trunk of an Algerian oak tree. The sample was stored dry in a labeled paper bag and then sent to the Laboratory of Evolutionary Zoology at the Department of Life Sciences, University of Modena and Reggio Emilia (UNIMORE), Italy. In order to extract tardigrades from the sample, a small portion of the collected substrate was placed and maintained in tap water at room temperature (20°C) for about half an hour. The sample was then sieved to separate tardigrades and their eggs from the substrate, and both animals and eggs were isolated using a glass pipette under a stereo microscope.

Microscopy and imaging

The collected specimens (eggs and animals) were mounted on slides in Faure-Berlese fluid (permanent slides) or fixed in Carnoy fixative (methanol: acetic acid, 3:1) and in the latter case then stained with a drop of acetic-lactic orcein (Rebecchi & Bertolani 1988) for LM observations (not permanent slides).

Observations and measurements of the sclerified parts of animals and eggshells were carried out with LM under PhC and DIC up to the maximum magnification (100× oil objective) with a Leica DM RB microscope equipped with a Nikon DS-Fi 1 digital camera, at the Department of Life Sciences, UNIMORE. The species was identified by comparing the specimens with the original descriptions of similar species of *Macrobotus* (for references see Results: Differential diagnosis) and, when possible, with the original type material of some of those species. In particular, we have examined the following material from Maucci's collection (thanks to the Civic Museum of Natural History of Verona, Italy): slides CT14009 holotype and CT14015 egg of *Macrobotus madegassus* Maucci, 1993; slide CT14701 one animal and one egg (paratypes) of *Macrobotus personatus* Biserov, 1990; from Bertolani's collection of the Department of Life Sciences, UNIMORE, Italy: slide C460–S97 holotype of *Macrobotus sandrae* Bertolani & Rebecchi, 1993 and for the same species slides from the type locality C442–S79 (Animal) and C450–S01 and C2346–S2 (eggs). These specimens and eggs were analyzed under LM.

Measurements of animal length, claws and buccal-pharyngeal apparatus details were taken according to Pilato (1981). The external buccal tube diameter was measured at the level of the stylet support insertion point. We calculated the *pt* index which is the percentage ratio between the length of a structure and the length of the buccal tube measured from the medio-dorsal ridge of the buccal armature to the base of the pharyngeal apophyses (Pilato 1981). Single measurements are available in Supp. file 1.

Fifteen additional specimens were prepared for SEM analyses according to Bertolani *et al.* (2014), and observed with a Philips SEM XL 40, available at the Centro Interdipartimentale Grandi Strumenti (CIGS) of UNIMORE.

Molecular analysis

Before molecular analysis, each specimen was identified and photographed in vivo with LM using the method described by Cesari *et al.* (2011) in order to obtain voucher specimens. Supp. file 2 shows the buccal-pharyngeal apparatus of a hologenophore voucher specimen, showing how it is possible to obtain good morphological information on alive animals even when the entire animal has to be used

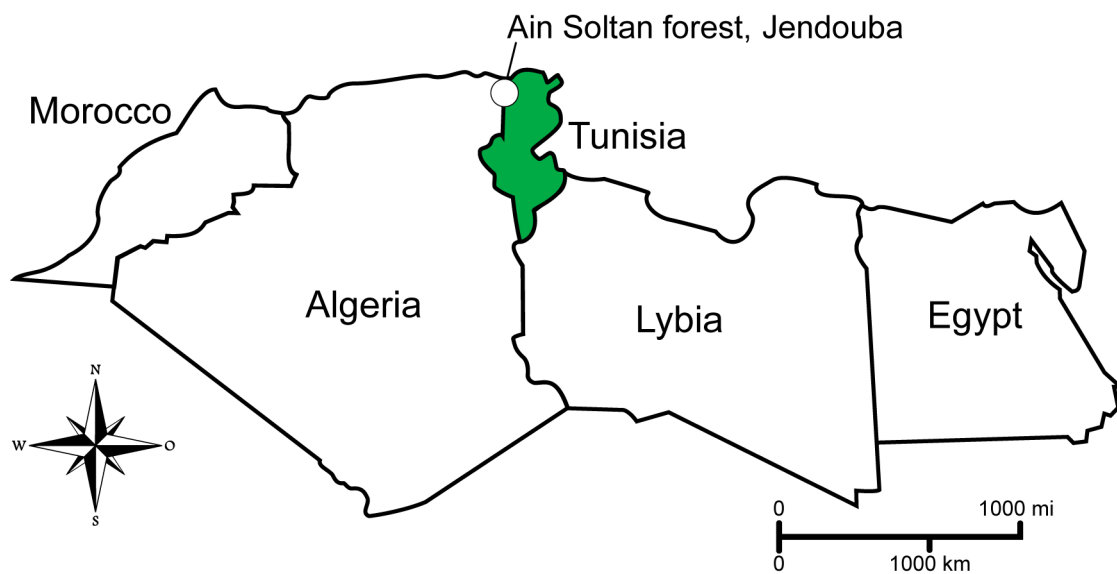


Fig. 1. Map of the sampling locality (circle) in the North-West of Tunisia.

Table 1. List of specimens of *M. azzunae* sp. nov. and *M. sandrae* Bertolani & Rebecchi, 1993 utilized for molecular analyses. Abbreviation: N/A = not available. * These specimens were analyzed for the COI gene in Bertolani *et al.* (2011b).

Specimen	18S	28S	ITS-2	COI
<i>Macrobiotus azzunae</i>				
C4218 V1	N/A	N/A	N/A	MW698695
C4218 V2	N/A	N/A	N/A	MW698696
C4218 V4	MW695447	MW695450	MW695454	MW698697
C4218 T5	MW695448	MW695451	MW695455	MW698698
C4218 T6	MW695449	MW695452	MW695456	N/A
C4218 V7	N/A	MW695453	MW695457	MW698699
<i>Macrobiotus sandrae</i>				
C2945 b	MW695445	N/A	N/A	HQ876573*
C2945 e	MW695446	N/A	N/A	HQ876577*

for molecular analysis. Table 1 lists the specimens utilized. Genomic DNA was extracted from six Tunisian single animals using a rapid salt and ethanol precipitation following the protocol described by Cesari *et al.* (2009). We amplified four DNA fragments: the small ribosome subunit (18S rRNA), the large ribosome subunit (28S rRNA), the internal transcribed spacer (ITS-2), and the cytochrome oxidase subunit I (COI), using the primers and protocols described by Bertolani *et al.* (2014), Cesari *et al.* (2016), Stec *et al.* (2018d) and Cesari *et al.* (2009), respectively. Additionally, a portion of the 18S gene was sequenced for two additional specimens of *M. sandrae* sampled in the locus typicus of the species (Black Forest, Germany). GenBank accession numbers of obtained sequences are listed in Table 1. The amplified products were gel purified using the Wizard Gel and PCR cleaning (Promega) kit, and fragments were sequenced according to the protocols described in Bertolani *et al.* (2014). All COI sequences were translated into protein sequences in MEGAX (Kumar *et al.* 2018) to check for the presence of stop codons, and therefore for the presence of pseudogenes. Sequences of the four genes pertaining to species of the *hufelandi* group were used for molecular comparisons (Table 2) and aligned with the newly produced sequences using the MAFFT algorithm (Katoh *et al.* 2002) as implemented in the MAFFT online service (Katoh *et al.* 2017) and checked by visual inspection (Supp. file 3, Supp. file 4, Supp. file 5, Supp. file 6). Uncorrected pairwise distances were computed using MEGAX (Supp. file 7, Supp. file 8, Supp. file 9, Supp. file 10). Furthermore, relationships between COI haplotypes pertaining to the clade A nested in the *M. hufelandi* complex sensu Stec *et al.* (2021) were estimated using a parsimony network, by applying the method described in Templeton *et al.* (1992), as implemented in TCS ver. 1.21 (Clement *et al.* 2000) and visualized using tcsBU (Múrias dos Santos *et al.* 2016). A 95% connection limit was employed, as it has been suggested as a useful general tool in species assignments and discovery (Hart & Sunday 2007). Putative species were also inferred by using the Poisson Tree Process (PTP; Zhang *et al.* 2013) and the Automatic Barcode Gap Discovery method (ABGD; Puillandre *et al.* 2012). PTP is a coalescent-based species delimitation method that uses non-ultrameric gene trees as input, and utilizes heuristic algorithms to identify speciation events relative to numbers of substitutions. The PTP method produces robust diversity estimates, in some cases more robust than those estimated under the generalized mixed Yule coalescent model (Tang *et al.* 2014). The starting gene tree was a maximum likelihood (ML) tree computed using RAxML ver. 7.2.4 (Stamatakis 2006), as implemented in CIPRES (Miller *et al.* 2010), under the GTR+I+G model, as inferred by using the Akaike Information Criterion on jModelTest2 (Guindon & Gascuel 2003; Durriba *et al.* 2012). A sequence of *Mesobiotus hilariae* Vecchi, Cesari, Bertolani, Jönsson, Rebecchi & Guidetti, 2016 (GenBank accession number: KT226108) was used as outgroup. Bootstrap

resampling with 1000 replicates was undertaken via the rapid bootstrap procedure of Stamatakis *et al.* (2008) to assign support to branches in the ML tree. In order to consider different evolutionary models for the three COI codons, a Bayesian tree was computed using the following models, as inferred by MrModeltest ver. 2 (Nylander 2004): SYM+I+G for the first position of the codon, GTR+I+G for the second position of the codon and GTR+G for the third position of the codon. The Bayesian dendrogram was computed with the program MrBayes ver. 3.2.7a (Huelsenbeck & Ronquist 2001; Ronquist & Huelsenbeck 2003), as implemented in CIPRES. Two independent runs, each of four Metropolis-coupled Markov chains Monte Carlo method, were launched for 2×10^7 generations, and trees were sampled every 1000 generations. Convergence of runs was assessed by tracking average standard deviation of split frequencies between runs and by plotting the log likelihood of sampled trees in TRACER ver. 1.7 (Rambaut *et al.* 2018) and the first 2×10^6 sampled generations were discarded as burn-in. In the distance-based ABGD method, the sequences are sorted into hypothetical species based on the barcode gap (i.e., whenever the divergence among organisms belonging to the same species is smaller than divergence among organisms from different species). The method first detects the barcode gap as the first significant gap beyond a model-based one-sided confidence limit for intraspecific divergence, and then uses it to partition the data. ABGD settings for the COI dataset were: prior minimum divergence of intraspecific diversity (Pmin) = 0.001; prior maximum divergence of intraspecific diversity (Pmax) = 0.1; Steps = 10 and gap width = 1.5. The analysis was performed on the ABGD website (<https://bioinfo.mnhn.fr/abi/public/abgd/abgdweb.html>).

