Redefinition of the sexstrigatus group of Lasioglossum (Hemihalictus) Cockerell, 1897 (Hymenoptera, Apoidea, Halictidae), with a revision of Japanese species

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Abstract. I propose a redefinition of the sexstrigatus group of Lasioglossum (Hemihalictus) Cockerell, 1897, including a phylogenetic analysis. This group is characterised by a combination of the following 12 characteristics: male antenna short, not attaining to metasoma, male labrum with distal process and well-developed basal elevation, male head with genal process as variation, female mesepisternum reticulate-punctate on lower area, mesepisternum without tubercle in both sexes, female metasomal terga with distinct fimbriae on posterior margin, male S8 with well-developed median process, gonobase ventral arm of male genitalia connected with each other at upper ends, gonocoxite of male genitalia smooth, gonostylus of male genitalia small and simple, bud-like, and the ventral retrorse lobe of male genitalia not attaining to gonobase. The Japanese species of the sexstrigatus group are revised. Thirteen species in total are recognised, including three new species: Lasioglossum (Hemihalictus) ikudomei sp. nov., L. (H.) spectrum sp. nov., and L. (H.) subsimplicior sp. nov. Lasioglossum (Hemihalictus) perplexans (Cockerell, 1925) is synonymised under L. (H.) kiautschouense (Strand, 1910). A key to the Japanese species is provided. Bionomical data, such as flight and flower records or habitat, are reported for some species. The distributions of all species are mapped. DNA sequences including a part of the barcode region are given for L. (H.) kiautschouense, L. (H.) ohei Hirashima & Sakagami, 1966, L. (H.) speculinum (Cockerell, 1925), L. (H.) spectrum sp. nov., L. (H.) subsimplicior sp. nov., and L. (H.) taeniolellum (Vachal, 1903).

Keywords. Halictidae, Lasioglossum, sexstrigatus group, Eastern Asia.

Introduction

Hemihalictus Cockerell, 1897 is a subgenus of Lasioglossum Curtis, 1833 (Halictidae Thomson, 1869, Halictinae Thomson, 1869), with approximately 250 species worldwide (Ascher & Pickering 2020). Lasioglossum (Hemihalictus) was considered monotypic (Michener 2007) until it was reclassified by a recent study (Gibbs et al. 2013) as a group that includes L. (Hemihalictus) s. str., L. (Studila) Cameron, 1898, and the black L. (Dialictus) Robertson, 1902, as classified by Michener (2007). The
composition of *L. (Hemihalictus)* sensu Gibbs et al. (2013) considerably differs from the systematics of Palearctic researchers (e.g., Ebmer 1976, 1997, 2000; Ebmer & Sakagami 1985; Ebmer et al. 1994; Pesenko 2007a). In Ebmer’s system, the Palearctic species of *L. (Hemihalictus)* sensu Gibbs et al. (2013) correspond to the carinateless *L. (Evylaeus)* Robertson, 1902. In the system of Pesenko (2007a), *Evylaeus* is treated as a valid genus with 12 subgenera (*E. (Limbevylaeus)* Pesenko, 2007, *E. (Crassevylaeus)* Pesenko, 2007, *E. (Laevinodilaeus)* Pesenko, 2007, *E. (Pallidevylaeus)* Pesenko, 2007, *E. (Pauphalictus)* Warncke, 1975, *E. (Marghalictus)* Warncke, 1975, *E. (Microhalictus)* Warncke, 1975, *E. (Nitidiusculaeus)* Pesenko, 2007, *E. (Rostrohalictus)* Warncke, 1975, *E. (Truncevylaeus)* Pesenko, 2007) corresponding to *L. (Hemihalictus)* sensu Gibbs et al. (2013). In the present study, I follow the recent systematics of *Lasioglossum* proposed by Gibbs et al. (2013). Many species groups have been recognised in the Palearctic *L. (Hemihalictus)* by various researchers (e.g., Blüthgen 1934; Ebmer 1975, 1986; Warncke 1975; Ebmer & Sakagami 1985; Sakagami & Ebmer 1996). At present, these species groups have been reclassified by Pesenko (2007a) into the following 12 groups: 1) *clypearis* group, 2) *crassepunctatum* group, 3) *laevinode* group, 4) *limbellum* group, 5) *longirostre* group, 6) *nitidiusculum* group, 7) *marginellum* group, 8) *pallidum* group, 9) *pauperatum* group, 10) *semilucens* group, 11) *sexstrigatus* group, and 12) *villosulum* group. These groups are treated as being of subgeneric status in Pesenko (2007a). The species of *L. (Hemihalictus)* are morphologically similar to each other, therefore, even in the Pesenko’s system, there remains ambiguity in the definition of each group.

One of these, the *sexstrigatus* group, includes over 30 species and is diverse in eastern Asia (Pesenko 2007b; Murao et al. 2010). This group is mainly characterised by the female metasomal terga with distinct fimbriae on the lateral-apical margin and the male head with a size-linked gigantism with genal process (Sakagami & Ebmer 1996). However, some species in the group do not necessarily share these characteristics (Murao 2017a). This group is also quite difficult to identify at the species level in *Lasioglossum* because identification is often based on subtle morphological differences (e.g., Sakagami & Ebmer 1996). A few species are known as solitary or communal (Sakagami et al. 1966; Sakagami 1992), but no eusocial species are known. Some species often dominate in the Japanese bee fauna when considering the number of individuals (Ikudome 1995; Iwata 1997; Minagi et al. 2000; Negoro 2001a, 2001b; Maeta et al. 2003; Gôukon 2006; Hisamatsu & Yamane 2006; Hisamatsu 2011, 2017; Murao, Murase & Iwata unpublished).

Twenty-two species of the *sexstrigatus* group have been partially described or recorded in Japan by various researchers (Smith 1873; Vachal 1903; Strand 1910; Blüthgen 1925; Sakagami et al. 1966; Usui et al. 1976; Takahashi & Sakagami 1993; Ebmer et al. 1994; Sakagami & Tadauchi 1995; Sakagami & Ebmer 1996; Murao et al. 2010; Murao 2012).

As a first step in the taxonomic study of the *sexstrigatus* group in Asia, this study revises the definition of this group, including a phylogenetic analysis based on the morphological characteristics and the Japanese fauna.

**Material and methods**

**Collection**

This study is based on the specimens deposited in the following institutions and personal collections, which are referred to using the following abbreviations:

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
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<tbody>
<tr>
<td>AETU</td>
<td>Applied Entomological Laboratory, Tokai University, Kumamoto, Japan</td>
</tr>
<tr>
<td>AMNH</td>
<td>American Museum of Natural History, New York, USA</td>
</tr>
<tr>
<td>cGou</td>
<td>private collection of Mr Katsu Gôukon, Miyagi, Japan</td>
</tr>
<tr>
<td>cMur</td>
<td>the first author’s private collection, Fukuoka, Japan</td>
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</table>
MURAO R., Redefinition and revision of the *Lasioglossum sexstrigatus* group in Japan

ELKU = Entomological Laboratory, Faculty of Agriculture, Kyushu University, Fukuoka, Japan
KWC = Laboratory of Biology, Kagoshima Women’s College, Kagoshima, Japan
MNHAH = the late Dr Shoichi F. Sakagami’s collection, deposited in the Museum of Nature and Human Activities, Hyogo, Japan
SULE = Dr Yasuo Maeta’s collection, deposited in the Laboratory of Insect Ecology, Faculty of Life and Environmental Science, Shimane University, Shimane, Japan
USNM = National Museum of Natural History, Washington DC, USA
ZMHB = Museum für Naturkunde an der Humboldt Universität zu Berlin, Germany

Terminology
Terminology and style used in the description follows Murao et al. (2015b). Abbreviations used in the text are as follows:

AOD = antennocular distance (shortest distance between outer margin of antennal socket and inner margin of compound eye)
BL = body length (from antennal base to tip of pygidial plate)
CAL = clypealveolar distance (between lower margin of antennal socket and lower margin of supra-clypeus in frontal view)
CPL = clypeal length (between upper and lower margins of clypeus in frontal view)
EL = eye length
EW = eye width (maximum length and width of the compound eye)
Fn = n° antennal flagellomere
FnL = length of n° flagellomere (measured along the ventral surface)
FnW = width of n° flagellomere (measured from dorsal and ventral surfaces of flagellomere)
GW = genal width (maximum width of the genal area when seen in lateral view)
HL = head length (from top of vertex to lower margin of clypeus)
HW = head width (between outer margins of compound eyes in frontal view)
IAD = interantennal distance (between inner margins of antennal sockets)
IOD = interocellar distance (between lateral ocelli)
IS = interspace between punctures (e.g., IS 0.5 d means ½ of the diameter of a puncture)
LOD = lower interorbital distance
MNL = metanotal length
MOD = maximum interorbital distance
MPL = metapostnotal length
MsW = maximum mesosomal width
MtW = maximum metasomal width
OCD = ocellocapital distance (shortest distance between margins of lateral ocellus and vertex when seen in upper view)
OOD = ocellocular distance (shortest distance between lateral ocellus and inner margin of compound eye)
PP = punctures
SCL = mesoscutellar length
Sn = n° metasomal sternum
SPL = scape length (a straight line from base to tip of scape)
Tn = n° metasomal tergum
UOD = upper interorbital distance
WL = wing length (length of fore wing from the apical point to the base including tegula)

Flower records
Flower records visited by each species are based on specimen label data. The scientific names of flowering plants visited by bees are cited from Yonekura & Kajita (2003-).
**Distribution maps**

Distribution maps were made by QGIS ver. 3.6 (QGIS Development Team 2020) using map data download from Natural Earth (https://www.naturalearthdata.com/). The point data for each map are based mainly on the examined specimens, but also partially on the following publications: Sakagami & Tadauchi (1995), Sakagami & Ebmer (1996), Goubara & Maeta (2002), Goubara et al. (2002), Haneda & Tano (2003), Ikudome (2005), Gôukon (2006), Yamane & Ikudome (2008), Ikudome & Yamane (2009), Fukasawa & Miyano (2010), Murao (2015), and Hiraiwa & Ushimaru (2017). Locality data which lacked both latitude and longitude were geocoded by GeoNames (https://www.geonames.org/).

**Cladistic analysis**

To test monophyly of the *sexstrigatus* group, I selected two to 16 species from each of 10 species groups of *Lasioglossum* (*Hemihalictus*) as the ingroup and two species of *L. (Dialictus)* as an outgroup (Appendix 1). A total of 32 morphological characters were coded using MESQUITE ver. 3.61 (Maddison & Maddison 2019) (Appendix 2). Characters are coded as ‘0’, ‘1’, ‘2’ or ‘3’ and the data matrix is shown in Appendix 3. A cladistic analysis was performed using TNT ver. 1.1 (Goloboff et al. 2008) traditional heuristic search that generated 1000 Wagner trees with random addition sequence. Symmetric resampling was performed using default settings for 1000 replicates. Unambiguous character state changes were mapped on the most parsimonious trees using WINCLADA ver. 1.00.88 (Nixon 2002).

**DNA analysis**

DNA extraction and PCR were conducted at the Kyushu University Museum (Fukuoka, Japan). DNA was extracted using a DNeasy Blood and Tissue kit (Qiagen, Tokyo, Japan) following the manufacture’s instructions. A 707 bp including a part of DNA barcode region of the cytochrome oxidase subunit I (COI) gene fragment of mtDNA was amplified (using the primers COI_pF2 and COI_2437d), purified, and electrophoresed following the methods described by Murao et al. (2015c). DNA sequencing was outsourced to the FASMAC Co., Ltd (Kanagawa, Japan). The sequences analyzed in the present study are deposited in GenBank through the DNA Database of Japan (DDBJ). Appendix 4 lists the GenBank accession numbers used in the present paper. Pairwise sequence divergences within each specis of *sexstrigatus* group were calculated using Kimura 2-parameter distance (Kimura 1980). The analyses were conducted using MEGA5 (Tamura et al. 2011).

**Results**

**Phylogenetic analysis**

The phylogenetic analysis resulted in a most parsimonious tree (L=137, Ci=33, Ri=57) (Fig. 1). In the present analysis, monophyly of the *sexstrigatus* group sensu Sakagami & Ebmer (1996), Pesenko (2007a), and Murao et al. (2010) is not supported and members are split into at least two clades: the *sexstrigatus* clade, comprising *Lasioglossum* (*Hemihalictus*) amamiense Ebmer & Sakagami, 1994, *L. (H.) frigidum* Sakagami & Ebmer, 1996, *L. (H.) ohei* Hirashima & Sakagami, 1966, *L. (H.) sexstrigatus* (Schenck, 1869), *L. (H.) simplicior* (Cockerell, 1931), *L. (H.) smilodon* Ebmer & Sakagami, 1994, *L. (H.) spectrum* sp. nov., *L. (H.) sphecodicolor* Sakagami & Tadauchi, 1995, and *L. (H.) taeniolellum* (Vachal, 1903) and the *japonicum* clade, comprising *L. (H.) canaliculatum* Murao, 2010, *L. (H.) donanense* Murao, 2010, *L. (H.) japonicum* (Dalla Torre, 1896), *L. (H.) urumaense* Murao, 2010, *L. (H.) yonaguniense* Murao, 2010, and *L. (H.) zipangu* Ebmer & Sakagami, 1994. The *sexstrigatus* clade is defined by six homoplasious synapomorphies: 1) the male head with genal process (variable with allometry) (3:0); 2) the distal process of male labrum present (13:0); 3) the female metasomal terga with filmbriae on apical margin (21:0) (secondarily lost in *L. (H.) sphecodicolor*); 4) the male S8 with developed median process (23:0); 5) gonobase ventral arms of male genitalia connected with each other at upper ends (24:0); and 6) the ventral retrorse lobe of male genitalia not reaching gonobase (28:1). I here propose a
Fig. 1. A most parsimonious tree resulting from analysis of the matrix in Appendix 3. Black circles indicate non-homoplasic changes, white circles indicate homoplasious change. For characters and character states see Appendix 2.
revised classification that the *sexstrigatus* group is restricted to species sharing the characteristics of the *sexstrigatus* clade.