Abbreviations

DIC = Differential Interference Contrast light Microscopy
LM = Light Microscopy
PhC = Phase Contrast light Microscopy
SEM = Scanning Electron Microscopy

Repository

UNIMORE = Department of Life Sciences, University of Modena and Reggio Emilia, via G. Campi 213/d, Modena, Italy

Results

Taxonomic account of the new species

Phylum Tardigrada Doyère, 1840
Class Eutardigrada Richters, 1926
Order Parachela Schuster, Nelson, Grigarick & Christenberry, 1980
Superfamily Macrobiotidea Thulin, 1928 in Marley *et al.* 2011
Family Macrobiotidae Thulin, 1928
Genus *Macrobiotus* C.A.S. Schultze, 1834

Macrobiotus azzunae sp. nov.

urn:lsid:zoobank.org:act:933CCC06-F69D-49E2-AF4F-0C042D8F5C99

Figs 1–4, 5A, C, 7

Etymology

The new species is dedicated in honor of Atf Azzouna, professor in the Faculty of Mathematical, Physical and Natural Sciences of Tunis and supervisor of the PhD thesis of Jamila Ben Marnissi.

Table 2 (continued on next page). Sequences of species of the *Macrobiotus hufelandi* group used for molecular comparisons with *M. azzunae* sp. nov.

Species	Accession number			Country	Reference
	18S	28S	COI		
<i>Macrobiotus caelestis</i>	MK737073		MK737922	Kyrgyzstan	Coughlan <i>et al.</i> 2019
<i>Macrobiotus canarius</i>	MH063922		MH057765–6	Canary Island	Stec <i>et al.</i> 2018c
<i>Macrobiotus crustulus</i>	MT261912		MT260371	French Guiana	Stec <i>et al.</i> 2020a
<i>Macrobiotus engbergi</i>	MN443039		MN444824–6	Greenland	Stec <i>et al.</i> 2020b
<i>Macrobiotus hannaе</i>	MH063922		MH057764	Poland	Nowak & Stec 2018
<i>Macrobiotus hufelandi</i>	GQ849024			Greenland	Jørgensen <i>et al.</i> 2010
			HQ876584–88	Germany	Bertolani <i>et al.</i> 2011b
<i>Macrobiotus cf. hufelandi</i> sp. 1			HQ876589	Germany	
			HQ876590–4	Switzerland	Bertolani <i>et al.</i> 2011b
			HQ876595–7	Italy	
<i>Macrobiotus</i> group <i>hufelandi</i>	JX888491–8538			USA	Adams, unpublished
<i>Macrobiotus joannae</i>	HQ604974–5			Australia	Bertolani <i>et al.</i> 2014
<i>Macrobiotus kamilae</i>	MK737070		MK737920–1	India	Coughlan & Stec 2019
<i>Macrobiotus kristenseni</i>	KC193577		KC193572–6	Argentina	Guidetti <i>et al.</i> 2013
<i>Macrobiotus macrocalix</i>	HQ604976		FJ176203–17	Italy	Bertolani <i>et al.</i> 2014
	MH063926		MH057767	Poland	Stec <i>et al.</i> 2018c
<i>Macrobiotus cf. macrocalix</i>			JX683812–20	Portugal	Vicente <i>et al.</i> 2013
<i>Macrobiotus cf. nelsonae</i>	HQ604965–6			USA	Bertolani <i>et al.</i> 2014
<i>Macrobiotus noongaris</i>	MK737069		MK737919	Australia	Coughlan & Stec 2019
<i>Macrobiotus papei</i>	MH063881		MH057763	Tanzania	Stec <i>et al.</i> 2018b
<i>Macrobiotus paulinae</i>	KT935502		KT951668	Kenya	Stec <i>et al.</i> 2015
<i>Macrobiotus persimilis</i>			EU244608	Germany	Schill, unpublished
<i>Macrobiotus polonicus</i>	HM187580		HM150647	Poland	Wehnicz <i>et al.</i> 2011
	MN888369		MN888317–19	Austria	Stec <i>et al.</i> 2021
	MN888370		MN888320–21	Slovakia	Stec <i>et al.</i> 2021

Table 2 (continued).

Species	Accession number			Country	Reference
	18S	28S	ITS2		
<i>Macrobiotus polyptiformis</i>	KX810008	KX810011–2	KX810010	Ecuador	Roszkowska <i>et al.</i> 2017
<i>Macrobiotus cf. recens</i>	MH063927	MH057768–9	MH063932–3	Spain	Stec <i>et al.</i> 2018
<i>Macrobiotus sandrae</i>		HQ876567–73		Italy	Bertolani <i>et al.</i> 2011b
		HQ876574–83		Germany	
<i>Macrobiotus sapiens</i>	DQ839601		GQ403680	Croatia	Schill <i>et al.</i> 2010
	HQ604971			Italy	Bertolani <i>et al.</i> 2014
<i>Macrobiotus scoiticus</i>	KY797265	KY797267	KY797268	Scotland	Stec <i>et al.</i> 2017
<i>Macrobiotus shonaticus</i>	MG757132	MG757136–7	MG757134–5	Japan	Stec <i>et al.</i> 2018a
<i>Macrobiotus cf. shonaticus</i>		LC431582–90	LC431591–9	Japan	Sugiura <i>et al.</i> 2020
<i>Macrobiotus terminalis</i>		JN673958–62		Italy	Cesari <i>et al.</i> 2011
<i>Macrobiotus cf. terminalis</i>		AY598775		Italy	Guidetti <i>et al.</i> 2005
<i>Macrobiotus vladimiri</i>		JX683810–11		Portugal	Vicente <i>et al.</i> 2013
		HM136931–4		Italy	Bertolani <i>et al.</i> 2011a
<i>Macrobiotus wandae</i>	FJ435738–42	FJ435751–5		Spain	Guil & Giribet 2012
	MN888375		MN888347	Finland	Stec <i>et al.</i> 2021
	MN435109–13		MN435120–22	Nepal	Kayastha <i>et al.</i> 2020

Type material

Holotype

TUNISIA • spec. of unidentified sex; North-West Tunisia, Kroumiri Mountains, Ain Soltan forest, Jendouba; 36°31'21.788" N, 8°19'57.741" E; 893 m a.s.l.; Apr. 2017; Marnissi leg.; moss on trunk of *Quercus canariensis*; UNIMORE, slide code C4218–S32.

Paratypes

TUNISIA • 17 specs, sex unidentified; same collection data as for holotype; UNIMORE, slide codes C4218–S2 to C4218–S7, C4218–S9, C4218–S17, C4218–S30, C4218–S31, C4218–S33 to C4218–S35 • 3 eggs; same collection data as for holotype; UNIMORE, slide codes C4218–S10, C4218–S11, C4218–S25.

Type depositories

The holotype (slide: C4218–S32), 17 paratypes (slides C4218–S2 to C4218–S7, C4218–S17, C4218–S30, C4218–S31, C4218–S33 to C4218–S35), 3 eggs (slides: C4218–S10/11/25) and two vouchers (slides SP04 and SP07, corresponding to specimens C4218 V4 and C4218 V7, respectively) mounted in Faure-Berlese fluid, are deposited in the Bertolani collection at the Department of Life Science, UNIMORE, Modena, Italy.

Type locality

NW Tunisia, Kroumirie mountains, Ain Soltan forest, Jendouba, 36°31'21.788" N, 8°19'57.741" E. Altitude 893 m a.s.l.

Description

Adult specimens

Body white, transparent after mounting in Faure-Berlese, from 162.2 to 410.3 μm in length (Fig. 2A, Table 3; structures measured only in the animals more than 200 μm in length). Eye spots present, even after mounting. Cuticle smooth but with small round or oval pores, 1–1.5 μm in diameter (Fig. 2B), better visible after fixation in Carnoy and orcein staining (Fig. 3C), scattered randomly on the entire cuticle, including the dorsal surface of all legs. With SEM, pores look oval or in the shape of a seed (Fig. 3A, D) with the largest diameter of 0.7–0.8 μm . Weak cuticular granulation also present on the lateral surface of all legs and specially on legs IV (Fig. 2B, arrow). Only with SEM is it possible to define the shape of the granulation on the legs, which looks as a regular disposition of star-shaped protuberances (about 0.3 μm ; Fig. 3F). Six buccal sensory lobes around the mouth, well recognizable with SEM. Mouth antero-ventral; buccal-pharyngeal apparatus of the *Macrobiotus* type (sensu Pilato & Binda 2010), with ventral lamina and ten small peribuccal lamellae (in the holotype, after mounting, separated from the mouth). Buccal armature, corresponding to oral cavity armature, OCA, according to Michalczyk & Kaczmarek (2003), without an anterior band of teeth visible, corresponding to the first band of teeth according to Michalczyk & Kaczmarek (2003), and to the anterior band of the buccal ring according to Guidetti *et al.* (2012); posterior band of teeth poorly visible, corresponding to second band of teeth, according to Michalczyk & Kaczmarek (2003), followed by three dorsal and three ventral crests, corresponding to third band of teeth according to Michalczyk & Kaczmarek (2003); the dorsal crests (Fig. 2D) are distinct transverse ridges, whereas the ventral crests (Fig. 2E) appear as two separate lateral transverse ridges and a roundish median tooth. The posterior band of teeth and the transverse ridges are part of the buccal tube, according to Guidetti *et al.* (2012). Buccal tube narrow; pharyngeal bulb spherical with triangular apophyses, two rod-shaped macroplacoids, relatively short, the first longer than the second and evidently but not deeply narrowed at its middle (Fig. 2C), the second with a not particularly evident subterminal constriction. Microplacoid present. Slender claws of the *hufelandi* type (sensu Pilato & Binda 2010); the external claw longer than the internal one and the posterior longer than

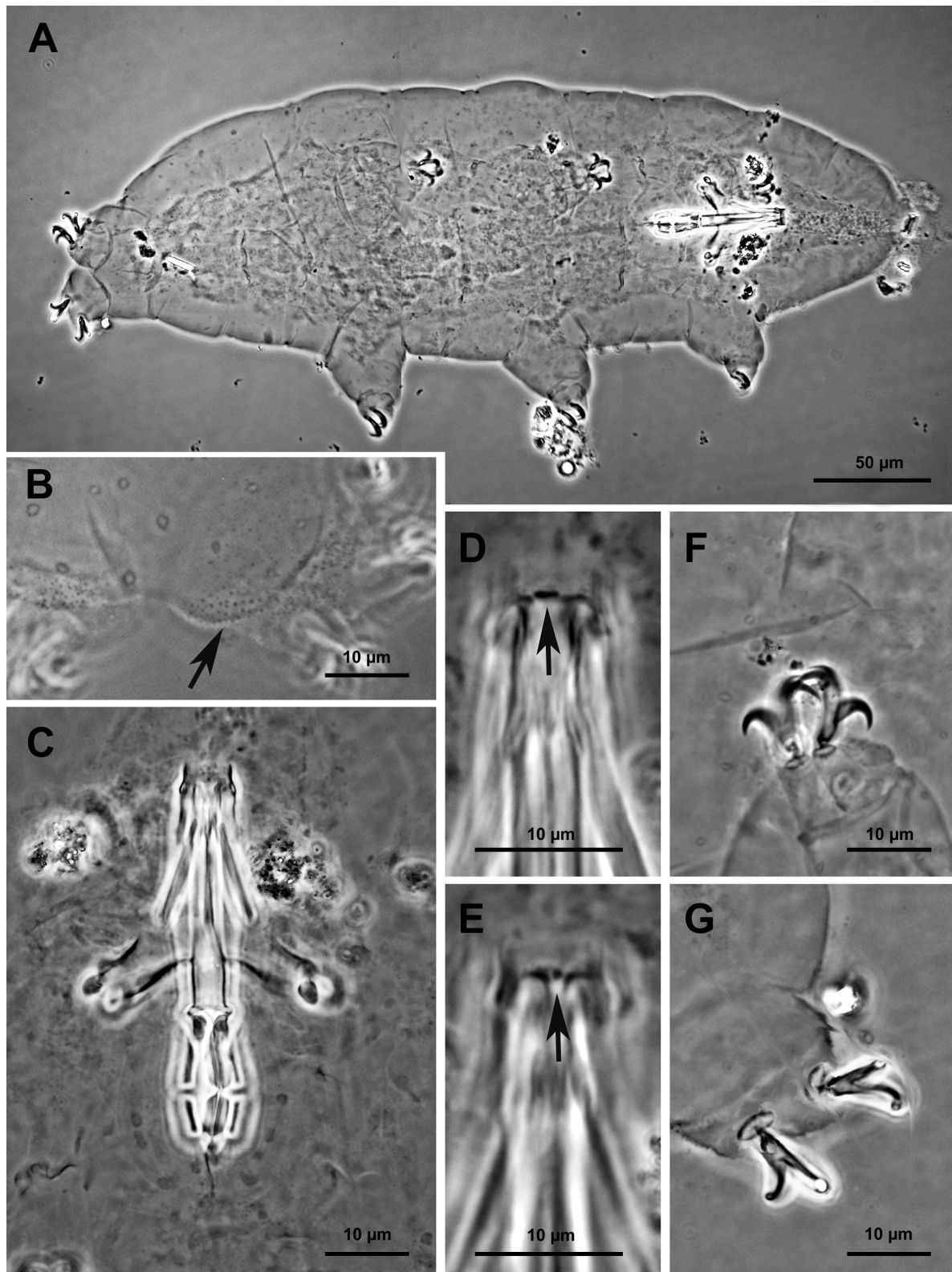


Fig. 2. *Macrobiotus azzunae* sp. nov., holotype (UNIMORE, slide C4218–S32). A. In toto animal. B. Cuticular pores and leg granulation (arrow) on the hind legs. C. Buccal-pharyngeal apparatus. D. Buccal armature: dorsal crests (arrow). E. Buccal armature: ventral crests (arrow). F. Claw and lunulae of the third pair of legs. G. Claw and lunulae of the fourth pair of legs. A–G: PhC.

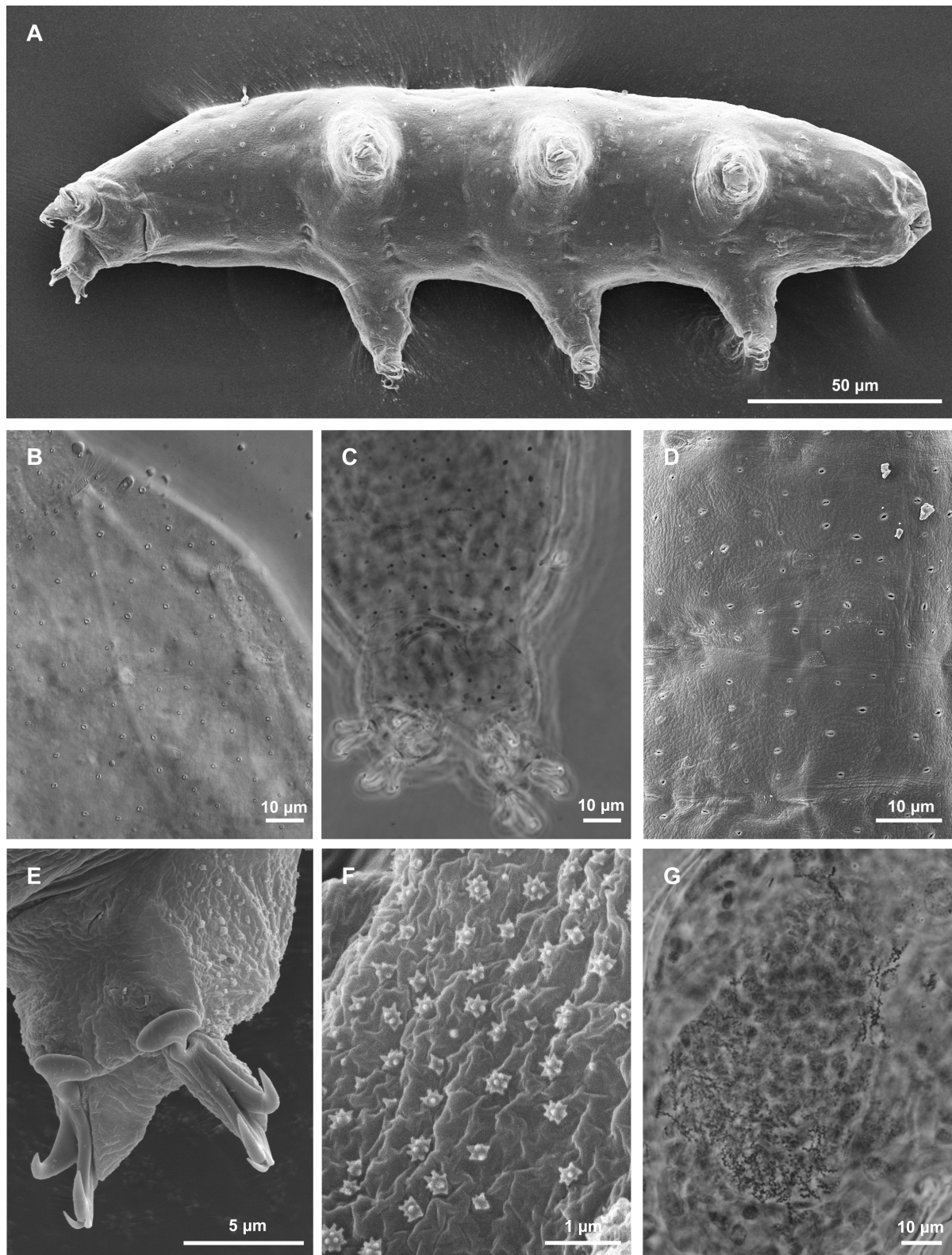


Fig. 3. *Macrobiotus azzunae* sp. nov., paratypes. **A.** In toto animal. **B–D.** Cuticular pores. **E.** Fourth pair of legs with smooth lunules and peculiar granulation on the legs. **F.** Granulation on the legs with a star-shaped organization. **G.** Male with testis full of mature spermatozoa with elongate, helicoidal nucleus. A, D–F: SEM (stub-C4218); B: in vivo DIC; C, G: orcein (not permanent slide TN02–04) PhC.

Table 3. Measurements (in μm) and *pt* of selected morphological structures of individuals of *Macrobiotus azzunae* sp. nov. mounted in Faure-Berlese. Abbreviations: Nr = number of specimens/structures measured; Range = smallest–largest value; SD = standard deviation. * Three animals below 200 μm in length were not considered in the analysis. Two were not measurable, while data of the only measurable animal below 200 μm is available in the supplementary material (Supp. file 1).

Character	Nr	Range μm	Mean \pm SD	Holotype μm	
Body length	10*	246.0–410.3		374.0	
Buccal tube length	13	29.4–38.5		36.6	
			<i>pt</i>	μm	<i>pt</i>
Buccal tube external width	13	11.6–15.9	13.3 \pm 1.2	5.8	15.9
Stylet support insertion point	13	75.1–80.5	76.7 \pm 1.5	27.9	76.1
Placoid row	13	50.1–57.7	51.8 \pm 2.4	18.9	51.5
Macroplacoid row	13	37.7–48.7	44.6 \pm 3.1	15.7	42.7
First macroplacoid	13	24.3–29.7	26.6 \pm 2.2	9.3	25.5
Second macroplacoid	13	16.0–20.7	18.3 \pm 1.5	6.6	18.1
Microplacoid	13	5.9–8.6	6.9 \pm 1.0	2.6	7.0
External Claws III main branch	13	27.1–31.3	29.8 \pm 1.3	11.5	31.3
External Claws III secondary branch	13	20.1–25.8	22.5 \pm 1.6	8.1	22.1
Internal Claws III main branch	13	24.9–30.2	27.7 \pm 1.6	10.2	27.9
Internal Claws III secondary branch	13	18.8–24.2	21.4 \pm 1.7	7.2	19.7
Posterior Claws IV main branch	9	29.1–34.6	32.7 \pm 1.8	12.2	33.2
Posterior Claws IV secondary branch	9	21.6–24.9	23.8 \pm 1.6	9.1	24.9
Anterior Claws IV main branch	10	28.2–32.5	30.1 \pm 1.9	11.1	30.3
Anterior Claws IV secondary branch	10	20.4–26.2	23.2 \pm 1.7	8.8	24.0

the anterior. Primary branches of each claw with distinct accessory points (Fig. 2F), a common tract of medium length (about a third of the total claw length) and an evident stalk connecting the claw to the lunule. Lunules under all claws, smooth, larger on the hind legs (Figs 2G, 3E). Cuticular bars under claws absent.

The population is dioecious (gonochoristic). Males were recognized using orcein staining, which revealed that the testis is filled with spermatozoa with a coiled head (Fig. 3G) and spermatids. No morphological secondary sexual dimorphism, such as gibbosities on legs IV in males, was identified.

Eggs

Eggs are laid freely, and are white, spherical or slightly oval. One egg containing a fully developed embryo showed the shape of the buccal-pharyngeal apparatus (Fig. 4A). Processes of the eggshell are in the shape of inverted goblets (Fig. 4B) with conical trunks and well-defined distal discs as large as the process bases (for measurements see Table 4). Distal discs concave, with a median small protuberance and, using PhC, with border often smooth, or sometimes slightly jagged, or slightly ragged (Fig. 4C),

Table 4. Measurements (in μm) of selected morphological structures of the eggs of *Macrobiotus azzunae* sp. nov. mounted in Faure-Berlese. Abbreviations: Nr = number of eggs/structures measured; Range = smallest–largest value; SD = standard deviation.