The *japonicum* clade is also proposed here as a new species group (as the *japonicum* group) because its monophyly is supported by three synapomorphies, one of which, male S4–S5 latero-posteriorly with thin hair tufts (22:1) is non-homoplasious. For the remaining nine species groups, monophyly is supported for the following groups in the present analysis: the *alphenum* group (= *L. (Sudila])*), *nitidiusculum* group, *marginellum* group, *villosulum* group, and the *clypearis + longirostre* groups clade.

**Taxonomy**

Class Insecta Linnaeus, 1758  
Order Hymenoptera Linnaeus, 1758  
Superfamily Apoidea Latreille, 1802  
Family Halictidae Thomson, 1869  
Subfamily Halictinae Thomson, 1869  
Tribe Halictini Thomson, 1869  
Genus *Lasioglossum* Curtis, 1833

*Hemihalictus* Cockerell, 1897: 288 (also published as new by Cockerell 1898) (type species: *Panurgus lustrans* Cockerell, 1897, by original designation).

*Sudila* Cameron, 1898: 52 (type species: *Sudila bidentata* Cameron, 1898, by designation of Sandhouse 1943).

*Prosopalictus* Strand, 1913: 26 (type species: *Prosopalictus micans* Strand, 1913, by original designation and monotypy).

*Ceylonicola* Friese, 1918: 501 (type species: *Ceylonicola atra* Friese, 1918 = *Sudila bidentata* Cameron, 1898, by designation of Sandhouse 1943).


*Halictus* (*Marghalictus*) Warncke, 1975: 95 (type species: *Hylaeus marginellus* Schenck, 1853, by original designation).


**Diagnosis**

In eastern Asia, the species of *Hemihalictus* series are classified into four subgenera (*Acanthalictus*, *Dialictus*, *Hemihalictus*, and *Sphecodogastra*). The subgenus *Hemihalictus* is separated from the other three subgenera by the following key:

1. Lower margin of male clypeus deflected forward; female labrum without basal elevation; basal elevation of male labrum stongly swollen, with longitudinal furrow; female mandible with two preapical teeth; male S2 gently swollen on apical part; male S6 expanded apically (Murao et al. 2014: figs 5–8, 15, 20) ................................................................. *Acanthalictus* Cockerell, 1924
   - Male clypeus normal, not deflected; female labrum with basal elevation (as in Fig. 4D); male labrum without basal elevation or with low basal elevation; female mandible with a preapical tooth; male S2 flat; male S6 straight apically ........................................ 2

2. Head and mesosoma with brilliant or dull green-metallic luster in both sexes (mesepisternum with shallow PP or reticulate PP in both sexes, see Murao et al. 2015b: figs 1e, 6e, 20e) ................................................................. *Dialictus* Robertson, 1902
   - Body generally black ........................................................................................................ 3

3. Posterior surface of propodeum generally with complete lateral carina (Murao 2017a: fig. 3b); mesepisternum with coarse reticulate-rugulae over entire surface (Murao 2017a: fig. 2c)................................................................. *Sphecodogastra* Ashmead, 1899
   - Posterior surface of propodeum with incomplete lateral carina not reaching metapostnotum (Murao 2017a: fig. 3c); mesepisternum with shallow or reticulate PP (Fig. 2E; Murao 2017b: fig. 1d) ................................. *Hemihalictus* Cockerell, 1897

Three species, *L.* (*Sphecodogastra*) boreale Svensson, Ebmer & Sakagami, 1977, *L.* (*S.*) solisortus Ebmer & Maeta, 1994, and *L.* (*S.*) subtropicum Sakagami, Miyanaga & Maeta, 1994, from Japan are similar to *L.* (*Hemihalictus*) by having the posterior surface of the propodeum with an incomplete lateral carina (sometimes as variation) (Murao & Tadauchi 2007). However, these species can be separated from *L.* (*Hemihalictus*) by the mesepisternum with coarse reticulate-rugulae.

**The *sexstrigatus* group**

**Diagnosis**

Species of the *sexstrigatus* group are characterized by a combination of the following characters: 1) male antenna short, not reaching metasoma (Fig. 2A); 2) male labrum with well-developed basal elevation (Fig. 2B) (except for *Lasioglossum* (*Hemihalictus*) ohei Hirashima & Sakagami, 1966); 3) male labrum with distal process (Fig. 2B); 4) male head with genal process as variation (Fig. 2C–D); 5) female mesepisternum reticulate-punctate on lower area (Fig. 2E); 6) mesepisternum without tubercle in both sexes; 7) female metasomal terga with distinct fimbriae on posterior margin (Fig. 2F) (except for *L.* (*H.*) sphecodicolor Sakagami & Tadauchi, 1995); 8) male S8 with well-developed (over S7) median process (Fig. 3A); 9) gonobase ventral arms of male genitalia connected with each other at upper ends (Fig. 3C); 10) gonocoxite of male genitalia smooth (Fig. 3B–D); 11) gonostylus of male genitalia small and simple, bud-like (Fig. 3B–C); and 12) the ventral retrorse lobe of male genitalia not reaching gonobase (Fig. 3B–C).
Fig. 2. A, E. Lasioglossum (Hemihalictus) sexstrigatus (Schenck, 1869). A. ♂, lateral habitus. E. ♀, mesepisternum. — B. L. (H.) taeniolellum (Vachal, 1903), ♂, labrum. — C, F. L. (H.) frigidum Sakagami & Ebmer, 1996. C. ♂, cephalic polymorphism. F. ♀, metasomal terga. — D. L. (H.) simplicior (Cockerell, 1931), ♂, cephalic polymorphism. Scale bars: A = 2 mm; B = 0.2 mm; C, E = 0.5 mm; D, F = 1 mm.
Fig. 3. *Lasioglossum (Hemihalictus) sexstrigatus* (Schenck, 1869), ♂. A. S7–S8. B. Genitalia in lateral view. C. Genitalia in ventral view. D. Genitalia in dorsal view. Scale bars: 0.1 mm.
The Japanese species of *Hemihalictus* are classified into five species groups (*nitidiusculum*, *japonicum*, *semilucens*, *sexstrigatus*, and *villosulum* groups). The *sexstrigatus* group is separated from the other four groups by the male head with genal process, the female metasomal terga generally with distinct fimbriae on posterior margin, and male S8 with developed median process.

**Variation** (male cephalic polymorphism)

The members of *sexstrigatus* group except for *Lasioglossum (Hemihalictus) frigidum* Sakagami & Ebmer, 1996 display male cephalic polymorphism (Sakagami *et al*. 1966; Ebmer *et al*. 1994; Sakagami & Ebmer 1996; Murao *et al*. 2010). This polymorphism is caused by the allometric development of the head (Sakagami *et al*. 1966). The presence of a genal process is characteristic of the *sexstrigatus* group, but is not known from the *japonicum* group (Sakagami *et al*. 1966; Ebmer *et al*. 1994; Sakagami & Tadauchi 1995; Murao *et al*. 2010). Male cephalic polymorphism with allometric variation is known to occur in various bee families such as Andrenidae Latreille, 1802, Colletidae Lepeletier, 1841, and Halictidae Thomson, 1869 (summarised in Danforth *et al*. 2019). It also appears that such male cephalic polymorphism often occurs in communal species (Maeta 2000; Danforth *et al*. 2019). In the Japanese species of the *sexstrigatus* group, *L. (H.) ohei* has indeed been reported as a communal species (Sakagami *et al*. 1966). The other Japanese species with male cephalic polymorphism may also be communal.

**Distribution**

This group is distributed from the Palearctic to northern Oriental Region. It is diverse in eastern Asia.

**Comments**

*Lasioglossum sexstrigatum* was originally described as *Halictus sexstrigatus* by Schenck. From the scientific name ‘-strigatus’, the original spelling was retained in accordance with Article 31.2.2 of the ICZN (International Commission on Zoological Nomenclature) Code.

**Species included in Japan**

1) *L. amamiense* Ebmer & Sakagami, 1994  
2) *L. frigidum* Sakagami & Ebmer, 1996  
3) *L. ikudomei* sp. nov.  
4) *L. kiautschouense* (Strand, 1910)  
5) *L. ohei* Hirashima & Sakagami, 1966  
6) *L. simplicior* (Cockerell, 1931)  
7) *L. smilodon* Ebmer & Sakagami, 1994  
8) *L. spectrum* sp. nov.  
9) *L. speculunum* (Cockerell, 1925)  
10) *L. sphecodicolor* Sakagami & Tadauchi, 1995  
11) *L. subsimplicior* sp. nov.  
12) *L. tadauchii* Murao, 2012  
13) *L. taeniolellum* (Vachal, 1903)

As stated above, these species, except for *L. (H.) bicornutum*, *L. (H.) kankauchare*, *L. (H.) latifacies*, and *L. (H.) silvicolum*, form a separate clade as the *japonicum* group (Fig. 1). Both *L. (H.) latifacies* and *L. (H.) silvicolum* are included in the *japonicum* group because males (undescribed) of both species share a non-homoplasious syapomorphy of this group (Murao unpublished). The Japanese specimens of *L. (H.) kankauchare* recorded by Blüthgen (1925, as *Halictus kankaucharis*) have been preserved in ZMHB. I visited ZMHB in 2012 to examine the bee specimens from the Oriental Region. At that time, I also examined the Japanese specimens of *L. (H.) kankauchare*. As a result, the Japanese specimens of *L. (H.) kankauchare* recorded by Blüthgen (1925) proved to be a misidentification of *L. (H.) japonicum*.

**Comments on non-Japanese species excluded from the *sexstrigatus* group**

*Lasioglossum (Hemihalictus) micante* (Michener, 1993) endemic in Taiwan belongs to the *sexstrigatus* group sensu Sakagami & Ebmer (1996), Pesenko (2007a) and Murao *et al.* (2010) (Michener 1993). According to Michener (1993), a male (holotype) of *L. (H.) micante* (female unknown) lacks both the genal process of the head and the median process of S8. This species probably belongs to a different species group in the *sexstrigatus* group. However, since only a male of this species is known, future taxonomic studies of the genus *Lasioglossum* in Taiwan will be necessary.

*Lasioglossum (Hemihalictus) amamiense* Ebmer & Sakagami, 1994

Fig. 18D


**Diagnosis**

Females are similar to *Lasioglossum (Hemihalictus) taeniolellum* (Vachal, 1903) but differ by the labrum without lateral projection on distal process (Murao *et al.* 2010: fig. 1m) and mesepisternum with shallow PP on upper area (Murao *et al.* 2010: fig. 1f). In contrast, in *L. (H.) taeniolellum*, the distal process of labrum has a horn-like lateral projection (Fig. 14D) and a mesepisternum with deep PP on the upper area.

**Distribution**

Japan (central Ryukyus: Amami-Ohshima Is., Kikai-jima Is., and Tokunoshima Is.).

**Flight records**

Female: March to October.

Male: May to September (Murao *et al.* 2010).

**Flower records**

Thirteen species in 9 families were reported as floral records by Murao *et al.* (2010).

**Habitat**

This species has been collected from cultivated and urban areas at low elevations and seaside wasteland (Murao *et al.* 2010).
Lasioglossum (Hemihalictus) frigidum Sakagami & Ebmer, 1996
Figs 2C, F, 4, 15A, 17A, 19D, 20A

Lasioglossum (Evylaeus) frigidum Sakagami & Ebmer, 1996: 899, figs 1–2, 3a–b, 4a–b, 5a, 6b (♀♂)
(holotype: Systematic Entomology, Faculty of Agriculture, Hokkaido University, Japan, ♂, type locality=Amagamori (in Misawa), Aomori Pref., Honshu, Japan).

Diagnosis
Females are similar to Lasioglossum (Hemihalictus) epicinctus (Strand, 1914) from Taiwan. According to Sakagami & Ebmer (1996), the differences between L. (H.) frigidum and L. (H.) epicinctus are not clearly described. The type specimen of L. (H.) epicinctus at the Senckenberg Deutsches Entomologisches Institut (Müncheberg, Germany) was examined in 2012. Based on this examination, the female of L. (H.) frigidum is separated from L. (H.) epicinctus by the supraclypeal area dimly shiny (IS weakly tessellate), the ridges of metapostnotum long, nearly reaching the posterior margin as in Fig. 4F, and the lineolation of T1 present over the entire surface (Fig. 15A). In contrast, in L. (H.) epicinctus, the supraclypeal area weakly shiny (IS nearly smooth), the ridges of metapostnotum short (only present on basal area), and the lineolation of T1 present on basal and apical areas.