Character	Nr	Range	Mean \pm SD
Egg diameter without processes	2	64.7–80.6	–
Egg diameter with processes	2	72.4–89.2	–
Process nr on egg circumference	2	29–33	–
Process height	10	4.2–6.4	5.4 \pm 0.6
Process base width	10	3.2–5.2	4.2 \pm 0.5
Distal disc width	10	3.2–5.2	4.4 \pm 0.6
Inter process distance	10	2.7–4.8	3.5 \pm 0.5

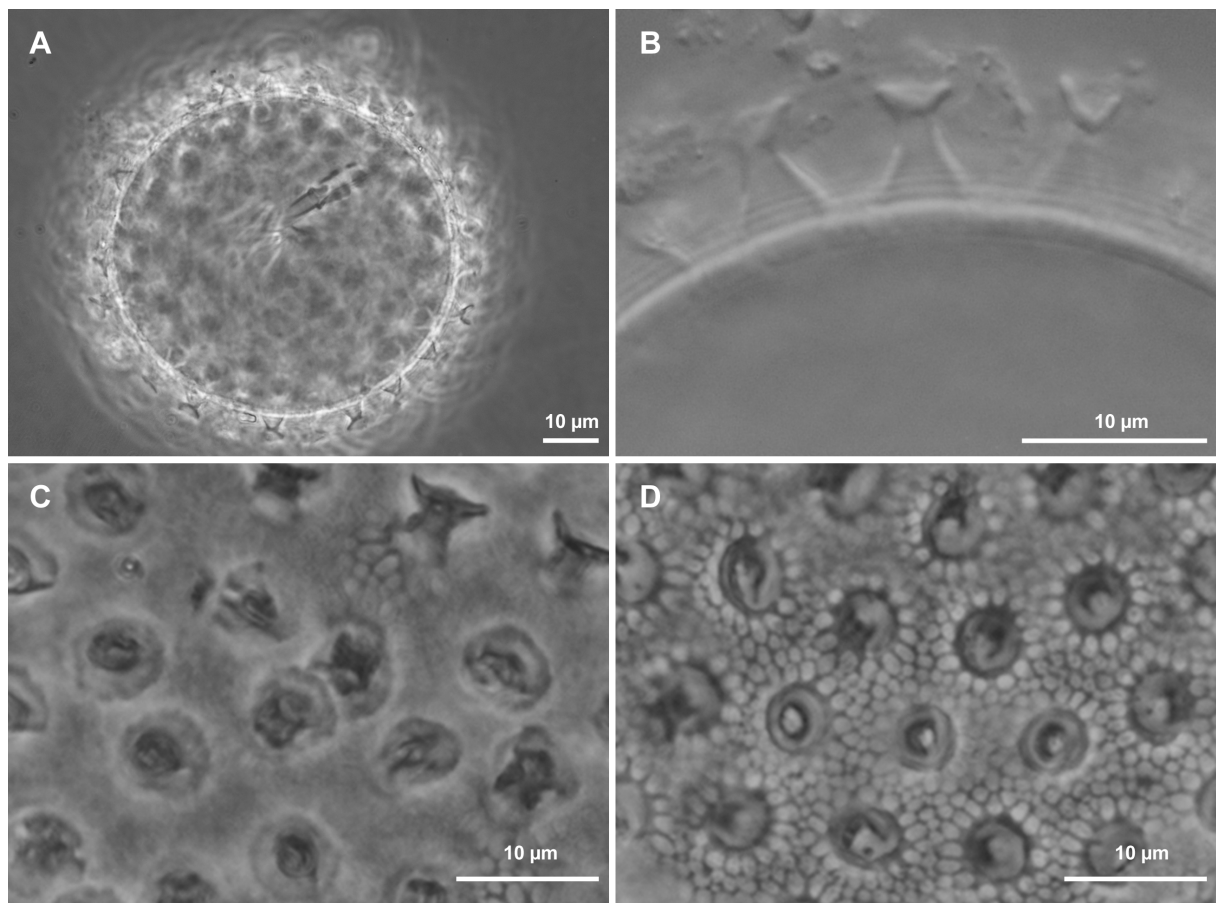


Fig. 4. Egg of *Macrobiotus azzunae* sp. nov., paratype (UNIMORE, slide C4218–S11). **A.** In toto egg with buccal-pharyngeal apparatus of its embryo at the end of development. **B.** Processes of the eggshell (midsection). **C.** Distal discs of the eggshell processes. **D.** Surface of the eggshell between processes. A, C–D: PhC; B: DIC.

but never clearly jagged, serrated or dentate. Surface among processes of the *hufelandi* type (sensu Kaczmarek & Michalczyk 2017a), i.e., covered by a very thin grid (Fig. 4D). Meshes around the process bases slightly larger and with slightly thicker wires compared with interbasal meshes. Mesh diameter around 0.5 μm .

Comparisons

Macrobotus azzunae sp. nov. has eggs with processes as inverted goblets and a reticulate eggshell between the processes. Consequently, a comparison must be done with the *Macrobotus* species listed by Kaczmarek & Michalczyk (2017a) with *hufelandi* type eggshells, excluding the species with processes that are not like inverted goblets, and adding the species with *hufelandi* type chorion eggs described after that publication. The species with *hufelandi* type chorion eggs that do not have processes as inverted goblets are *Macrobotus acadianus* (Meyer & Domingue, 2011), *M. dariae* Pilato & Bertolani, 2004, *M. lissostomus* Durante Pasa & Maucci, 1979, *M. santoroi* Pilato & D'Urso, 1976, and *M. scoticus* Stec, Morek, Gąsiorek, Blagden & Michalczyk, 2017. Moreover, *M. azzunae* sp. nov. has egg processes with distal discs with a smooth or slightly jagged border, therefore it differs from all the species that have clearly indented, serrated or clearly jagged distal discs, such as: *Macrobotus canaricus* Stec, Krzywański & Michalczyk, 2018, *M. crustulus* Stec, Dudziak & Michalczyk, 2020, *M. hanna*e Nowak & Stec, 2018 (whose egg surface is more cribriform than reticulate), *M. hibiscus* de Barros, 1942, *M. horningi* Kaczmarek & Michalczyk, 2017b (which also has very high processes), *M. hufelandi* C.A.S. Schultze, 1834, *M. humilis* Binda & Pilato, 2001, *M. iharosi* Pilato, Binda & Catanzaro 1991, *M. joannae* Pilato & Binda, 1983, *M. julianae* (Meyer, 2012), *M. kamilae* Coughlan & Stec, 2019, *M. modestus* Pilato & Lisi, 2009, *M. noonragis* Coughlan & Stec, 2019, *M. papei* Stec, Kristensen & Michalczyk, 2018 (with particularly long filaments starting from the distal disc), *M. paulinae* Stec, Smolak, Kaczmarek & Michalczyk, 2015, *M. polypiformis* Roszkowska, Ostrowska, Stec, Janko & Kaczmarek, 2017 (even with cog-teeth extended to form a long, thin, hair-like and flexible filament), *M. punctillus* Pilato, Binda & Azzaro, 1990, *M. sapiens* Binda & Pilato, 1984, *M. sottilei* Pilato, Kiosya, Lisi & Sabella, 2012.

For the shape of the egg *Macrobotus azzunae* sp. nov. differs from *M. rawsoni* Horning, Schuster & Grigarick, 1978 because this species has only one strip of meshes around each egg process (see Kaczmarek & Michalczyk 2017b); it differs from *M. serratus* Bertolani, Guidi & Rebecchi, 1996 because in this species the egg surface is porous more than reticulated, with pores small and spaced from each other, and its egg processes have a large, often square, distal disc; it differs from *M. seychellensis* Biserov, 1994 because the distal disc of the egg processes of this species, even though not dentate, has long and very developed lobes.

The remaining nine species of the *hufelandi* group should be compared singularly.

***Macrobotus almadai* Fontoura, Pilato & Lisi, 2008**

Macrobotus azzunae sp. nov. differs from *M. almadai* in having a posterior band of teeth in the buccal cavity visible with LM (not visible in *M. almadai*), and distal disc with a jagged margin instead of very small teeth as in *M. almadai*.

***Macrobotus canaricus* Stec, Krzywański & Michalczyk, 2018**

With LM the margin of the distal disc of *M. azzunae* sp. nov., never dentate in this species, looks similar to that of *M. canaricus*, but the SEM images of the eggs of the latter species evidence the presence of an almost dentate disc. Moreover, the peribasal meshes of the eggshell are larger than interbasal ones in the new species while they do not differ from the interbasal ones in *M. canaricus*; regarding the animals there are differences in the buccal armature: in *M. azzunae* sp. nov. the posterior band of teeth is visible with LM (even though poorly) and the three dorsal crests are distinct transverse ridges, while

in *M. canaricus* the posterior band of teeth is visible only with SEM and with LM the dorsal teeth form a transversal ridge weakly divided into three teeth.

***Macrobotus madegassus* Maucci, 1993**

The new species differs from *M. madegassus* by the presence of the eye spots (absent in *M. madegassus*), pores on the cuticle (absent in *M. madegassus*), presence in the buccal armature of posterior band of teeth, even though weak (fully absent in *M. madegassus*), buccal tube much larger (*pt* of the holotypes 15.9 vs 7), insertion of the stylet supports on the buccal tube much more posterior (*pt* of the holotypes 76.1 vs 68), first and second macroplacoid longer (*pt* of the holotypes 25.5 and 18.1 vs 21.3 and 12.0), lunules on the hind legs without kerning (crenate in *M. madegassus*), eggshell processes with distal disc as large as the base (similar range 3.2–5.2 for both measurements) with respect to that of *M. madegassus* (disc vs base: 4.3–5.4 vs 2.3–2.6).

***Macrobotus martini* Bartels, Pilato, Lisi & Nelson, 2009**

The cuticular pores in *M. azzunae* sp. nov. are much smaller than those of *M. martini*; the distal disc of the egg processes in *M. azzunae* sp. nov. has a diameter similar to that of the process base, while in *M. martini* the distal disc is much narrower than the base.

***Macrobotus nebrodensis* Pilato, Sabella, D'Urso & Lisi, 2017**

Macrobotus azzunae sp. nov. differs from *M. nebrodensis* by the absence of the cuticular bar near the lunules on the first three pairs of legs (a faint bar is present in *M. nebrodensis*). The egg processes of *M. azzunae* sp. nov. are in higher number on the circumference (29–33) with respect to those of *M. nebrodensis* (18). In the latter species there are some egg processes very high (up to 20.6 μm), while in the new species process height and shape are more uniform. The difference in the eggshell between meshes around the process base and the others is much less evident in *M. azzunae* sp. nov. than in *M. nebrodensis*.