Material examined
Paratypes
JAPAN – Hokkaido • 2 ♀♂; Abashiri-district, Hamakoshimizu; 19 Jun. 1967; H. Fukuda and K. Yamauchi leg.; MNHAH. – Honshu • 1 ♀; Aomori Pref., Misawa, Amagamori; 16 Aug. 1986; M. Yamada leg.; MNHAH • 1 ♀; Ibaraki Pref., Muramatsu; 13 May 1981; M. Takahashi leg.; MNHAH.

Other material
JAPAN – Honshu • 1 ♀; Tottori Pref., Yumiga-hama; 8 Aug. 1993; Y. Maeta leg.; MNHAH • 1 ♂; same location as for preceding; 6 Sep. 1993; Y. Maeta leg.; MNHAH • 1 ♂; Shimane Pref., Izumo-shi, Sotozono-cho; 12 Jul. 1994; Y. Maeta and K. Minagi leg.; SULE • 1 ♂; same location as for preceding; 10 Jun. 1995; Y. Maeta and K. Minagi leg.; SULE. – Kyushu • 2 ♀♂; Fukuoka Pref., Itoshima-shi, Shima-machi, Nigino-hama; 33°35′22.329″ N, 130°8′8.868″ E; 17 Aug. 2013; R. Murao leg.; ELKU • 1 ♂; same location as for preceding; 12 Jul. 2014; R. Murao leg.; ELKU • 1 ♂; same location as for preceding; 6 Jun. 2015; R. Murao leg.; ELKU • 1 ♀; Fukuoka Pref., Fukuoka-shi, Nishi-ku, Imazu-nagahama; 33°36′37.64″ N, 130°15′34.919″ E; 2 Jul. 2006; R. Murao leg.; ELKU • 1 ♀; same collection data as for preceding; 28 Jun. 2009; R. Murao leg.; ELKU • 1 ♂; same collection data as for preceding; ELKU • 1 ♂; same collection data as for preceding; 2 Aug. 2009; R. Murao leg.; ELKU • 1 ♂; same collection data as for preceding; ELKU • 1 ♀; same location as for preceding; 16 Aug. 2011; R. Murao leg.; ELKU • 3 ♀♂; same location as for preceding; 29 Jun. 2013; R. Murao leg.; ELKU • 6 ♂; same collection data as for preceding; ELKU • 1 ♂; same location as for preceding; 29 Jun. 2013; Y. Murao leg.; ELKU • 4 ♂♂; same collection data as for preceding; ELKU • 1 ♂; same location as for preceding; 17 Aug. 2013; R. Murao leg.; ELKU • 1 ♀; same collection data as for preceding; ELKU • 1 ♀; same location as for preceding; 17 May 2014; R. Murao leg.; ELKU • 2 ♂♂; Fukuoka Pref., Fukuoka-shi, Nishi-ku, Ikino-matsubara; 33°34′51.671″ N, 130°18′1.335″ E; 8 Jul. 2013; R. Murao leg.; ELKU • 1 ♀; Fukuoka Pref., Fukuoka-shi, Higashi-ku, Shikano-shima, Gebaga-hama; 33°40′57.566″ N, 130°17′18.933″ E; 20 Sep. 2012; R. Murao leg.; ELKU • 1 ♀; same location as for preceding; 15 Aug. 2013; R. Murao leg.; ELKU • 1 ♀; Fukuoka Pref., Kasuya-gun, Shingu-machi, Shingu-hama; 33°43′8.861″ N, 130°26′15.07″ E; 29 Jul. 2006; R. Murao leg.; ELKU • 1 ♂; same location as for preceding; 14 Oct. 2007; R. Murao leg.; ELKU • 1 ♂; same collection data as for preceding; 10 May 2009; R. Murao leg.; ELKU •
39 ♀♀; same location as for preceding; 3 Jun. 2011; R. Murao leg.; ELKU • 1 ♂; same location as for preceding; 16 Jul. 2011; R. Murao leg.; ELKU • 1 ♀; same location as for preceding; 20 Sep. 2012; R. Murao leg.; ELKU • 1 ♀; same location as for preceding; 4 May 2014; R. Murao leg.; ELKU • 3 ♀♂; Fukuoka Pref., Fukutsu-shi, Koinoura-kaigan; 33°48′00.327″ N, 130°27′02.674″ E; 2 May 2015; R. Murao leg.; ELKU • 5 ♀♀; Kumamoto Pref., Amakusa, Tomioka-shiki; 19 Jun. 1931; Esaki and Hori

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**Fig. 4. Lasioglossum (Hemihalictus) frigidum** Sakagami & Ebmer, 1996. A. ♀, lateral habitus. B. ♂, head in frontal view. C. ♂, head in frontal view. D. ♀, labrum. E. ♀, mesoscutum. F. ♀, metapostnotum. Scale bars: A = 2 mm; B, E–F = 0.5 mm; C–D = 0.2 mm.
Distribution

Japan (Hokkaido, Honshu, Izu-shotô Islands, Shikoku, Kyushu, northern Ryukyus).

Flight records

Female: April to October.

Male: June to October. The flight records of male are based on the phenological data reported by Minagi et al. (2000).

Flower records


Habitat

Lasioglossum frigidum has been collected only in coastal sand dunes. One of the collecting sites is shown in Fig. 19D.

Lasioglossum (Hemihalictus) ikudomei sp. nov.

Figs 5, 15B, 18D, 20B

Diagnosis

Females are similar to Lasioglossum (Hemihalictus) smilodon Ebmer & Sakagami, 1994 from Japan but are separated from them by the supraclypeal area shinier (IS weakly tessellate), the PP on the mesoscutum sparser (IS = 3 d in maximum) (Fig. 5E), and T1 with very weak lineolation. In contrast, in L. (H.) smilodon, the supraclypeal area is dimly shiny (IS distinctly tessellate), the PP on the mesoscutum denser (IS = 2 d at maximum) (Fig. 9E), and the lineolation of T1 clearer.

Etymology

The specific name is dedicated to Dr Shuichi Ikudome (KWC), who contributed greatly to clarify the bee fauna of Ryukyus Islands, southwestern Japan.

Material examined

Holotype

Paratypes
JAPAN – Ryukyus • 1 ♀; Yoron Is., Maeno-hama; 4 Jun. 1989; KWC • 1 ♀; Okinawa Pref., Kunigami, Hedo; 5 Apr. 1979; K. Kusigemati leg.; ELKU • 2 ♀; Okinawa Pref., Hedo; 5 Apr. 1979; K. Ohara leg.; ELKU • 2 ♀; Okinawa Pref., Kunigami-gun, Kunigami-son, Cape Hedo; 26°52′ N, 128°15′ E; 2 Apr. 2017; K. Otsui leg.; cMur • 1 ♀; Okinawa Pref., Okinawa-jima, Kunigami-gun, Kunigami-son, Ada;

Fig. 5. Lasioglossum (Hemihalictus) ikudomei sp. nov. A. ♀, lateral habitus. B. ♀, head in frontal view. C. ♂, head in frontal view. D. ♀, labrum. E. ♀, mesoscutum. F. ♀, metapostnotum. Scale bars: A = 2 mm; B–C, E = 1 mm; D = 0.2 mm; F = 0.5 mm.
Description

Female

Measurements (n=5, unit mm). BL = 5.13–5.63 (5.43 ± 0.23), WL = 4.25–4.88 (4.65 ± 0.26), HL = 1.55–1.65 (1.61 ± 0.04), HW = 1.48–1.65 (1.57 ± 0.06), IOD = 0.26–0.29 (0.28 ± 0.02), OOD = 0.24–0.31 (0.26 ± 0.02), OCD = 0.18–0.19 (0.19 ± 0.01, n = 4), UOD = 0.94–1.03 (0.98 ± 0.04), MOD = 1.10–1.23 (1.15 ± 0.05), LOD = 0.84–0.94 (0.88 ± 0.04), IAD = 0.15–0.16 (0.16 ± 0.01), AOD = 0.26–0.31 (0.28 ± 0.02), CAL = 0.29–0.32 (0.31 ± 0.01), CPL = 0.34–0.37 (0.35 ± 0.01), EL = 1.75–1.85 (1.80 ± 0.04), EW = 0.39–0.45 (0.41 ± 0.04), GW = 0.29–0.39 (0.35 ± 0.04), SPL = 0.61–0.66 (0.64 ± 0.02), F1L = 0.10 (0.10 ± 0.00), F2L = 0.08 (0.08 ± 0.00), F3L = 0.08 (0.08 ± 0.00), F2W = 0.13–0.15 (0.14 ± 0.01), MsW = 1.80–2.05 (1.93 ± 0.09), SCL = 0.38–0.45 (0.43 ± 0.03), MNL = 0.25–0.30 (0.27 ± 0.02), MPL = 0.25–0.33 (0.28 ± 0.03), MtW = 1.75–2.10 (1.97 ± 0.13).

Coloration. Body black except for the following parts: mandible reddish brown apically; flagellum brown or blackish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga narrowly yellowish brown transluent apically. Wings transparent, veins and stigma brown.

Pubescence. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotum moderately densely tomentose dorsally and around lobe; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa. Disc of T1 without distinct short hairs on medial area.

Structure and sculpture head. Slightly longer than wide or nearly as long as wide; HW:HL = 1:1.03. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.85:0.76. IOD:OOD:OCD = 1:0.95:0.69. IAD:AOD = 1:1.86. Ocellocular area with moderately dense PP, IS smooth (IS = 0.5–3 d). Paraocular area and frons weakly shiny, with shallow reticulate PP. Supracypeal area slightly convex, weakly shiny, with sparse PP, IS weakly tessellate (IS = 1–4 d). Clypeus nearly flat, with sparse PP over entire surface, IS very weakly tessellate on upper half or ⅗ and smooth on lower half or ⅔ (IS = 1–5 d). EW:GW = 1:0.86. Genal area to postgena with distinct straight ridges. Malar space linear. Occiput not carinate. Hypostomal carinae nearly parallel. Mandible bidentate. Labrum (Fig. 5D): basal area approximately 1.9 × as wide as long; distal process approximately 0.6 × as long as basal area, tongue-like, and without lateral projection; distal keel rounded, pointed apically. Antenna short, not reaching metasoma. F2L:F2W = 1:1.72; flagellum nearly flattened ventrally.

Thorax. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 5E) with moderately dense PP over entire surface; IS weakly tessellate over entire surface (particularly very weak on posterior area) (IS = 0.5–3 d); parapsidal line a narrow groove. Mesoscutellum with 2–4 PP on submedian area and denser PP on marginal area; IS nearly smooth on submedian area and very weakly tessellate on marginal area (IS = 2–5 d on submedian area, = 0.5–2 d on marginal area). Metanotum weakly rugulose. Mesepisternum weakly shiny, with dense shallow PP on upper area and reticulate PP on lower area; IS nearly smooth on upper area (IS = 0.5–1 d on upper area). SCL:MNL:MPL = 1:0.62:0.66. Propodeum: metapostnotum (Fig. 5F) gently inclined, with irregular sinuate ridges on anterior half in holotype and two paratypes (remaining five paratypes with short longitudinal ridges), with weak tessellation on posterior half, and nearly smooth among ridges; junction between metapostnotum and posterior surface not carinate, with weak tessellation; lateral surface weakly rugulose; posterior surface with lateral carina on lower half, without oblique carina. Coxae usual shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial
plate of hind leg carinate marginally. Inner hind tibial spur with slender 2–4 teeth as in Fig. 20B (n = 7). Fore wing with three submarginal cells.

**Abdomen.** Disc of T1 without distinct PP on medial area, and with very weak lineolation interrupted on medial area (Fig. 15B). Disc of T2 nearly smooth on anterior to medial area in holotype and two paratypes, and with weak lineolation on posterior area (anterior and posterior area with weak lineolation in four paratypes). Discs of T3–T4 with weak lineolation over entire surface.

**Male**

**Measurements** (n = 1, unit mm). BL = 4.23, WL = 3.92, HL = 1.56, HW = 1.51, IOD = 0.29, OOD = 0.29, OCD = 0.16, UOD = 0.98, MOD = 1.04, LOD = 0.84, IAD = 0.20, AOD = 0.24, CAL = 0.31, CPL = 0.33, EL = 1.07, EW = 0.40, GW = 0.44, SPL = 0.44, F1L = 0.11, F2L = 0.18, F3L = 0.16, F2W = 0.13, MsW = 1.55, SCL = 0.36, MNL = 0.22, MPL = 0.27, MtW = 1.35.

**Coloration.** Body black except for the following parts: mandible yellowish brown except for apically reddish brown; labrum dark yellow; pronotal lobe yellowish brown; tegula yellowish brown translucent; legs brown, without distinct yellow marks; tibial spur yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma pale brown.

**Pubescence.** Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: lower paraocular area and pronotal dorsum to lobe sparsely tomentose. Disc of T1 with sparse hairs only on marginal area. Disc of T2–T4 with sparse short hairs over entire surface. T2–T3 with thin apical fimbriae, less distinct than in the female.


**Thorax.** Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum with moderately dense PP over entire surface; IS weakly tessellate on anterior margin, nearly smooth on rest parts (IS = 1.5–3 d); parapsidal line a narrow groove. Mesoscutellum with sparse PP over entire surface; IS smooth (IS = 2.5–6 d). Metanotum weakly rugulose. Mesepisternum with moderately dense PP over entire surface; IS smooth (IS = 1–3 d). SCL:MNL:MPL = 1:0.63:0.75. Propodeum: metapostnotum weakly shiny and gently inclined, with short straight ridges occupying anterior ⅔; junction between metapostnotum and posterior surface not carinate, nearly smooth; lateral surface weakly reticulate; posterior surface nearly smooth, with lateral carina on lower ⅓, and without oblique carina. Coxae usual shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur serrate. Fore wing with three submarginal cells.

**Abdomen.** Disc of T1 nearly smooth. Disc of T2–T3 with sparse fine PP; T2 weakly lineolate on apical margin; T3 weakly lioleolate on apical half. Disc of T4 weakly lineolate over entire surface. S7 with moderately long, apically rounded median process.
GENITALIA. Gonobase flat at bottom; gonocoxite smooth; ventral retrorse lobe tongue-like, moderately long but not reaching gonobase, with sparse short hairs ventrally.

Distribution
Japan (central to southern Ryukyus: Yoron-jima Is., Okinawa-jima Is., Miyako-jima Is., Iriomote-jima Is.).

Flight period
Female: middle March to late August.
Male: July.

Flower records
Ixeris japonica (Burm.f.) Nakai (Asteraceae).

_Lasioglossum (Hemihalictus) kiautschouense_ (Strand, 1910)
Figs 6, 15C, 17B, 19A, 20C

_Halictus kiautschouense_ Strand, 1910: 195 (holotype: ZMHB, ♀, type locality = Kiautshou, Tsingtou, China, examined).
_Halictus perplexans_ Cockerell, 1925: 10 (syntype: ♀, ZMHB, type locality = Preobrageny Bay, Russia, examined; 4 ♀♀, USNM, type locality = Preobrageny Bay and Low Lighthouse, Russia). _Syn. nov._

_Evylaeus (Prospalictus) kiautschouensis_ – Pesenko 2007b: 111.
_Evylaeus (Prospalictus) perplexans_ – Pesenko 2007b: 111.

Diagnosis
Females are separated from the other members of the _sexstrigatus_ group occurring in Japan by a combination of the following character states: IS of mesoscutum nearly smooth on posterior area; metasoma entirely black; metasomal terga with white fimbriae on lateral-apical margin; and disc of T1 with distinct sparse fine PP (Fig. 15C) and without lineolation (Murao 2017a).

Material examined
Holotype
CHINA • ♀; Tsingtou, Kiautshou; Jun.–Jul. 1903; S. Glaue leg.; ZMHB.

Other material
JAPAN – Hokkaido • 1 ♀; Ashoro-cho, Ashoro; 43°20′0″ N, 143°40′0″ E; 2 Jul. 2013; O. Tadauchi leg.; ELKU • 1 ♀; Asahikawa, Asahiyama; 24 Sep. 1970; MNHAH • 2 ♀♀; Ebetsubuto; 8–9 Jun. 1974; M. Ishikawa leg.; MNHAH • 3 ♀♀; Asahikawa, Inosawa; 27 Jun. 1969; MNHAH • 10 ♀♀; Kiritapp; 1972; MNHAH • 1 ♀; Moiwa; 3 Jun. 1972; Kawano leg.; MNHAH • 6 ♀♀; Kushiro, Tenneru n.; 1968; E. Ohtsuka leg.; MNHAH • 5 ♀♀; Tobetsu; 22 May–9 Jul. 1974; M. Ishikawa leg.; MNHAH • 6 ♀♀; Lake Shikotsu (Iburi), Morappu; 11–13 Aug. 1953; Y. Hirashima leg.; ELKU • 2 ♀♀; Mombetsu-gun, Engaru; 11 Aug. 1955; K. Morimoto leg.; ELKU. – Honshu • 2 ♀♀; Aomori Pref., Mt Iwaki; 23 Jun.
1981; M. Yamada leg.; MNHAH • 1 ♀; Iwate Pref., Morioka, Kuriyagawa; 16 May 1964; Y. Maeta leg.; ELKU • 1 ♀; Yamagata Pref., Murayama, Tochiuda; 26 May 1975; O. Tadauchi leg.; ELKU • 1 ♀; Yamaguchi Pref., Hagi, Sengokudai; 23 May 1960; Y. Hirashima leg.; ELKU. – Izu Islands • 28 ♀;&

♀; Hachijo Is., Okago-Sokoto; 5 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU • 3 ♀; Hachijo Is., Mitsune-Kantoyama; 30 May 1964; Y. Hirashima and M. Shiga leg.; ELKU • 2 ♀; Okago-Fuji; 26 May 1964; Y. Hirashima and M. Shiga leg.; ELKU • 5 ♀; Hachijo Is., Sokoto; 4 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU • 2 ♀; Okago-Fuji; 26 May 1964; Y. Hirashima and M. Shiga leg.; ELKU • 5 ♀; Hachijo Is., Nakanogo-Daigo yama-Mitsune; 1 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU. – Kyushu • 1 ♀; Hiraodai (Buzen); 6 Jul. 1952; K. Yasumatsu leg.; ELKU • 2 ♀; Mt Oita Pref., Kokonoe-machi, Sensui-san; 23 May 2005; K. Mitai leg.; cMur • 1 ♀; Oita Pref., Kusu-gun, Kokonoe-machi, Chojyabaru; 33°7′6.773″ N, 131°13′49.331″ E; 1050 m a.s.l.; 14 Aug. 2010; R. Murao leg.; cMur • 1 ♀; same location as for preceding; 5 Sep. 2010; R. Murao leg.; cMur • 1 ♀; Kumamoto Pref., Aso-shi; 33°0′9.146″ N, 131°8′11.141″ E; 21 Jul. 2013; R. Murao leg.; ELKU.

Redescription

MEASUREMENTS (n = 5, unit mm). BL = 4.50–4.88 (4.68 ± 0.17), WL = 4.00–4.63 (4.38 ± 0.27), HL = 1.42–1.55 (1.47 ± 0.06), HW = 1.45–1.58 (1.51 ± 0.07), IOD = 0.26–0.29 (0.27 ± 0.02), OOD = 0.29–0.31 (0.29 ± 0.01), OCD = 0.18–0.19 (0.18 ± 0.01), UOD = 0.97–1.03 (0.99 ± 0.03), LOD = 1.10–1.16 (1.12 ± 0.04), IAD = 0.18–0.19 (0.18 ± 0.01), AOD = 0.97–1.03 (0.99 ± 0.03), MOD = 0.26–0.29 (0.27 ± 0.02), OOD = 0.29–0.31 (0.29 ± 0.01), CAL = 0.24–0.29 (0.27 ± 0.02), CPL = 0.97–1.03 (0.99 ± 0.03), F1L = 0.08–0.10 (0.09 ± 0.01), F2L = 0.08 (0.08 ± 0.00), F3L = 0.08 (0.08 ± 0.00), F2W = 1.60–1.75 (1.67 ± 0.07), EW = 0.35–0.42 (0.39 ± 0.03), GW = 0.29–0.32 (0.30 ± 0.01), SPL = 0.58–0.68 (0.62 ± 0.04), F1L = 0.08–0.10 (0.09 ± 0.01), F2L = 0.08 (0.08 ± 0.00), F3L = 0.08 (0.08 ± 0.00), F2W = 0.11–0.15 (0.14 ± 0.01), MsW = 1.60–1.80 (1.69 ± 0.10), SCL = 0.33–0.40 (0.36 ± 0.03), MNL = 0.20–0.23 (0.21 ± 0.01), MPL = 0.23–0.25 (0.25 ± 0.01), MtW = 1.60–1.90 (1.78 ± 0.13).

Female

COLORATION. Body black except for the following parts: mandible reddish brown apically; F4–F10 or F5–F10 yellowish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga narrowly yellowish brown translucent apically. Wings transparent, veins and stigma yellowish brown.

PUBESCENCE. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotum moderately densely tomentose on dorsal area and around lobe; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa.

STRUCTURE AND SCULPTURE HEAD. Nearly as long as wide; HW:HL = 1:0.97. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.88:0.73. IOD:OCD:OD = 1:1.07:0.67. IAD:AOD = 1:1.96. Ocellocular area densely punctate, IS smooth (IS = 0.5–1.5 d). Paraocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area nearly flat, weakly shiny, with moderately dense PP, IS weakly tessellate (IS = 0.5–1 d). CPL:CAL = 1:0.87. Clypeus nearly flat, with dense PP on upper half and larger shallow PP on lower half, IS nearly smooth (IS = 0.5 d on upper half), EW = 1.07. Genal area with weak straight ridges. Malar space linear. Occiput not carinate. Postgena distinctly tessellate. Hypostomal carinae nearly parallel. Mandible bidentate. Labrum (Fig. 6C): basal area approximately 2× as wide as long; distal process approximately 0.7× as long as basal area, narrow, and without lateral projection; distal keel pointed apically. Antenna short, not reaching metasoma. F2L:F2W = 1:1.68; flagellum nearly flattened ventrally.

THOARX. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 6D) with dense PP over entire surface; IS nearly smooth on posterior area, and distinctly tessellate on the rest area (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with dense PP over entire surface, IS smooth (IS = 0.5–2 d). Metanotum weakly rugulose.
Mesepisternum weakly shiny, with reticulate PP over entire surface. SCL:MNL:MPL = 1:0.57:0.68. Propodeum: metapostnotum (Fig. 6E) gently inclined, with straight ridges nearly attaining to posterior margin, and nearly smooth among ridges; the junction between metapostnotum and posterior surface not carinate; lateral surface weakly rugulose; posterior surface with lateral carina on lower half, without oblique carina. Coxae normal shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial

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**Fig. 6.** *Lasioglossum (Hemihalictus) kiautschouense* (Strand, 1910), ♀.  
A. Lateral habitus.  
B. Head in frontal view.  
C. Labrum.  
D. Mesoscutum.  
E. Metapostnotum.  
Scale bars: A = 3 mm; B, D–E = 0.5 mm; C = 0.25 mm.
plate of hind leg carinate marginally. Inner hind tibial spur with slender 2–4 teeth as in Fig. 20C (n = 17). Fore wing with three submarginal cells.

**ABDOMEN.** Disc of T1 with sparse distinct fine PP on medial area and without lineolation over entire surface (Fig. 15C). Discs of T2–T4 without lineolation over entire surface (sometimes T4 with very weak lineolation).

**Male**
Not examined in the present study.

**Distribution**
Japan (Hokkaido, Honshu, Kyushu, Izu-shotô Islands), Korean Peninsula, Russian Far East, China.

**Flight records**
Female: May to September.

Males have been collected from July to August in Primorsky, Russian Far East (Ebmer 1996, 2006).

**Flower records**
The specimens examined in this paper were collected on the flowers of 7 species in 3 families as follows.

**Habitat**
This species has been collected from semi-natural grassland in western Japan. One of the collecting sites is shown in Fig. 19A.

**Remarks**
According to Cockerell (1925), the syntypes of *Halictus perplexans* are preserved in USNM. During my visit to ZMHB in 2012, I found and examined a syntype of *H. perplexans*.

**Lasioglossum (Hemihalictus) ohei** Hirashima & Sakagami, 1966
Figs 7, 15D, 17C, 19A, 20D


**Diagnosis**
Females are similar to *L. (H.) simplicior* but separated from them by the head slightly longer than wide or nearly as long as wide (HL/HW ratio 1.01 ± 0.02), IS of mesoscutum nearly smooth on posterior margin, and metasomal terga with silky dull luster. In contrast, in *L. (H.) simplicior*, the head wider than long (HL/HW ratio 0.97±0.03), IS of mesoscutum with distinct tessellation over entire surface, and metasomal terga with enamel-like luster as in most species of *Lasioglossum*.

**Material examined**

**Paratypes**
JAPAN – Hokkaido • 1 ♀; Kitami; 18 Jul. 1965; S.F. Sakagami leg.; MNHAH • 16 ♀♀, 7 ♂♂; same location as for preceding; 22 Jul.–14 Aug. 1964; S.F. Sakagami leg.; MNHAH • 3 ♀♀; same location as
for preceding; 14 Aug. 1964; S.F. Sakagami leg.; MNNAH • 6 ♀♀, 2 ♂♂; same location as for preceding; 23 Aug. 1959; S.F. Sakagami leg.; MNNAH • 1 ♂; Kitamoshiri, Hokkaido Uryu Experimental Forest; 12 Sep. 1969; S.F. Sakagami and H. Fukuda leg.; MNNAH • 1 ♂, 4 ♂♂; same location as for preceding; 14 Sep. 1969; S.F. Sakagami and H. Fukuda leg.; MNNAH.