***Macrobotus personatus* Biserov, 1990**

The new species differs from *M. personatus* by the posterior band of the buccal armature less evident, the presence of a clear constriction in the first macroplacoid (Fig. 5A), in the paratype of *M. personatus* examined by us barely identifiable (Fig. 6A) and, according to Biserov (1990) usually absent in the type material of that species. The pores on the cuticle of *M. azzunae* sp. nov. are small, approximately 1 μm in diameter, while in *M. personatus* they are strongly elliptic and about 3 μm in length (Fig. 6B). Lunules on leg IV are always smooth in *M. azzunae* sp. nov., sometimes indented in *M. personatus*. With respect to the eggs of *M. personatus* (Fig. 6C–D), the egg processes of *M. azzunae* sp. nov. (Figs 4C–D, 5C) are clearly shorter, 5.4 ± 0.6 vs 9.5 ± 0.5 (range 4.2–6.4 vs 9–10.5) and with a narrower base and distal disc (both 3.2–5.2 vs 7–10.5 and 7–9 respectively). Males are present in the new species, while in *M. personatus* only females were found (Biserov 1990), suggesting parthenogenesis in that species.

***Macrobotus sandrae* Bertolani & Rebecchi, 1993**

The new species differs from *M. sandrae* for the eggshell shape, with thinner wires of the reticulum and meshes around the processes larger than the inter-process meshes in *M. azzunae* sp. nov. (Fig. 5C), all meshes similar in size in *M. sandrae* (Fig. 5D). Figure 5C–D also show a difference in the process base diameter, narrower in *M. azzunae* sp. nov. With regard to the animals, *M. azzunae* sp. nov. differs from *M. sandrae* by a constriction of the first macroplacoid much more pronounced (Fig. 5A; it is hardly visible in *M. sandrae*; Fig. 5B). Moreover, in animals of similar size the posterior band of the buccal armature is just less evident in the new species, and lunules on the hind legs are without hint of teeth (but teeth, present in the holotype of *M. sandrae*, are often difficult to identify in other specimens of that species).

***Macrobiotus terminalis* Bertolani & Rebecchi, 1993**

Macrobiotus azzunae sp. nov. differs from *M. terminalis* for the absence of granulation on the cuticle (noted only in the redescription of *M. terminalis*; see Cesari *et al.* 2011), for the absence of teeth on the lunules, especially evident on the hind legs of *M. terminalis*, and for the presence of males, absent in *M. terminalis* (see redescription by Cesari *et al.* 2011).

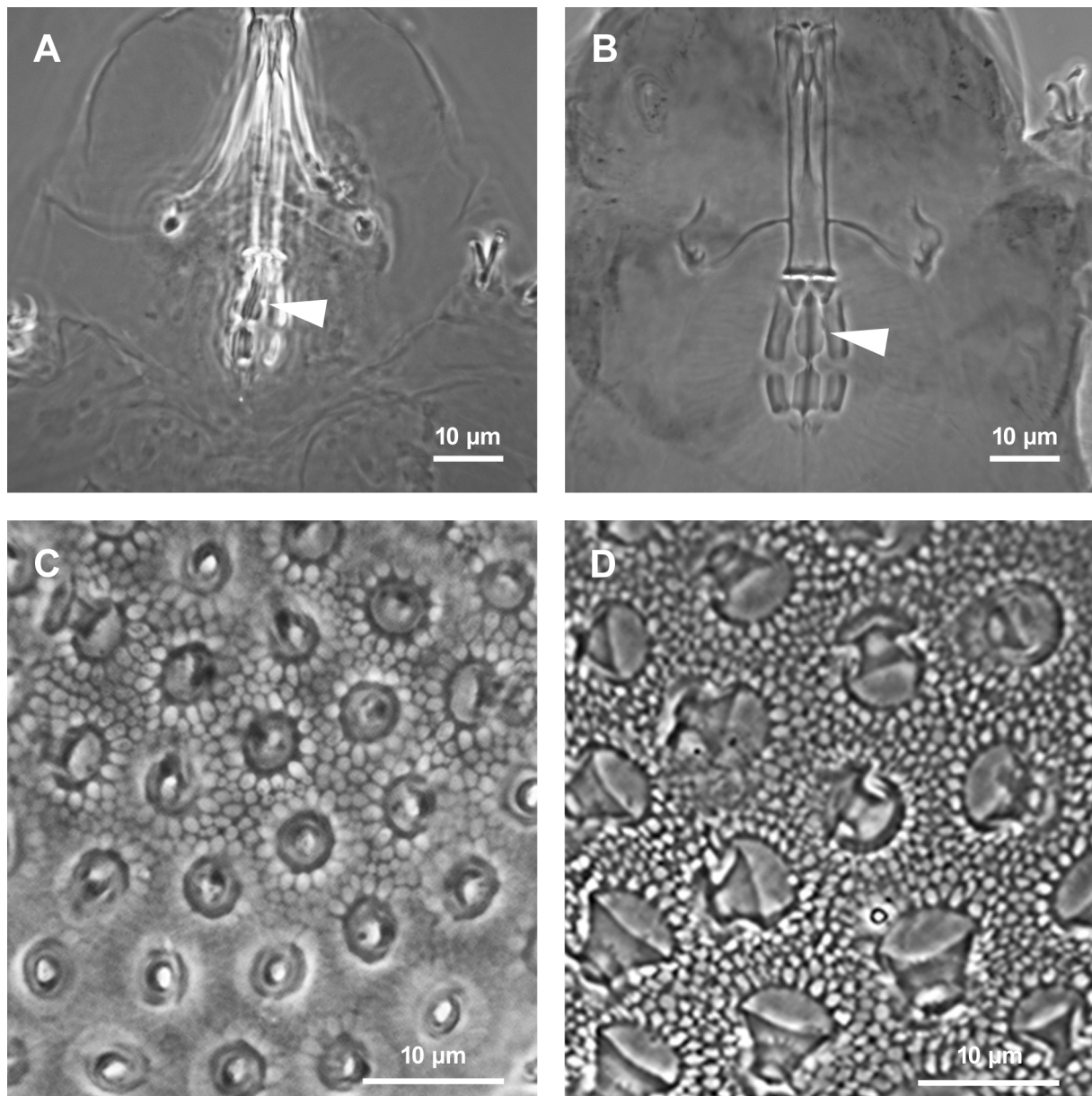


Fig. 5. Comparison between *M. azzunae* sp. nov. and *M. sandrae* Bertolani & Rebecchi, 1993. **A.** Placoids in *M. azzunae* sp. nov., paratype (UNIMORE, slide C4218–S30). **B.** Placoids in *M. sandrae* (UNIMORE, slide C442–S79); arrowheads evidence the different constriction depth of the first macroplacoid. **C.** Eggshell in *M. azzunae* sp. nov., paratype (UNIMORE, slide C4218–S4). **D.** Eggshell in *M. sandrae* (UNIMORE, slide C2346–S2); in *M. azzunae* sp. nov. there are smaller processes and reticulation with thinner wires and larger net around the processes than in *M. sandrae*. A–D: PhC.

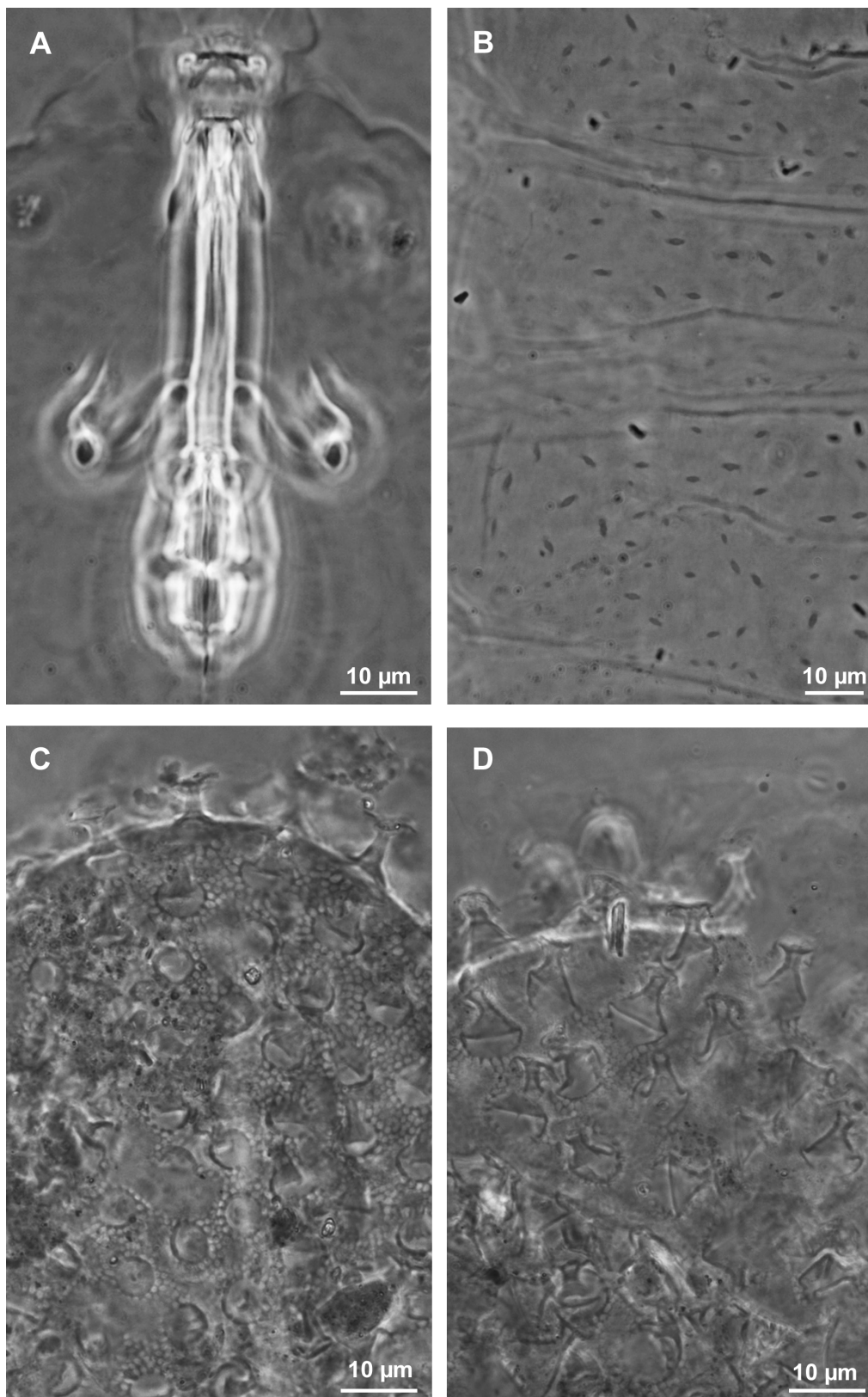


Fig. 6. *Macrobiotus personatus* Biserov, 1990, paratypes (Civic Museum of Natural History of Verona, Italy, CT14701). **A.** Buccal-pharyngeal apparatus with macroplacoids. **B.** Pores on the cuticle. **C.** Eggshell reticulation and egg processes. **D.** Egg processes. A–D: PhC.