Other material
JAPAN – Hokkaido • 1 ♀; Mt Daisetsu, Aizankei; 30 Jul.–3 Aug. 1955; Y. Hirashima leg.; ELKU • 1 ♀; Asahikawa, Asahiyama; 18 Jun. 1985; T. Inaoka leg.; MNNAH • 1 ♀; Sapporo, Barato; 3 Jun. 1973; M. Matsumoto leg.; MNNAH • 1 ♀; same location as for preceding; 7 Jun. 1973; M. Matsumoto leg.; MNNAH • 1 ♀; same location as for preceding; 14 Jun. 1973; M. Matsumoto leg.; MNNAH • 7 ♀♀; same location as for preceding; 21 Jun. 1973; M. Matsumoto leg.; MNNAH • 12 ♀♀; same location as for preceding; 28 Jun. 1973; M. Matsumoto leg.; MNNAH • 1 ♀; same location as for preceding; 27 Jul. 1973; M. Matsumoto leg.; MNNAH • 2 ♀♀; same location as for preceding; 3 Aug. 1973; M. Matsumoto leg.; MNNAH • 1 ♀; same location as for preceding; 8 Aug. 1973; M. Matsumoto leg.; MNNAH • 1 ♀; Fukushima; 27 Jul. 1965; M. Munakata leg.; MNNAH • 1 ♀; Asahikawa, Inosawa; 30 May 1969; MNNAH • 4 ♀♀; same location as for preceding; 25 Jun. 1969; MNNAH • 6 ♀♀; same location as for preceding; 27 Jun. 1969; MNNAH • 14 ♀♀; same location as for preceding; 13 Jul. 1969; MNNAH • 1 ♀; same location as for preceding; 15 Jul. 1969; MNNAH • 1 ♀; same location as for preceding; 15 Aug. 1969; MNNAH • 2 ♀♀; same location as for preceding; 29 May 1970; MNNAH • 14 ♀♀; same location as for preceding; 10 Jun. 1970; MNNAH • 7 ♀♀; Nokanan; 8 Jun. 1967; MNNAH • 6 ♀♀; same location as for preceding; 22 Jun. 1967; MNNAH • 1 ♀; same location as for preceding; 27 Jun. 1967; MNNAH • 1 ♀; same location as for preceding; 6 Jul. 1967; MNNAH • 17 ♀♀; same location as for preceding; 21 Jul. 1967; MNNAH • 3 ♀♀; Sapporo, Hokkaido University Campus; 6 Jun. 1959; S.F. Sakagami leg.; MNNAH • 1 ♀; same location as for preceding; 11 Jun. 1959; S.F. Sakagami leg.; MNNAH • 1 ♀; same location as for preceding; 14 Jun. 1959; S.F. Sakagami leg.; MNNAH • 2 ♀♀; same location as for preceding; 18 Jun. 1959; S.F. Sakagami leg.; MNNAH • 4 ♀♀; same location as for preceding; 30 Jun. 1959; S.F. Sakagami leg.; MNNAH • 1 ♀; Yukomanbetsu; 22 Jul. 1967; MNNAH • 1 ♀; same location as for preceding; 4 Jul. 1968; MNNAH • 1 ♀; Ebetsubuto; 18 Jul. 1974; M. Ishikawa leg.; MNNAH. – Honshu • 1 ♀; Aomori Pref., Mt Iwaki; 5 Oct. 1980; M. Yamada leg.; MNHAN • 1 ♀; same location as for preceding; 14 Jul. 1981; M. Yamada leg.; MNHAN • 1 ♀; Miyagi Pref., Shiogama-shi, Hojima; 19 May 1995; K. Gōukon leg.; cGou • 1 ♀; same location as for preceding; 25 May 1996; K. Gōukon leg.; cGou • 1 ♀; Niigata Pref., Sado Is., Dondonyama; 9 Jul. 1999; K. Gōukon leg.; cGou • 1 ♀; same location as for preceding; 10 Jul. 1999; K. Gōukon leg.; cGou • 1 ♀; Saitama Pref., Kodama; 6 Jun. 1968; T. Nambu leg.; ELKU • 4 ♀♀; same location as for preceding; 3 Jul. 1968; T. Nambu leg.; ELKU • 1 ♀; Nagano Pref., Yamaguchi-mura, Magome; 15 May 1975; O. Tadauchi leg.; ELKU • 5 ♀♀; Gifu Pref., Hikie; 5 Jun. 1978; K. Yamauchi leg.; MNNAH • 3 ♀♀; same location as for preceding; 28 Jun. 1978; K. Yamauchi leg.; MNNAH • 1 ♀; Wakayama Pref., Kibi; 2 Apr. 1969; M. Matsuura leg.; MNNAH • 1 ♀; same location as for preceding; 26 May 1969; M. Matsuura leg.; MNNAH • 1 ♀; Yamaguchi Pref., Toyoda-machi, Houra; 12 May 2004; T. Sugimoto leg.; cMur. – Kyushu • 5 ♀♀; Fukuoka Pref., Mt Wakasugi-yama; 22 Apr. 1973; O. Tadauchi leg.; ELKU • 1 ♀; Oita Pref., Kokonoe-machi, Jizoyabaru; 1 Nov. 1970; K. Kaniyama leg.; ELKU • 2 ♀♀; Oita Pref., Kusu-gun, Kokonoe-machi, Handakougen; 5 Jun. 2004; T. Sugimoto leg.; cMur • 1 ♀; Oita Pref., Kusu-gun, Kokonoe-machi, Chojyabaru; 33°7′6.773″ N, 131°3′49.331″ E; 1050 m a.s.l.; 13 Aug. 2010; R. Murao leg.; cMur • 3 ♀♀; same location as for preceding; 14 Aug. 2010; R. and Y. Murao leg.; cMur • 1 ♀; same location as for preceding; 5 Sep. 2010; Y. Murao leg.; cMur • 8 ♀♀; same location as for preceding; 15 May 2011; R. and Y. Murao leg.; cMur • 3 ♀♀; same location as for preceding; 12 Aug. 2011; R. Murao leg.; cMur • 1 ♀; same location as for preceding; 28 Aug. 2011; R. Murao leg.; cMur • 1 ♀; same location as for preceding; 5 Aug. 2013; R. Murao leg.; ELKU • 2 ♀♀; Kumamoto Pref., Aso-gun, Aso-machi, Matoishi-Kario; 28 Sep. 2007; R. Murao and Y. Nishimura leg.; cMur • 2 ♀♀, 3 ♂♂; Kumamoto Pref.,

**Distribution**

Japan (Hokkaido, Honshu, Kyushu, northern Ryukyus).

![Fig. 7. Lasioglossum (Hemihalictus) ohei Hirashima & Sakagami, 1966. A. ♀, lateral habitus. B. ♀, head in frontal view. C. ♂, head in frontal view. D. ♀, labrum. E. ♀, mesoscutum. F. ♀, metapostnotum. Scale bars: A = 2 mm; B = 1 mm; C, E–F = 0.5 mm; D = 0.2 mm.](image-url)
Flight period
Female: April to November.

Male: July to September. The flight records of males are based on the paratypes collected or reared from the nest (Sakagami et al. 1966).

Flower records
The specimens examined in this paper were collected on the flowers of 50 species in 19 families as follows.

- Amaryllidaceae: Allium fistulosum L.
- Apiaceae: Sanicula chinensis Bunge.
- Asteraceae: Achillea alpina L. var. longiligulata H.Hara; Aster microcephalus (Miq.) Franch. & Sav. var. ovatus (Franch. & Sav.) Soejima & Mot.Ito; Eupatorium glehni F.Schmidt ex Trautv.; Ixeridium dentatum (Thunb.) Tzvelev.
- Brassicaceae: Brassica rapa L. var. glabra Regel ‘Pe-tsai’; B. rapa L. var. oleifera DC.; B. rapa L. var. rapa; Capsella bursa-pastoris (L.) Medik.; Raphanus sativus L. var. hortensis Backer; Thlaspi arvense L.
- Campanulaceae: Lobelia sessilifolia Lamb.
- Commelinaceae: Commelina communis L.
- Convolvulaceae: Calystegia pubescens Lindl.
- Hydrangeaceae: Deutzia scabra Thunb.; Hydrangea macrophylla (Thunb.) Ser. f. macrophylla; H. serrata (Thunb.) Ser. var. serrata.
- Hypericaceae: Hypericum patulum Thunb.
- Iridaceae: Iris sanguinea Hornem.
- Lamiaceae: Prunella vulgaris L. subsp. asiatica (Nakai) H.Hara; Hylomecon japonica (Thunb.) Prantl & Kündig.
- Papaveraceae: Chelidonium majus L. subsp. asiaticum H.Hara; Paeonia suffruticosa Andrews.
- Polygonaceae: Persicaria filiformis (Thunb.) Nakai ex W.T.Lee; Persicaria longiseta (Brujin) Kitag.
- Saururaceae: Houttuynia cordata Thunb.

Habitat
This species has been collected mainly from the mountain of western Japan. One of the collecting sites is shown in Fig. 19A.

Biological reference
Sakagami et al. (1966) and Sakagami (1992) reported on the biology of this species as univoltine and communal, with nest structure type Ia of Sakagami & Michener (1962).

_Lasioglossum (Hemihalictus) simplicior_ (Cockerell, 1931)
Figs 2D, 8, 15E, 17D

_Halictus simplicior_ Cockerell, 1931: 16 (holotype: AMNH, ♀, type locality = Shanghai, China, examined).

_Evylaeus (Prosopalictus) simplicior_ – Pesenko 2007b: 111.

Diagnosis
Females are similar to _L. (H.) ohei_, but separated from them by the head wider than long (HL/HW ratio 0.97 ± 0.03), IS of mesoscutum with distinct tessellation over entire surface, and metasomal terga with enamel-like luster. In contrast, in _L. (H.) ohei_, the head is slightly longer than wide or nearly as long as wide (HL/HW ratio 1.01 ± 0.02), IS of mesoscutum nearly smooth on posterior margin, and metasomal terga with silky dull luster.
Material examined

Holotype
CHINA • ♀; Prov. Kiangsu, Shanghai, Zô-Sè; AMNH.

Other material
JAPAN – Izu Islands • 44 ♀♂; Hachijo Is., Okago-Fuji; 26 May 1964; Y. Hirashima and M. Shiga leg.; ELKU • 4 ♂♂; same collection data as for preceding; ELKU • 3 ♀♀♀; Hachijo Is., Okago-Sokoto; 5 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU • 1 ♂; same collection data as for preceding; ELKU • 2 ♀♀♀; Mt Kanto-yama, Hachijo Is.; 2 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU • 1 ♂; same collection data as for preceding; ELKU • 1 ♀; Hachijo Is., Kamogawa; 27 May 1964; Y. Hirashima and M. Shiga leg.; ELKU • 2 ♂♂; same collection data as for preceding; ELKU • 3 ♀♀♀; Hachijo Is., Okago-Sokoto; 5 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU • 1 ♀; same location as for preceding; ELKU • 1 ♀; same location as for preceding; 15 Aug. 1987; H. Takahashi leg.; ELKU • 1 ♂; same location as for preceding; 26 Sep. 1987; H. Takahashi leg.; MNHAH • 1 ♀; same location as for preceding; 4 Oct. 1987; H. Takahashi leg.; MNHAH • 1 ♂; Hachijo Is., Mt Mitsune-yama, Nakanogo-Daigo; 1 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU • 1 ♀; Hachijo Is., Bouei Road; 27 Sep. 1987; H. Takahashi leg.; MNHAH • 2 ♂♂; same collection data as for preceding; MNHAH • 3 ♂♂; same location as for preceding; 3 Oct. 1987; H. Takahashi leg.; ELKU • 1 ♀; Hachijo Is., Okago; 10 Jul. 1987; H. Takahashi leg.; MNHAH • 1 ♂; Hachijo Is., Sokoto; 4 Jun. 1964; Y. Hirashima and M. Shiga leg.; ELKU • 1 ♀; Tokyo, Aoga Is., Yasundogou; 10 Aug. 1987; H. Takahashi leg.; MNHAH.

Redescription

Female

MEASUREMENTS (n = 6, unit mm). BL = 5.25–5.75 (5.41 ± 0.24), WL = 3.85–5.25 (4.81 ± 0.52), HL = 1.48–1.58 (1.54 ± 0.05), HW = 1.52–1.68 (1.59 ± 0.07), IOD = 0.22–0.31 (0.27 ± 0.03), OOD = 0.26–0.33 (0.28 ± 0.03), OCD = 0.13–0.21 (0.18 ± 0.03), UOD = 0.97–1.03 (1.01 ± 0.02), MOD = 1.09–1.23 (1.16 ± 0.06), LOD = 0.82–0.94 (0.88 ± 0.05), IAD = 0.16 (0.16 ± 0.00), AOD = 0.27–0.32 (0.29 ± 0.02), CAL = 0.22–0.31 (0.28 ± 0.03), CPL = 0.32–0.35 (0.34 ± 0.01), EL = 1.09–1.85 (1.66 ± 0.29), EW = 0.39–0.45 (0.42 ± 0.02), GW = 0.29–0.35 (0.33 ± 0.02), SPL = 0.60–0.66 (0.64 ± 0.02), F1L = 0.07–0.10 (0.09 ± 0.01), F2L = 0.07–0.08 (0.08 ± 0.01), F3L = 0.08–0.10 (0.09 ± 0.01), F2W = 0.11–0.15 (0.13 ± 0.01), MsW = 1.71–2.00 (1.88 ± 0.12), SCL = 0.31–0.43 (0.39 ± 0.05), MNL = 0.22–0.28 (0.25 ± 0.02), MPL = 0.24–0.28 (0.26 ± 0.01), MtW = 1.75–2.05 (1.90 ± 0.11).

COLORATION. Body black except for the following parts: mandible reddish brown apically; flagellum blackish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma yellowish brown.

PUBESCENCE. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotum moderately densely tomentose on dorsal area and around lobe; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa.

STRUCTURE AND SCULPTURE HEAD. Wider than long or nearly as long as wide; HW:HL = 1:0.97. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.87:0.76. IOD:OOD:OCD = 1:1.05:0.68. IAD:AOD = 1:1.84. Ocellocular and paraocular areas, frons weakly shiny, with shallow reticulate PP. Supracypeal area slightly convex, dull, with dense PP, IS distinctly tessellate (IS = 0.5–2 d). CPL:CAL = 1:0.82. Clypeus nearly flat, with dense PP on upper half and larger shallow PP on lower half, IS nearly smooth (IS = 0.5–1 d on upper half). EW:GW = 1:0.78. Genal area with weak straight ridges. Malar space linear. Occiput not carinate. Postgena distinctly tessellate. Hypostomal carinae nearly parallel. Mandible bidentate.
Labrum (Fig. 8D): basal area approximately 2.2 × as wide as long; distal process approximately 0.7 × as long as basal area, narrow, and without lateral projection; distal keel pointed apically. Antenna short, not reaching metasoma. F2L:F2W = 1:1.71; flagellum nearly flattened ventrally.

THOARX. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 8E) with dense PP over entire surface; IS distinctly tessellate over entire surface (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with dense PP over entire surface, IS with weak tessellation (IS = 0.5–2 d). Metanotum weakly rugulose. Mesepisternum weakly shiny, with reticulate PP over entire surface. SCL:MNL:MPL = 1:0.63:0.65. Propodeum: metapostnotum (Fig. 8F) gently inclined, with irregular sinuate ridges nearly attaining to posterior margin; junction between metapostnotum and posterior surface not carinate; lateral surface weakly rugulose; posterior surface with lateral carina on lower ⅔, without oblique carina. Coxae usual shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur with slender 2–5 teeth (n = 50). Fore wing with three submarginal cells.

ABDOMEN. Disc of T1 with sparse fine PP on medial area and lineolation over entire surface (Fig. 15E). Lineolation on T2–T5 nearly over entire surface.

First description of male

Measurements (n = 5, unit mm). BL = 4.38–5.62 (4.94 ± 0.41), WL = 3.92–5.15 (4.37 ± 0.43), HL = 1.44–1.49 (1.45 ± 0.02), HW = 1.51–1.62 (1.56 ± 0.04), IOD = 0.27–0.29 (0.28 ± 0.02), OOD = 0.29–0.31 (0.30 ± 0.01), OCD = 0.20–0.22 (0.21 ± 0.01), UOD = 0.98–1.04 (1.02 ± 0.03), MOD = 1.04–1.11 (1.08 ± 0.03), LOD = 0.80–0.91 (0.85 ± 0.05), IAD = 0.20–0.22 (0.20 ± 0.01), AOD = 0.22–0.24 (0.24 ± 0.01), CAL = 0.22–0.24 (0.24 ± 0.01), CPL = 0.31–0.38 (0.34 ± 0.02), EL = 1.07–1.11 (1.08 ± 0.02), EW = 0.44–0.49 (0.46 ± 0.02), GW = 0.36–0.44 (0.40 ± 0.03), SPL = 0.42–0.44 (0.44 ± 0.01), F1L = 0.11 (0.11 ± 0.00), F2L = 0.16–0.18 (0.17 ± 0.01), F3L = 0.16–0.18 (0.17 ± 0.01), F2W = 0.11–0.13 (0.13 ± 0.01), MsW = 1.39–1.55 (1.47 ± 0.05), SCL = 0.36–0.38 (0.36 ± 0.01), MNL = 0.18–0.20 (0.18 ± 0.01), MPL = 0.22 (0.22 ± 0.00), MtW = 1.19–1.32 (1.24 ± 0.05).

Coloration. Body black except for the following parts: mandible yellow except for apically reddish; labrum and lower half of clypeus yellow; flagellum yellowish brown ventrally; tegula yellowish brown translucent; tibiae basally and apically yellow; tibial spur yellow; tarsi yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma pale yellowish brown.

Pubescence. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: lower paraocular area sparsely tomentose.

Structure and sculpture head. Wider than long; HW:HL = 1:0.93. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.94:0.78. IOD:OOD:OCD = 1:1.06:0.73. IAD:AOD = 1:1.11. Ocellocular and paraocular areas, frons weakly shiny, with shallow reticulate PP. Supracypeal area nearly flat, dull, with reticulate PP, IS distinctly tessellate. CPL:CAL = 1:0.71. Clypeus nearly flat, with dense PP over entire surface, IS smooth (IS = 0.5–1 d). EW:GW = 1:0.88. Genal area with weak straight ridges. Malar space linear. Occiput not carinate. Postgena distinctly lineolate. Hypostomal carinae nearly parallel. Mandible edentate. Antenna short, not reaching metasoma. F2L:F2W = 1:0.74; flagellum nearly flattened ventrally.

THOARX. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum with dense PP over entire surface; IS weakly tessellate on anterior margin, otherwise smooth (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with moderately dense over entire surface, IS smooth (IS = 0.5–3 d). Metanotum weakly rugulose. Mesepisternum weakly shiny, with reticulate PP over entire surface. SCL:MNL:MPL = 1:0.50:0.61. Propodeum: metapostnotum gently inclined, with longitudinal ridges on anterior ⅔, not reaching
posterior margin; junction between metapostnotum and posterior surface not carinate and nearly smooth; lateral surface weakly rugulose; posterior surface with lateral carina on lower half, without oblique carina. Coxae usual shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur serrate. Fore wing with three submarginal cells.

Fig. 8. Lasioglossum (Hemihalictus) simplicior (Cockerell, 1931). A. ♀, lateral habitus. B. ♀, head in frontal view. C. ♂, head in frontal view. D. ♀, labrum. E. ♀, mesoscutum. F. ♀, metapostnotum. Scale bars: A = 3 mm; B–C = 1 mm; D = 0.25 mm; E–F = 0.5 mm.
ABDOMEN. Disc of T1 with sparse fine PP and without lineolation. Disc of T2–T3 with denser PP than T1; T2 without lineolation, and T3 posteriorly with weak lineolation. T4 with weak lineolation over entire surface. S7 with moderately long, apically rounded median process.

GENITALIA. Gonobase flat at bottom; gonocoxite smooth; ventral retrorse lobe tongue-like, moderately long reaching gonobasal ventral arm, with sparse short hairs ventrally.

Distribution

Flight period
Female: late May to early October.
Male: late May to middle October.

Flower record
Hydrangea macrophylla (Thunb.) Ser. (Hydrangeaceae).

Comments
The type locality of this species is Shanghai in China, so it is not surprising that L. simplicior is also distributed on the Japanese mainland. Interestingly, Japanese specimens matching the holotype were found only on the Izu Islands and not on the Japanese mainland. In the future, it may be worth while to verify whether the population of the Izu Islands indeed represents L. simplicior, after comparison of the genes of both the continental and Izu Islands populations.

This species has been recorded from Japan (Takahashi & Sakagami 1993; Goubara et al. 2004; Fukasawa & Miyano 2010). However, these records were excluded from the Japanese bee fauna because of the need to re-examine them (Tadauchi & Murao 2014; Murao 2020). In the present study, the distribution of this species in Japan was reconfirmed by comparing with the type specimen.

Lasioglossum (Hemihalictus) smilodon Ebmer & Sakagami, 1994
Figs 9, 15F, 18D, 20E

Lasioglossum (Evylaeus) smilodon Ebmer & Sakagami in Ebmer et al., 1994: 32, figs 26–33 (♀♂)
(holotype: ELKU, ♀, type locality = (TOKARA) Suwanose-Jima, Kagoshima Pref., Japan, examined).

Lasioglossum (Evylaeus) smilodon – Sakagami & Ebmer 1996: figs 1c, 3c, 4c, 5b, 6a (♀♂). — Murao et al. 2010: 31–32 (in key).

Diagnosis
Females are similar to L. (H.) ikudomei sp. nov. but are separated from them by the supraclypeal area dimly shiny (IS distinctly tessellate), the PP on the mesoscutum denser (IS=2 d in maximum) (Fig. 9E), and the lineolation of T1 more clear. In contrast, in L. (H.) ikudomei sp. nov., the supraclypeal area is more shiny (IS weakly tessellate), the PP on the mesoscutum sparser (IS=3 d in maximum) (Fig. 5E), and T1 with very weak lineolation.

Material examined

Holotype
JAPAN – Ryukyus • ♀; Kagoshima Pref., Suwanose-jima; 2 Aug. 1985; S. Ikudome leg.; ELKU.
Paratype

JAPAN – Ryukyus • 1 ♂; Kagoshima Pref., Akuseki-jima; 150–230 m a.s.l.; 2 Aug. 1985; S. Ikudome leg.; KWC.

Fig. 9. Lasioglossum (Hemihalictus) smilodon Ebmer & Sakagami, 1994. A. ♀, lateral habitus. B. ♀, head in frontal view. C. ♂, head in frontal view. D. ♀, labrum. E. ♀, mesoscutum. F. ♀, metapostnotum. Scale bars: A = 2 mm; B = 1 mm; C, E–F = 0.5 mm; D = 0.2 mm.
**Other material**


**Redescription**

**Female**

Measurements (n = 5, unit mm). BL = 4.88–5.75 (5.48 ± 0.35), WL = 4.50–5.13 (4.80 ± 0.24), HL = 1.48–1.68 (1.59 ± 0.07), HW = 1.45–1.61 (1.55 ± 0.06), IOD = 0.26–0.29 (0.28 ± 0.01), OOD = 0.24–0.26 (0.25 ± 0.01), OCD = 0.19–0.24 (0.21 ± 0.02), UOD = 0.87–1.00 (0.95 ± 0.05), MOD = 1.03–1.18 (1.13 ± 0.06), LOD = 0.77–0.90 (0.86 ± 0.05), IAD = 0.13–0.16 (0.15 ± 0.01), AOD = 0.26–0.31 (0.28 ± 0.02), CAL = 0.27–0.29 (0.28 ± 0.01), CPL = 0.32–0.35 (0.34 ± 0.01), EL = 1.60–1.85 (1.79 ± 0.11), EW = 0.39–0.45 (0.41 ± 0.03), GW = 0.29–0.35 (0.34 ± 0.03), SPL = 0.63–0.66 (0.65 ± 0.01), F1L = 0.08–0.10 (0.09 ± 0.01), F2L = 0.08–0.11 (0.09 ± 0.01), F3L = 0.08–0.10 (0.08 ± 0.01), F2W = 0.11–0.15 (0.13 ± 0.01), MsW = 1.75–2.05 (1.92 ± 0.12), SCL = 0.38–0.43 (0.41 ± 0.02), MNL = 0.23–0.28 (0.26 ± 0.02), MPL = 0.28–0.33 (0.31 ± 0.02), MtW = 1.80–2.15 (2.00 ± 0.14).

**Coloration.** Body black except for the following parts: mandible reddish brown apically; F4–F10 yellowish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga narrowly yellowish brown translucent apically. Wings transparent, veins and stigma blackish brown.

**Pubescence.** Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotal dorsum to lobe and metanotum densely tomentose; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa. Disc of T1 without short hairs. Discs of T2–T4 with moderately dense short hairs over entire surface.

**Structure and sculpture head.** Nearly as long as wide; HW:HL = 1:1.02. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.84:0.76. IOD:OOD:OCD = 1:0.88:0.76. IAD:AOD = 1:1.91. Occipital area moderately densely punctate, IS smooth (IS = 1–2.5 d). Paraocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area slightly convex, weakly shiny, with dense PP, IS weakly tessellate (IS = 1–1.5 d). CPL:CAL = 1:0.83. Clypeus nearly flat, with dense PP on upper half and larger shallow PP on lower half; IS weakly tessellate on upper half and nearly smooth on lower half (IS = 1–2 d on upper half). EW:GW = 1:0.81. Genal area to postgena with straight ridges. Malar space linear. Occiput not carinate. Hypostomal carinae nearly parallel. Mandible bidentate. Labrum (Fig. 9D): basal area approximately 1.9 × as wide as long; distal process approximately 0.6 × as long as basal area, narrow, and without lateral projection; distal keel pointed apically. Antenna short, not reaching metasoma. F2L:F2W = 1:1.48; flagellum nearly flattened ventrally.

**Thorax.** Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 9E) with dense PP over entire surface; IS distinctly tessellate over entire surface (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with sparser PP on submedian area and denser PP on marginal area; IS weakly tessellate over entire surface (IS = 1–4 d on submedian area, and 0.5–2 d on marginal area). Metanotum weakly rugulose. Mesepisternum shallow reticulate-punctate on upper area and weakly rugulate on lower area. SCL:MNL:MPL = 1:0.63:0.74.
Propodeum: metapostnotum (Fig. 9F) dimly shiny and gently inclined, with straight ridges occupying anterior half, and distinctly tessellate on posterior half; junction between metapostnotum and posterior surface not carinate, distinctly tessellate; lateral surface distinctly tessellate; posterior surface with lateral carina on lower half, without oblique carina. Coxae usual shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur pectinate, with 3–4 teeth as in Fig. 20E (n = 4). Fore wing with three submarginal cells.