***Macrobotus vladimiri* Bertolani, Biserov, Rebecchi & Cesari, 2011**

With respect to *M. vladimiri*, animals of *M. azzunae* sp. nov. reach a shorter length (up to 410.3 μm vs 515.1 μm), in *M. azzunae* sp. nov. the posterior band of teeth of the buccal armature is less evident and the lunules on the hind legs are not indented. In *M. azzunae* sp. nov. the egg diameter without processes (64.7–80.6 μm) is less than that of the eggs of *M. vladimiri* (89.9–92.0 μm); the processes are shorter (4.2–6.4 μm in the new species vs 6.5–8 μm in *M. vladimiri*). In the new species the base process diameter is narrower (3.2–5.2 μm) than in *M. vladimiri* (5.1–7.3 μm), the distal disc is weakly or not jagged (clearly jagged in *M. terminalis*). In *M. azzunae* sp. nov. males are present, while they are absent in *M. vladimiri*.

Genetic distances

The ranges of uncorrected genetic p-distances between *M. azzunae* sp. nov. and the other species of the *M. hufelandi* group (Supp. file 7, Supp. file 8, Supp. file 9, Supp. file 10), are as follows:

- 18S 0.1–5.6%, with the most similar being *M. sandrae* from Germany (present paper)
- 28S 0.1%, with the only available *M. vladimiri* from Spain (FJ435751–5)
- ITS-2 7.7–32.2%, with the most similar being *Macrobotus vladimiri* (MN888347) from Finland
- COI 6.3–25.6%, with the most similar being *Macrobotus sandrae* (HQ876574, HQ876577, HQ876578, HQ876579, HQ876581) from Germany

The COI dataset is the most complete and informative for species delimitation investigation. Both phylogenetic reconstructions on the COI dataset resulted in the same topology, and thus the ML tree was utilized for the PTP analysis (Fig. 7, left), which shows 14 putative species clusters: *M. crustulus*, *M. hanna*e, *M. cf. recens*, *M. canaricus*, *M. hufelandi*, *M. cf. hufelandi* sp.1, *M. terminalis*, *M. cf. terminalis*, *M. wandae*, *M. macrocalix*, *M. cf. macrocalix*, *M. vladimiri*, *M. sandrae* and *M. azzunae* sp. nov. This subdivision is further validated by both the ABGD and the haplotype network analysis (Fig. 7, centre and right). Present molecular data therefore confirms the validity of the erection of *M. azzunae* sp. nov.

Discussion

With the discovery of *M. azzunae* sp. nov. the species identified in Tunisia now comes to 33, all non-marine. This number is much lower than the number of non-marine species, 120, found in nearby Sicily (Pilato *et al.* 2017, 2019), an island extensively studied from a tardigradological point of view. This means that there is considerable potential for further discoveries that could come from the study of the various habitats colonized by tardigrades in Tunisia. Furthermore, the results will undoubtedly be greater and more informative if the research is carried out with an integrated approach, as done in this work.

Acknowledgments

This study was conducted within the framework of collaboration between Jamila Ben Marnissi who obtained personal financial assistance for the travel costs awarded by Prof. Monia Trabelsi (Department of Biological Sciences, Faculty of Mathematical, Physical and Natural Sciences of Tunis, University of Tunis El Manar), the director of the Laboratory of Ecology, Biology and Physiology of aquatic organisms, University of Tunis El Manar of Tunis, and Prof. Lorena Rebecchi, head of the research team of the laboratory of Evolutionary Zoology (EvoZoo, Department of Life Sciences of the University of Modena and Reggio Emilia, Italy), who supported all the investigations from an economical point of view. We would like to thank Paul Bartels (Warren Wilson College, Asheville, NC) for suggestions and English revision of the manuscript and Prof. Roberto Guidetti ADD: (Department of Life Sciences of the University of Modena and Reggio Emilia, Italy) for his collaboration and support.

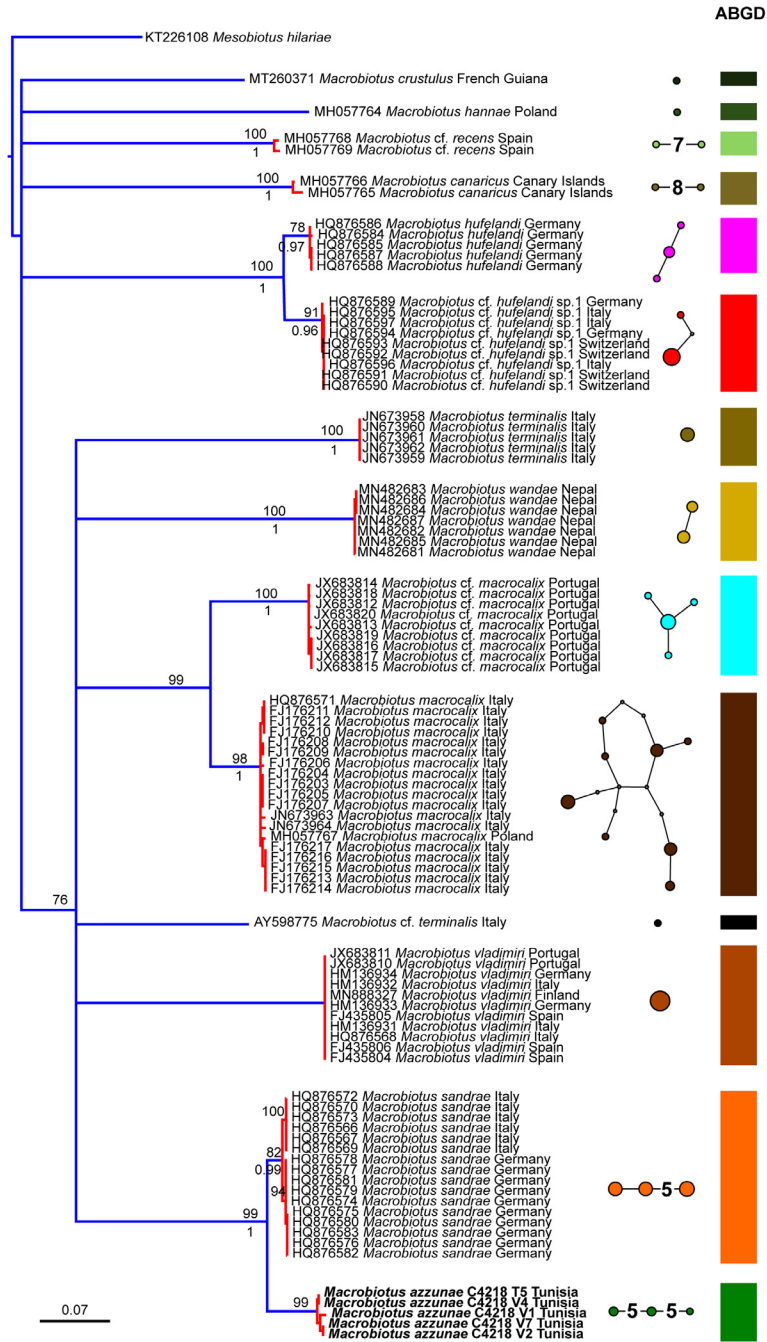


Fig. 7. Left: tree resulting from both the maximum likelihood analysis and the Bayesian inference of cytochrome c oxidase subunit I (COI) in *M. azzunae* sp. nov. specimens and sequences from GenBank. Values above branches point out bootstrap values, while values under branches represent posterior probability values. Results of the Poisson tree process analysis are provided using differently coloured branches: putative species are indicated using transitions from blue-coloured branches to red-coloured branches. Newly scored haplotypes are in bold. The scale bar shows the number of substitutions per nucleotide position. Centre: haplotype network of COI gene in *M. hufelandi* complex. Circles represent haplotypes, while circle surface denotes haplotype frequency. Networks falling below the value of the 95% connection limit are disconnected. Right: rectangles denote specimens grouped by ABGD analysis.

Contribution statement

The authors contributed equally to this work.

References

- Bertolani R., Biserov V., Rebecchi L. & Cesari M. 2011a. Taxonomy and biogeography of tardigrades using an integrated approach: new results on species of the *Macrobiotus hufelandi* group. *Invertebrate Zoology* 8 (1): 23–26. <https://doi.org/10.15298/invertzool.08.1.05>
- Bertolani R., Rebecchi L., Giovannini I. & Cesari M. 2011b. DNA barcoding and integrative taxonomy of *Macrobiotus hufelandi* C.A.S. Schultze 1834, the first tardigrade species to be described, and some related species. *Zootaxa* 2997: 19–36. <https://doi.org/10.11646/zootaxa.2997.1.2>
- Bertolani R., Guidetti R., Marchioro T., Altiero T., Rebecchi L. & Cesari M. 2014. Phylogeny of Eutardigrada: new molecular data and their morphological support lead to the identification of new evolutionary lineages. *Molecular Phylogenetics and Evolution* 76: 110–126. <https://doi.org/10.1016/j.ympev.2014.03.006>
- Binda M.G. & Pilato G. 1987. Tardigradi dell’Africa. V: Notizie sui Tardigradi del Nordafrica e descrizione della nuova specie *Macrobiotus diffusus*. *Animalia* 14: 177–191.
- Biserov V.I. 1990. On the revision of the *Macrobiotus* genus. The subgenus *Macrobiotus* s. str. is a new taxonomic status of the *hufelandi* group (Tardigrada, Macrobiotidae). Communication 2. *Zoologicheskij Zhurnal* 69: 38–50. [In Russian.]
- Cesari M., Bertolani R., Rebecchi L. & Guidetti R. 2009. DNA barcoding in Tardigrada: the first case study on *Macrobiotus macrocalix* Bertolani & Rebecchi, 1993 (Eutardigrada, Macrobiotidae). *Molecular Ecology Resources* 9 (3): 699–706. <https://doi.org/10.1111/j.1755-0998.2009.02538.x>
- Cesari M., Giovannini I., Bertolani R. & Rebecchi L. 2011. An example of problems associated with DNA barcoding in tardigrades: a novel method for obtaining voucher specimens. *Zootaxa* 3104: 42–51. <https://doi.org/10.11646/zootaxa.3104.1.3>
- Cesari M., Vecchi M., Palmer A., Bertolani R., Pilato G., Rebecchi L. & Guidetti R. 2016. What if the claws are reduced? Morphological and molecular phylogenetic relationships of the genus *Haplomacrobiotus* May, 1948 (Eutardigrada, Parachela). *Zoological Journal of the Linnean Society* 178 (4): 819–827. <https://doi.org/10.1111/zoj.12424>
- Cesari M., Montanari M., Kristensen R.M., Bertolani R., Guidetti R. & Rebecchi L. 2020. An integrated study of the biodiversity within the *Pseudechiniscus suillus-facettalis* group (Heterotardigrada: Echiniscidae). *Zoological Journal of the Linnean Society* 188 (3): 717–732. <https://doi.org/10.1093/zoolinlean/zlz045>
- Clement M., Posada D. & Crandall K. 2000. TCS: a computer program to estimate gene genealogies. *Molecular Ecology* 9: 1657–1660. <https://doi.org/10.1046/j.1365-294x.2000.01020.x>
- Coughlan K. & Stec D. 2019. Two new species of the *Macrobiotus hufelandi* complex (Tardigrada: Eutardigrada: Macrobiotidae) from Australia and India, with notes on their phylogenetic position. *European Journal of Taxonomy* 573: 1–38. <https://doi.org/10.5852/ejt.2019.573>
- Coughlan K., Michalczyk Ł. & Stec D. 2019. *Macrobiotus caelestis* sp. nov., a new tardigrade species (Macrobiotidae: *hufelandi* group) from the Tien Shan Mountains (Kyrgyzstan). *Annales Zoologici* 69 (3): 499–513. <https://doi.org/10.3161/00034541ANZ2019.69.3.002>
- Darriba D., Taboada G.L., Doallo R. & Posada D. 2012. jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods* 9 (8): 772. <https://doi.org/10.1038/nmeth.2109>