**Abdomen.** Disc of T1 without distinct PP and with weak lineolation over entire surface (Fig. 15F). Disc of T2 weakly lineolate on anterior and posterior area, and nearly smooth on medial area. Discs of T3–T4 with weak lineolation over entire surface.

**Male**

**Measurements** (n = 3, unit mm). BL = 4.77–5.38 (4.90±0.36), WL = 3.77–4.46 (4.21±0.31), HL = 1.42–1.73 (1.60±0.13), HW = 1.36–1.76 (1.56±0.16), IOD = 0.27–0.31 (0.30±0.02), OOD = 0.27–0.29 (0.27±0.01), OCD = 0.16–0.22 (0.19±0.03), UOD = 0.89–1.13 (1.00±0.10), MOD = 0.93–1.24 (1.08±0.13), LOD = 0.69–1.02 (0.86±0.14), IAD = 0.16–0.24 (0.20±0.04), AOD = 0.22–0.31 (0.25±0.04), CAL = 0.27–0.31 (0.30±0.02), CPL = 0.29–0.38 (0.34±0.04), EL = 1.00–1.24 (1.13±0.10), EW = 0.38–0.47 (0.42±0.04), GW = 0.36–0.51 (0.44±0.06), SPL = 0.36–0.42 (0.46±0.04), F1L = 0.13–0.16 (0.14±0.01), F2L = 0.16–0.20 (0.18±0.02), F3L = 0.16–0.18 (0.17±0.01), F2W = 0.13–0.16 (0.15±0.01), MsW = 1.42–1.77 (1.60±0.15), SCL = 0.36–0.42 (0.39±0.03), MNL = 0.20–0.22 (0.21±0.01), MPL = 0.27–0.29 (0.28±0.01), MtW = 1.29–1.61 (1.46±0.13).

**Coloration.** Body black except for the following parts: lower half or margin dark yellow; mandible reddish brown; labrum dark yellow; pedicel and F1 yellowish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; tarsi yellowish brown or brown; metasomal terga narrowly yellowish brown translucent apically. Wings transparent, veins and stigma blackish brown.

**Pubescence.** Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotal dorsum to lobe and metanotum sparsely tomentose. Disc of T1 with sparse short hairs. Discs of T2–T4 with moderately dense short hairs over entire surface. T2–T3 with thin apical fimbriae, not clear in female.

**Structure and sculpture head.** Based on normal specimens (not cephalic polymorphism) Head nearly as long as wide; HW:HL = 1:1.03. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.92:0.79. IOD:OOD:OCD = 1:0.93:0.65. IAD:AOD = 1:1.11. Ocellocular area moderately densely puctate, IS smooth (IS = 1–4 d). Paraocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area slightly convex, weakly shiny, with moderately dense PP, IS weakly tessellate (IS = 1–2.5 d). CPL:CAL = 1:0.87. Clypeus nearly flat, with moderately dense PP over entire surface; IS smooth (IS = 1–3 d). EW:GW = 1:1.04. Genal area on lower margin and postgena with straight ridges. Malar space linear. Hypostomal carinae nearly parallel. Mandible edentate. Antenna short, not reaching metasoma. F2L:F2W = 1:0.83; flagellum nearly flattened ventrally.

**Thorax.** Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum with dense PP over entire surface; IS weakly tessellate on anterior half, nearly smooth on posterior half (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with sparser PP over entire surface; IS smooth (IS = 1–4 d). Metanotum weakly rugulose. Mesepisternum with moderately dense shallow PP on upper area and weak reticulate PP on lower area; IS smooth. SCL:MNL:MPL = 1:0.53:0.72. Propodeum: metapostnotum weakly shiny and gently inclined, with short straight ridges occupying anterior half and weakly tessellate on posterior half; junction between metapostnotum and posterior surface not carinate, weakly tessellate; lateral surface distinctly tessellate; posterior surface with lateral carina on lower half, without oblique carina. Coxae

**ABDOMEN.** Disc of T1–T3 with fine sparse PP. Disc of T1 basally weakly lineolate or smooth (paratype and 1 ♀ lineolate, but 1 ♂ not lineolate). Disc of T2–T3 weakly lineolate on anterior and posterior areas, and nearly smooth on medial area. Discs of T4 with weak lineolation over entire surface. S7 with moderately long, apically rounded median process.

**GENITALIA.** Gonobase flat at bottom, ventral arms connected with each other at upper ends; gonocoxite smooth; ventral retrorse lobe tongue-like, moderately long but not reaching gonobase, with sparse short hairs ventrally.

**Distribution**


**Flight period**

Female: April to October.

Male: August to October. The flight records of male are based on the collecting data of the original description (Ebmer et al. 1994).

**Flower records**


**Lasioglossum (Hemihalictus) spectrum** sp. nov.


Figs 10, 16A–B, 17E, 19B, 20F

**Diagnosis**

Females are similar to *L. (H.) epicinctus* but are separated from them by the frons with sparse hairs (not mixed with tomentose hairs) and the ridges of the metapostnotum long (nearly reaching posterior margin as in Fig. 10F). In contrast, in *L. (H.) epicinctus*, the frons is mixed with dense whitish tomentose hairs and the ridges of the metapostnotum are short (only present on basal area).

**Etymology**

The specific name is derived from ‘Obake’, meaning ‘ghost’ in Japanese. This species has been called “Obake-chibi-kohanabachi” in Japanese, hence its scientific name.

**Material examined**

**Holotype**

JAPAN – Kyushu ♀; Fukuoka Pref., Soeda-machi, Kyushu Univ., Hikosan Exp. St.; 33°28′48.746″ N, 130°54′55.452″ E; 18 Jul. 2014; R. Murao leg.; ELKU.

[Verbatim label: JAPAN: Kyushu / Kyushu Univ., Hikosan Exp. St. / Soeda-machi / Fukuoka Pref. / 18. vii. 2014 / Ryuki Murao leg. // N33°28′48.746″ E130°54′55.452″ // HOLOTYPE // Lasioglossum (Hemihalictus) spectrum Murao]
Paratypes
JAPAN – Honshu • 10 ♂; Iwate Pref., Morioka; 18 Jun. 1980; Y. Maeta leg., from nest; MNHAH • 20 ♂; same location as for preceding; 5 Jul. 1980; Y. Maeta leg., from nest; MNHAH • 11 ♂; Iwate Pref., Morioka, Kuriyagawa; 21 Jul. 1980; Y. Maeta leg., from nest; MNHAH • 3 ♂; same location as for preceding; 30 Jul. 1980; Y. Maeta leg., from nest; MNHAN • 2 ♂; same location as for preceding;

Fig. 10. Lasioglossum (Hemihalictus) spectrum sp. nov. A. ♀, lateral habitus. B. ♀, head in frontal view. C. ♂, head in frontal view. D. ♀, labrum. E. ♀, mesoscutum. F. ♀, metapostnotum. Scale bars: A = 3 mm; B = 1 mm; C, E–F = 0.5 mm; D = 0.2 mm.
1 Aug. 1980; Y. Maeta leg., from nest; MNHAH • 1 ♀; Shimane Pref., Okinoshima Is., Nagaobana, Chibu; 36° N, 133°03′ E; 28 Aug. 2014; K. Otsui leg.; cMUR • 1 ♀; Shimane Pref., Oda, Mt Sanbe; 35°99′ N, 132°37′ E; 6 Aug. 2015; K. Otsui leg.; cMUR • Kyushu • 2 ♀♀; Fukuoka Pref., Soeda-machi, Kyushu Univ., Hikosan Exp. St.; 33°28′48.746″ N, 130°54′55.452″ E; 13 Jun. 2014; R. Murao leg., ELKU • 1 ♀; same location as for preceding; 19 Sep. 2013; R. Murao leg.; ELKU • 2 ♀♀; same location as for preceding; 23 May 2014; R. Murao leg.; ELKU • 2 ♀♀; same location as for preceding; 18 Jul. 2014; R. Murao leg.; ELKU • ELKU • Fukuoka Pref., Tagawa-gun, Soeda-machi, Mt Hiko-san; 33°28′48.766″ N, 130°54′55.452″ E–33°29′16.788″ N, 130°54′56.461″ E; 17 Jul. 2014; R. Murao leg.; ELKU • 1 ♀; Hikosan (Buzen); 8 Jul. 1939; K. Yasumatsu leg.; ELKU; 1 ♀; same location as for preceding; 28 Jul. 1939; K. Yasumatsu leg.; ELKU • 1 ♀; Mt Hiko-san, Kajiya; 10 May 1973; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 4 Jun. 1973; K. Takeno leg.; ELKU • 1 ♀; Fukuoka Pref., Tagawa-gun, Nishi-Soeda; 18 Jun. 1968; K. Kaniyama leg.; ELKU • 1 ♀; Fukuoka, Mt Hiko; 4 May 1969; K. Kaniyama leg.; ELKU • 1 ♀; same location as for preceding; 11 Jul. 1969; K. Kaniyama leg.; ELKU • 1 ♀; same location as for preceding; 8 Jul. 1970; K. Nozato leg.; ELKU • 1 ♀; same location as for preceding; 2 May 1971; H. Makihara leg.; ELKU • 1 ♀; same location as for preceding; 7 Jul. 1971; M.T. Chujo leg.; ELKU • 2 ♀♀; same location as for preceding; 29 May 1972; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 1 Jun. 1972; K. Takeno leg.; ELKU • 2 ♀♀; same location as for preceding; 28 Apr. 1976; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 22 May 1980; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 14 Aug. 1980; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 13 Jun. 1969; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 21 Jun. 1966; A. Taketani leg.; ELKU • 2 ♀♀; same location as for preceding; 22 Jun. 1966; A. Taketani leg.; ELKU • 1 ♀; same location as for preceding; 23 Jun. 1966; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 25 Jun. 1966; A. Taketani leg.; ELKU • 1 ♀; same location as for preceding; 18 Jul. 1966; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 23 Jul. 1966; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 6 Aug. 1966; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 30 Aug. 1966; K. Takeno leg.; ELKU • 2 ♀♀; same location as for preceding; 15 May 1967; S. Kimoto leg.; ELKU • 1 ♀; same location as for preceding; 17 May 1967; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 25 May 1967; S. Kimoto leg.; ELKU • 1 ♀; same location as for preceding; 26 May 1967; S. Kimoto leg.; ELKU • 1 ♀; same location as for preceding; 3 Jun. 1967; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 4 Jul. 1967; K. Takeno leg.; ELKU • 1 ♀; same location as for preceding; 7 Jul. 1968; K. Kaniyama leg.; ELKU • 3 ♀♀; same location as for preceding; 21 Jul. 1968; K. Kaniyama leg.; ELKU • 1 ♀; same location as for preceding; 11 May 1971; Y. Hirashima leg.; ELKU • 1 ♀; same location as for preceding; 7 Jul. 1971; Y. Hirashima leg.; ELKU • 1 ♀; Fukuoka Pref., Fukuoka-shi, Nishi-ku, Genkai-jima; 33°41′24.799″ N, 130°13′52.528″ E; 19 Jul. 2009; R. Murao leg.; ELKU • 1 ♀; Fukuoka Pref., Fukuoka-shi, Nishiku, Nagahama-kai-gan; 33°36′37.64″ N, 130°15′34.919″ E; 2 Aug. 2009; R. Murao leg.; ELKU • 3 ♀♀; Fukuoka Pref., Ushikubi-dai, Ohnojo-shi; 29 Jun. 2011; R. Murao leg.; ELKU • 1 ♀; Fukuoka Pref., Fukuoka-shi, Sawara-ku, Shiibaru; 3 May 2010; R. Murao leg.; ELKU • 2 ♀♀; Fukuoka Pref., Chikushino-shi, Kouzono, Onejiruma-rindo; 4 Jul. 2009; R. Murao leg.; ELKU • 1 ♀; Fukuoka Pref., Fukuoka-shi, Higashi-ku, Shikanoshima; 14 Apr. 2013; R. Murao leg.; ELKU • 1 ♀; Saga Pref., Tosu-shi, Mt Kusasen-ya-yama; 33°25′1.353″ N, 130°26′46.819″ E; 6 Jun. 2010; R. Murao leg.; ELKU • 1 ♀; same location as for preceding; 12 Jun. 2010; R. Murao leg.; ELKU • 3 ♀♀; same location as for preceding; 19 Sep. 2013; Y. Murao leg.; ELKU • 2 ♀♀; Oita Pref., Kusumi-gun, Kokonoe-machi, Chojyabaru; 33°76′773″ N, 131°13′49.331″ E; 1050 m a.s.l.; 13 Aug. 2010; Y. Murao leg.; ELKU • 3 ♀♀; same location as for preceding; 15 May 2011; R. Murao leg.; ELKU • 1 ♀; same location as for preceding; 12 Aug. 2011; R. Murao leg.; ELKU • 1 ♀♀; Kumamoto Pref., Aso-gun, Minami-aso-mura, near Kusasenri; 30 Jul. 2009; R. Murao leg.; ELKU • 1 ♀; Kumamoto Pref., Aso-ishi, Aso-machi, Matoishi wilderness; 32°55′46.305″ N, 130°58′11.555″ E; 14 Aug. 2010; Y. Murao leg.; ELKU • 2 ♀♀; same location as for preceding; 5 Aug. 2013; Y. Murao leg.; ELKU • 1 ♀; Kumamoto Pref., Aso-gun, Minami-aso-mura, Asosan-ishi-eki; 30 Jul. 2009; R. Murao leg.; ELKU • 1 ♀; Kumamoto Pref., Aso-
gun, Nishihara-mura, near Mt Tawara; 30 Jul. 2009; R. Murao leg.; ELKU. – **Ryukyu** • 1 ♀; Kagoshima Pref., Yaku-shima Is., Onoaida; 40–200 m a.s.l.; 27 May 1982; S. Ikudome leg.; KWC.