- Degma P., Bertolani R. & Guidetti R. 2020. Actual checklist of Tardigrada. 36th Edition: 8–07–2020. Available from <https://iris.unimore.it/retrieve/handle/11380/1178608/227296/> [accessed 28 Jun. 2021].
- Gąsiorek P., Stec D., Morek W., Marnissi J. & Michalczyk Ł. 2017. The tardigrade fauna of Tunisia, with an integrative description of *Bryodelphax maculatus* sp. nov. (Heterotardigrada: Echiniscidae). *African Zoology* 52 (2): 77–89. <https://doi.org/10.1080/15627020.2017.1297688>
- Guidetti R. & Bertolani R. 2005. Tardigrade taxonomy: an updated checklist of the taxa and a list of characters for their identification. *Zootaxa* 845 (1): 1–46. <https://doi.org/10.11646/zootaxa.845.1.1>
- Guidetti R., Gandolfi A., Rossi V. & Bertolani R. 2005. Phylogenetic analysis of Macrobiotidae (Eutardigrada, Parachela): a combined morphological and molecular approach. *Zoologica Scripta* 34: 235–244. <https://doi.org/10.1111/j.1463-6409.2005.00193.x>
- Guidetti R., Altiero T., Marchioro T., Sarzi Amadè L., Avdonina A.M., Bertolani R. & Rebecchi L. 2012. Form and function of the feeding apparatus in Eutardigrada (Tardigrada). *Zoomorphology* 131: 127–148. <https://doi.org/10.1007/s00435-012-0149-0>
- Guidetti R., Peluffo J.R., Rocha A.M., Cesari M. & Moly de Peluffo M.C. 2013. The morphological and molecular analyses of a new South American urban tardigrade offer new insights on the biological meaning of the *Macrobiotus hufelandi* group of species (Tardigrada: Macrobiotidae). *Journal of Natural History* 47: 2409–2426. <https://doi.org/10.1080/00222933.2013.800610>
- Guil N. & Giribet G. 2012. A comprehensive molecular phylogeny of tardigrades — adding genes and taxa to a poorly resolved phylum-level phylogeny. *Cladistics* 27: 1–29. <https://doi.org/10.1111/j.1096-0031.2011.00364.x>
- Guindon S. & Gascuel O. 2003. A simple, fast and accurate method to estimate large phylogenies by maximum-likelihood. *Systematic Biology* 52: 696–704. <https://doi.org/10.1080/10635150390235520>
- Hart M.W. & Sunday J. 2007. Things fall apart: biological species form unconnected parsimony networks. *Biology Letters* 3: 509–512. <https://doi.org/10.1098/rsbl.2007.0307>
- Huelsenbeck J.P. & Ronquist F. 2001. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* 17: 754–755. <https://doi.org/10.1093/bioinformatics/17.8.754>
- Iharos G. 1978. Data to the knowledge of the Tardigrada fauna of Tunisia. *Folia Entomologica Hungarica* 31: 161–164.
- Jørgensen A., Faurby S., Hansen J.G., Møbjerg N. & Kristensen R.M. 2010. Molecular phylogeny of Arthrotardigrada (Tardigrada). *Molecular Phylogenetics and Evolution* 54: 1006–1015. <https://doi.org/10.1016/j.ympev.2009.10.006>
- Kaczmarek Ł. & Michalczyk Ł. 2017a. The *Macrobiotus hufelandi* (Tardigrada) group revisited. *Zootaxa* 4363: 101–123. <https://doi.org/10.11646/zootaxa.4363.1.4>
- Kaczmarek Ł. & Michalczyk Ł. 2017b. A description of *Macrobiotus horningi* sp. nov. and redescriptions of *M. maculatus* comb. nov. Iharos, 1973 and *M. rawsoni* Horning *et al.*, 1978 (Tardigrada: Eutardigrada: Macrobiotidae: *hufelandi* group). *Zootaxa* 4363: 79–100. <https://doi.org/10.11646/zootaxa.4363.1.3>
- Katoh K., Misawa K., Kuma K.I. & Miyata T. 2002. MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research* 30: 3059–3066. <https://doi.org/10.1093/nar/gkf436>
- Katoh K., Rozewicki J. & Yamada K.D. 2017. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in Bioinformatics* 20 (4): 1160–1166. <https://doi.org/10.1093/bib/bbx108>

- Kayastha P., Berdi D., Mioduchowska M., Gawlak M., Łukasiewicz A., Gołdyn B. & Kaczmarek Ł. 2020. Some tardigrades from Nepal (Asia) with integrative description of *Macrobiotus wandae* sp. nov. (Macrobiotidae: *hufelandi* group). *Annales Zoologici* 70 (1): 121–142. <https://doi.org/10.3161/00034541ANZ2020.70.1.007>
- Kumar A., Sharma P., Gomar Alba M., Shcheprova Z., Daulny A., Sanmartín T., Matucci I., Funaya C., Beato M. & Mendoza M. 2018. Daughter-cell-specific modulation of nuclear pore complexes controls cell cycle entry during asymmetric division. *Nature Cell Biology* 20 (4):432–442. <https://doi.org/10.1038/s41556-018-0056-9>
- McInnes S.J. 1994. Zoogeographic distribution of terrestrial/freshwater tardigrades from current literature. *Journal of the Natural History* 28: 257–352. <https://doi.org/10.1080/00222939400770131>
- McInnes S.J., Michalczyk Ł. & Kaczmarek Ł. 2017. Annotated zoogeography of non-marine Tardigrada. Part IV: Africa. *Zootaxa* 4284: 1–71. <https://doi.org/10.11646/zootaxa.4284.1.1>
- Michalczyk Ł. & Kaczmarek Ł. 2003. A description of the new tardigrade *Macrobiotus reinhardti* (Eutardigrada, Macrobiotidae, *harmsworthi* group) with some remarks on the oral cavity armature within the genus *Macrobiotus* Schultze. *Zootaxa* 331: 1–24. <https://doi.org/10.11646/zootaxa.331.1.1>
- Miller M.A., Pfeiffer W. & Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. *2010 Gateway Computing Environments Workshop (GCE)*: 1–8. <https://doi.org/10.1109/GCE.2010.5676129>
- Múrias dos Santos A., Cabezas M.P., Tavares A.I., Xavier R. & Branco M. 2016. tcsBU: a tool to extend TCS network layout and visualization. *Bioinformatics* 32 (4): 627–628. <https://doi.org/10.1093/bioinformatics/btv636>
- Nelson D.R., Guidetti R. & Rebecchi L. 2015. Phylum Tardigrada. In: Thorp J. & Rogers D.C. (eds) *Ecology and General Biology: Thorp and Covich's Freshwater Invertebrates*: 347–380. Academic Press, London. <https://doi.org/10.1016/B978-0-12-385026-3.00017-6>
- Nowak B. & Stec D. 2018 An integrative description of *Macrobiotus hanna*e sp. nov. (Tardigrada: Eutardigrada: Macrobiotidae: *hufelandi* group) from Poland. *Turkish Journal of Zoology* 42: 269–286. <https://doi.org/10.3906/zoo-1712-31>
- Nylander J.A.A. 2004. MrModeltest v2. Program distributed by the author. Evolutionary Biology Centre, Uppsala University.
- Pilato G. 1981. Analisi di nuovi caratteri nello studio degli Eutardigradi. *Animalia* 8: 51–57.
- Pilato G. & Binda M.G. 2010. Definition of families, subfamilies, genera and subgenera of the Eutardigrada, and keys to their identification. *Zootaxa* 2404: 1–54. <https://doi.org/10.11646/zootaxa.2404.1.1>
- Pilato G. & Lisi O. 2009. Tardigrades of the Seychelles Islands, with the description of three new species. *Zootaxa* 2124: 1–20. <https://doi.org/10.5281/zenodo.188178>
- Pilato G., Binda M.G. & Azzaro M. 1990. Tardigradi di Terra del Fuoco e Magallanes. III. *Macrobiotus punctillus*, nuova specie di Macrobiotidae del gruppo *hufelandi*. *Animalia* 17: 123–129.
- Pilato G., Binda M.G. & Catanzaro R. 1991. Remarks on some tardigrades of the African fauna with the description of three new species of *Macrobiotus* Schultze 1834. *Tropical Zoology* 4: 167–178. <https://doi.org/10.1080/03946975.1991.10539487>
- Pilato G., Kiosya Ye., Lisi O. & Sabella G. 2012. New records of Eutardigrada from Belarus with the description of three new species. *Zootaxa* 3179: 39–60. <https://doi.org/10.11646/zootaxa.3179.1.2>

- Pilato G., Sabella G., D'Urso V. & Lisi O. 2017. *Macrobotus nebrodensis* and *Adropion vexatum*, two new species of Eutardigrada (Tardigrada, Parachela) from Sicily. *Zootaxa* 4362 (2): 267–279. <https://doi.org/10.11646/zootaxa.4362.2.6>
- Pilato G., D'Urso V., Sabella G. & Lisi O. 2019. A new species of isohypsibiid (Eutardigrada) from Sicily. *Zootaxa* 4608 (3): 543–554. <https://doi.org/10.11646/zootaxa.4608.3.8>
- Puillandre N., Lambert A., Brouillet S. & Achaz G. 2012. ABGD, Automatic Barcode Gap Discovery for primary species delimitation. *Molecular Ecology* 21: 1864–1877. <https://doi.org/10.1111/j.1365-294X.2011.05239.x>
- Ramazzotti G. & Maucci W. 1983. Il Phylum Tardigrada. *Memorie dell'Istituto Italiano di Idrobiologia, Pallanza* 41: 1–1012.
- Rambaut A., Drummond A.J., Xie D., Baele G. & Suchard M.A. 2018. Posterior summarisation in Bayesian phylogenetics using Tracer 1.7. *Systematic Biology* 67 (5): 901–904. <https://doi.org/10.1093/sysbio/syy032>
- Rebecchi L. & Bertolani R. 1988. New cases of parthenogenesis and polyploidy in the genus *Ramazzottius* (Tardigrada, Hypsibiidae) and a hypothesis concerning their origin. *International Journal of Invertebrate Reproduction and Development* 14: 187–196. <https://doi.org/10.1080/01688170.1988.10510377>
- Richters F. 1926. Tardigrada. In: Kukenthal W. & Krumbach T. (eds) *Handbuch der Zoologie*, Vol. III: 58–61. Walter de Gruyter & Co, Berlin and Leipzig.
- Ronquist F. & Huelsenbeck J.P. 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574. <https://doi.org/10.1093/bioinformatics/btg180>
- Roszkowska M., Ostrowska M., Stec D., Janko K. & Kaczmarek Ł. 2017. *Macrobotus polypiformis* sp. nov., a new tardigrade (Macrobotidae; *hufelandi* group) from the Ecuadorian Pacific coast, with remarks on the claw abnormalities in eutardigrades. *European Journal of Taxonomy* 327: 1–19. <https://doi.org/10.5852/ejt.2017.327>
- Schill R.O. (ed.) 2018. *Water Bears: The Biology of Tardigrades*. Vol. 419. Springer, Cham, Switzerland.
- Schill R.O., Förster F., Dandekar T. & Wolf M. 2010. Using compensatory base change analysis of internal transcribed spacer 2 secondary structures to identify three new species in *Paramacrobotus* (Tardigrada). *Organisms Diversity and Evolution* 10: 287–296. <https://doi.org/10.1007/s13127-010-0025-z>
- Schultze C.A.S. 1834. *Macrobotus Hufelandii animal e crustaceorum classe novum, reviviscendi post diuturnam asphyxia et ariditatem potens, etc.* C. Curths, Berlin.
- Schuster R.O., Nelson D.R., Grigarick A.A. & Christenberry D. 1980. Systematic criteria of the Eutardigrada. *Transaction of the American Microscopical Society* 99: 284–303. <https://doi.org/10.2307/3226004>
- Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22: 2688–2690. <https://doi.org/10.1093/bioinformatics/btl446>
- Stamatakis A., Hoover P. & Rougemont J. 2008. A rapid bootstrap algorithm for the RAxML web servers. *Systematic Biology* 57: 758–771. <https://doi.org/10.1080/10635150802429642>
- Stambouli-Essassi S., Roche E. & Bouzid S. 2007. Evolution of vegetation and climatic changes in North-Western Tunisia during the last 40 millennia. *Geo-Eco-Trop* 31: 171–214.
- Stec D., Smolak R., Kaczmarek Ł. & Michalczyk Ł. 2015. An integrative description of *Macrobotus paulinae* sp. nov. (Tardigrada: Eutardigrada: Macrobotidae: *hufelandi* group) from Kenya. *Zootaxa* 4052 (2): 501–526. <https://doi.org/10.11646/zootaxa.4052.5.1>

- Stec D., Morek W., Gąsiorek P., Blagden B. & Michalczyk Ł. 2017. Description of *Macrobotus scoticus* sp. nov. (Tardigrada: Macrobiotidae: *hufelandi* Group) from Scotland by means of integrative taxonomy. *Annales Zoologici* 67 (2): 181–197. <https://doi.org/10.3161/00034541ANZ2017.67.2.001>
- Stec D., Arakawa K. & Michalczyk Ł. 2018a. An integrative description of *Macrobotus shonaicus* sp. nov. (Tardigrada: Macrobiotidae) from Japan with notes on its phylogenetic position within the *hufelandi* group. *PLoS ONE* 13 (2): e0192210. <https://doi.org/10.1371/journal.pone.0192210>
- Stec D., Kristensen R.M. & Michalczyk Ł. 2018b. Integrative taxonomy identifies *Macrobotus papei*, a new tardigrade species of the *Macrobotus hufelandi* complex (Eutardigrada: Macrobiotidae) from the Udzungwa Mountains National Park (Tanzania). *Zootaxa* 4446 (2): 273–291. <https://doi.org/10.11646/zootaxa.4446.2.7>
- Stec D., Krzywański Ł. & Michalczyk Ł. 2018c. Integrative description of *Macrobotus canaricus* sp. nov. with notes on *M. recens* (Eutardigrada: Macrobiotidae). *European Journal of Taxonomy* 452: 1–36. <https://doi.org/10.5852/ejt.2018.452>
- Stec D., Morek W., Gąsiorek P. & Michalczyk Ł. 2018d. Unmasking hidden species diversity within the *Ramazzottius oberhaeuseri* complex, with an integrative redescription of the nominal species for the family Ramazzottiidae (Tardigrada: Eutardigrada: Parachela). *Systematics and Biodiversity* 16 (4): 357–376. <https://doi.org/10.1080/14772000.2018.1424267>
- Stec D., Dudziak M. & Michalczyk Ł. 2020a. Integrative descriptions of two new Macrobiotidae species (Tardigrada: Eutardigrada: Macrobiotidae) from French Guiana and Malaysian Borneo. *Zoological Studies* 59 (23): 1–25. <https://doi.org/10.6620/ZS.2020.59-23>
- Stec D., Tumanov D.V. & Kristensen R.M. 2020b. Integrative taxonomy identifies two new tardigrade species (Eutardigrada: Macrobiotidae) from Greenland. *European Journal of Taxonomy* 614: 1–40. <https://doi.org/10.5852/ejt.2020.614>
- Stec D., Vecchi M., Calhim S. & Michalczyk Ł. 2021. New multilocus phylogeny reorganises the family Macrobiotidae (Eutardigrada) and unveils complex morphological evolution of the *Macrobotus hufelandi* group. *Molecular Phylogenetics and Evolution* 160. <https://doi.org/10.1016/j.ympev.2020.106987>
- Sugiura K., Arakawa K. & Matsumoto M. 2020. Distribution of *Macrobotus shonaicus* Stec, Arakawa & Michalczyk, 2018 (Tardigrada: Eutardigrada: Macrobiotidae) in Japan. *Zootaxa* 4767 (1): 56–70. <https://doi.org/10.11646/zootaxa.4778.3.11>
- Tang C.Q., Humphreys A.M., Fontaneto D. & Barraclough T.G. 2014. Effects of phylogenetic reconstruction method of the robustness of species delimitation using single-locus data. *Methods in Ecology and Evolution* 5: 1086–1094. <https://doi.org/10.1111/2041-210X.12246>
- Templeton A.R., Crandall K.A. & Sing C.F. 1992. A cladistic analysis of phenotypic association with haplotypes inferred from restriction endonuclease mapping and DNA sequence data. III. Cladogram estimation. *Genetics* 132 (2): 619–633.
- Thulin G. 1928. Über die Phylogenie und das System der Tardigraden. *Hereditas* 11 (2–3): 207–266. <https://doi.org/10.1111/j.1601-5223.1928.tb02488.x>
- Vecchi M., Cesari M., Bertolani R., Jönsson K.I., Rebecchi L. & Guidetti R. 2016. Integrative systematic studies on tardigrades from Antarctica identify new genera and new species within Macrobiotidae and Echiniscoidea. *Invertebrate Systematics* 30: 303–322. <https://doi.org/10.1071/IS15033>
- Vicente F., Cesari M., Serrano A. & Bertolani R. 2013. The impact of fire on terrestrial tardigrade biodiversity: a first case-study from Portugal. *Journal of Limnology* 72 (s1): 152–159. <https://doi.org/10.4081/jlimnol.2013.s1.e19>

Welnicz W., Grohme M.A., Kaczmarek Ł., Schill R.O. & Frohme L. 2011. ITS-2 and 18S rRNA data from *Macrobiotus polonicus* and *Milnesium tardigradum* (Eutardigrada, Tardigrada). *Journal of Systematics and Evolutionary Research* 49 (s1): 34–39. <https://doi.org/10.1111/j.1439-0469.2010.00595.x>

Zhang J., Kapli P., Pavlidis P. & Stamatakis A. 2013. A general species delimitation method with applications to phylogenetic placements. *Bioinformatics* 29: 2869–2876. <https://doi.org/10.1093/bioinformatics/btt499>

Manuscript received: 1 September 2020

Manuscript accepted: 16 April 2021

Published on: 9 July 2021

Topic editor: Rudy Jocqué

Section editor: Daniel Stec

Desk editor: Pepe Fernández

Printed versions of all papers are also deposited in the libraries of the institutes that are members of the *EJT* consortium: Muséum national d'histoire naturelle, Paris, France; Meise Botanic Garden, Belgium; Royal Museum for Central Africa, Tervuren, Belgium; Royal Belgian Institute of Natural Sciences, Brussels, Belgium; Natural History Museum of Denmark, Copenhagen, Denmark; Naturalis Biodiversity Center, Leiden, the Netherlands; Museo Nacional de Ciencias Naturales–CSIC, Madrid, Spain; Real Jardín Botánico de Madrid CSIC, Spain; Zoological Research Museum Alexander Koenig, Bonn, Germany; National Museum, Prague, Czech Republic.

Supp. file 1. Single measurements (in μm) and *pt* values of morphological structures of the holotype and paratypes of *Macrobiotus azzunae* sp. nov. <https://doi.org/10.5852/ejt.2021.758.1429.4625>

Supp. file 2. Buccal-pharyngeal apparatus of a hologenophore voucher specimen (C4218 V7) of *Macrobiotus azzunae* sp. nov. in vivo, PhC. <https://doi.org/10.5852/ejt.2021.758.1429.4627>

Supp. file 3. Alignment of all *Macrobiotus hufelandi* group sequences for the 18S gene. <https://doi.org/10.5852/ejt.2021.758.1429.4629>

Supp. file 4. Alignment of all *Macrobiotus hufelandi* group sequences for the 28S gene. <https://doi.org/10.5852/ejt.2021.758.1429.4631>

Supp. file 5. Alignment of all *Macrobiotus hufelandi* group sequences for the ITS-2 gene. <https://doi.org/10.5852/ejt.2021.758.1429.4633>

Supp. file 6. Alignment of all *Macrobiotus hufelandi* group sequences for the COI gene. <https://doi.org/10.5852/ejt.2021.758.1429.4635>

Supp. file 7. Genetic distances (p-distance) computed between species of the *Macrobiotus hufelandi* group for the 18S gene. <https://doi.org/10.5852/ejt.2021.758.1429.4637>

Supp. file 8. Genetic distances (p-distance) computed between species of the *Macrobiotus hufelandi* group for the 28S gene. <https://doi.org/10.5852/ejt.2021.758.1429.4639>

Supp. file 9. Genetic distances (p-distance) computed between species of the *Macrobiotus hufelandi* group for the ITS-2 gene. <https://doi.org/10.5852/ejt.2021.758.1429.4641>

Supp. file 10. Genetic distances (p-distance) computed between species of the *Macrobiotus hufelandi* group for the COI gene. <https://doi.org/10.5852/ejt.2021.758.1429.4643>