Non-type material

JAPAN – **Hokkaido** • 1 ♀; Hamakoshimizu; 27 May 1966; MNHAH • 1 ♀; same location as for preceding; 28 May 1966; MNHAH • 2 ♀♀; same location as for preceding; 11 Jun. 1966; MNHAH • 1 ♀; same location as for preceding; 8 Jul. 1966; K. Yamauchi leg.; MNHAH • 1 ♀; same location as for preceding; 29 Sep. 1966; MNHAH • 2 ♀♀; same location as for preceding; 20 Jun. 1967; MNHAH • 2 ♀♀; same location as for preceding; 9 Aug. 1967; MNHAH • 1 ♀; Asahikawa, Inosawa; 10 Jun. 1969; MNHAH • 1 ♀; Yukomanbetsu; 9 Jul. 1968; MNHAH • 4 ♀♀; Ahosoro; 12 Jun. 2010; O. Tadauchi leg.; ELKU • 1 ♀; Ashoro-cho, Ashoro; 43°20′0″ N, 143°40′0″ E; 29 Jun. 2013; O. Tadauchi leg.; ELKU • 2 ♀♀; same location as for preceding; 2 Jul. 2013; O. Tadauchi leg.; ELKU • 1 ♀; Honbetsu-cho, Hobetsu; 43°10′0″ N, 143°35′0″ E; 1 Jul. 2013; O. Tadauchi leg.; ELKU. – **Honshu** • 1 ♀; Aomori Pref., Namioka, Mt Bonzyu; 29 May 1983; M. Yamada leg.; MNHAH • 1 ♀; same location as for preceding; 1 Aug. 1983; M. Yamada leg.; MNHAH • 2 ♀♀; Miyagi Pref., Rifu-cho; 24 May 1980; K. Gôukon leg.; MNHAH • 2 ♀♀; Ibaraki Pref., Mt Gozen-yama; 30 Jul. 1976; M. Katsukawa leg.; MNHAH • 1 ♀; Shizukawa Pref., Ito-shi, Mt Omuro; 4 May 2005; K. Gôukon leg.; SULU • 2 ♀♀; Kyoto Pref., Botanical Garden, Kyoto Univ.; 3 Jun. 1986; T. Inoue leg.; MNHAH • 4 ♀♀; Hyogo Pref., Kita-ku, Ikuno-Doujyo-cho; 4 May 2008; R. Murao leg.; ELKU • 1 ♀; Shimane Pref., Mt Sanbe; 5 May 1992; T. Yamaguchi leg.; SULE • 1 ♀; same location as for preceding; 22 May 1992; T. Yamaguchi leg.; SULE • 1 ♀; same location as for preceding; 23 Aug. 1992; T. Yamaguchi leg.; SULE • 1 ♀; Shimane Pref., Campus of Shimane Univ.; 16 Jul. 1993; Y. Okajima leg.; SULE. – **Kyushu** • 1 ♀; Kumamoto Pref., Aso-gun, Choyo-son, Setaura; 30 Apr. 1986; M. Iwata leg.; AETU • 4 ♀♀; same location as for preceding; 8 May 1986; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 30 Jun. 1986; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 11 Jul. 1986; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 13 Sep. 1986; M. Iwata leg.; AETU • 2 ♀♀; same location as for preceding; 27 Apr. 1987; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 8 Jul. 1987; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 23 Jul. 1987; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 16 Nov. 1987; M. Iwata leg.; AETU • 3 ♀♀; same location as for preceding; 30 Apr. 1988; M. Iwata leg.; AETU • 2 ♀♀; same location as for preceding; 23 May 1988; M. Iwata leg.; AETU • 3 ♀♀; same location as for preceding; 13 Jun. 1988; M. Iwata leg.; AETU • 3 ♀♀; same location as for preceding; 11 Jul. 1988; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 12 Aug. 1988; M. Iwata leg.; AETU • 6 ♀♀; same location as for preceding; 17 Apr. 1989; M. Iwata leg.; AETU • 2 ♀♀; same location as for preceding; 27 Apr. 1989; M. Iwata leg.; AETU • 5 ♀♀; same location as for preceding; 8 May 1989; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 7 Jun. 1989; M. Iwata leg.; AETU • 8 ♀♀; same location as for preceding; 19 Jun. 1989; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 29 Jun. 1989; M. Iwata leg.; AETU • 6 ♀♀; same location as for preceding; 6 Jul. 1989; M. Iwata leg.; AETU • 2 ♀♀; same location as for preceding; 14 Jul. 1989; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 19 Jul. 1989; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 6 Aug. 1989; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 25 Sep. 1989; M. Iwata leg.; AETU • 1 ♀; same location as for preceding; 6 Oct. 1989; M. Iwata leg.; AETU • 1 ♀; Kumamoto Pref., Aso-gun, Nishihara-mura; 22 Apr. 2000; M. Murase leg.; AETU • 2 ♀♀; same location as for preceding; 29 May 2000; R. Murao leg.; AETU • 2 ♀♀; same location as for preceding; 29 May 2000; M. Murase leg.; AETU • 2 ♀♀; same location as for preceding; 6 Jul. 2000; R. Murao and M. Murase leg.; AETU • 2 ♀♀; same location as for preceding; 12 May 2000; R. Murao leg.; AETU • 13 ♀♀; same location as for preceding; 26 Jun. 2000; R. Murao and M. Murase leg.; AETU • 2 ♀♀; same location as for preceding; 14 Jun. 2000; R. Murao leg.; AETU • 13 ♀♀; same location as for preceding; 17 Jul. 2000; R. Murao leg.; AETU • 2 ♀♀; same location as for preceding; 8 Sep. 2000; M. Murase leg.; AETU • 14 ♀♀; Kumamoto Pref., Kikuchi-gun, Kikuyo-machi; 22 Apr. 2000; R. Murao and M. Murase leg.; AETU • 4 ♀♀; same location
as for preceding; 29 Apr. 2000; R. Murao and M. Murase leg.; AETU • 13 ♀♀; same location as for preceding; 12 May 2000; R. Murao and M. Murase leg.; AETU • 23 ♀♀; same location as for preceding; 20 May 2000; R. Murao and M. Murase leg.; AETU • 26 ♀♀; same location as for preceding; 29 May 2000; R. Murao and M. Murase leg.; AETU • 19 ♀♂; same location as for preceding; 6 Jun. 2000; R. Murao and M. Murase leg.; AETU • 24 ♀♀; same location as for preceding; 14 Jun. 2000; R. Murao and M. Murase leg.; AETU • 26 ♀♀; same location as for preceding; 26 Jun. 2000; R. Murao and M. Murase leg.; AETU • 7 ♀♂; same location as for preceding; 8 Sep. 2000; R. Murao and M. Murase leg.; AETU • 6 ♀♂; same location as for preceding; 19 Sep. 2000; R. Murao and M. Murase leg.; AETU • 5 ♀♂; same location as for preceding; 1 Oct. 2000; R. Murao and M. Murase leg.; AETU • 11 ♀♂; same location as for preceding; 10 Oct. 2000; R. Murao and M. Murase leg.; AETU • 3 ♀♂; same location as for preceding; 27 Oct. 2000; M. Murase leg.; AETU • 3 ♀♂; same location as for preceding; 6 Nov. 2000; R. Murao and M. Murase leg.; AETU • 3 ♀♂; same location as for preceding; 14 Nov. 2000; O. Tadauchi leg.; ELKU • 1 ♀; Nagasaki Pref., Tsushima Is., Izuhara, Yora-Naiin; 13 May 2010; O. Tadauchi leg.; ELKU.

**Description**

**Female**

**Measurements** (n=5, unit mm). BL = 4.75–5.00 (4.88 ± 0.13), WL = 3.88–4.50 (4.25 ± 0.25), HL = 1.42–1.48 (1.45 ± 0.03), HW = 1.35–1.45 (1.41 ± 0.04), IOD = 0.26 (0.26 ± 0.00), OOD = 0.26–0.27 (0.26 ± 0.01), OCD = 0.16 (0.16 ± 0.00), UOD = 0.87–0.94 (0.90 ± 0.02), MOD = 1.03–1.06 (1.04 ± 0.03), LOD = 0.68–0.77 (0.74 ± 0.04), IAD = 0.13–0.15 (0.14 ± 0.01), AOD = 0.21–0.27 (0.25 ± 0.02), CAL = 0.24–0.27 (0.26 ± 0.01), CPL = 0.27–0.32 (0.29 ± 0.02), EL = 1.55–1.70 (1.62 ± 0.06), EW = 0.35–0.42 (0.39 ± 0.03), GW = 0.23–0.32 (0.27 ± 0.04), SPL = 0.55–0.60 (0.57 ± 0.02), F1L = 0.08 (0.08 ± 0.00), F2L = 0.08 (0.08 ± 0.00), F3L = 0.08 (0.08 ± 0.00), F2W = 0.11–0.13 (0.12 ± 0.01), MsW = 1.55–1.80 (1.67 ± 0.10), SCL = 0.35–0.38 (0.37 ± 0.01), MNL = 0.18–0.23 (0.20 ± 0.02), MPL = 0.28 (0.28 ± 0.00), MtW = 1.65–1.85 (1.74 ± 0.08).

**Coloration.** Body black except for the following parts: mandible reddish brown apically; F4–F10 brown (holotype) or yellowish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma brown.

**Pubescence.** Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotal dorsum to lobe and metanotum moderately densely tomentose; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa. Disc of T1 with sparse short hairs on medial area. Discs of T2–T4 with moderately dense short hairs over entire surface.

**Structure and sculpture head.** Nearly as long as wide; HW:HL = 1:1.03. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.87:0.71. IOD:OOD:OCD = 1:1.03:0.63. IAD:AOD = 1:1.83. Ocellocular area densely punctate, IS nearly smooth (IS = 0.5–2 d). Paraocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area slightly convex, weakly shiny, with dense PP; IS weakly tessellate (IS = 0.5–2 d). CPL:CAL = 1:0.89. Clypeus nearly flat, with reticulate PP on upper half and larger shallow PP on lower half; IS nearly smooth over entire surface (IS = 0.5–1 d on lower area). EW:GW = 1:0.70. Genal area to postgena with weak straight ridges. Malar space linear. Occiput not
carinate. Hypostomal carinae nearly parallel. Mandible bidentate. Labrum (Fig. 10D): basal area approximately 1.9× as wide as long; distal process approximately 0.6× as long as basal area, tongue-like, and without lateral projection; distal keel pointed apically. Antenna short, not reaching metasoma. F2L:F2W = 1:1.44; flagellum nearly flattened ventrally.

THORAX. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 10E) with dense PP over entire surface; IS distinctly tessellate on anterior half, and weakly tessellate on posterior half (but nearly smooth on posterior margin) (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with moderately dense PP over entire surface; IS nearly smooth (holotype and some paratypes) or weakly tessellate (two paratypes) over entire surface (IS = 0.5–3 d). Metanotum weakly rugulose. Mesepisternum reticulate-punctate over entire surface. SCL:MNL:MPL = 1:0.55:0.75. Propodeum: metapostnotum (Fig. 10F) dimly shiny and gently inclined, with straight ridges occupying anterior ⅔ (holotype and some paratypes) or on anterior half, with coarse tessellation on posterior ⅓ or half; junction between metapostnotum and posterior surface not carinate, coarsely tessellate; lateral surface weakly rugulose and coarsely tessellate; posterior surface with lateral carina on lower half, without oblique carina. Fore trochanter narrow, longer than wide. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur pectinate, with 2–3 teeth as in Fig. 20F (n = 10). Fore wing with three submarginal cells.

ABDOMEN. Disc of T1 with weak lineolation on basal and apical areas (not overlapping in puncture zone on medial area) (Fig. 16A), and with sparse fine PP on medial area. Disc of T2 with weak lineolation on basal and apical area, and without lineolation on medial area. T3–T4 weakly lineolate over entire surface.

Male

Measurements (n = 5, unit mm). BL = 3.62–4.54 (4.32 ± 0.43), WL = 3.08–3.77 (3.58 ± 0.27), HL = 1.20–1.38 (1.32 ± 0.07), HW = 1.20–1.40 (1.32 ± 0.07), IOD = 0.20–0.24 (0.23 ± 0.02), OOD = 0.24–0.29 (0.27 ± 0.01), OCD = 0.16–0.20 (0.17 ± 0.02), UOD = 0.82–0.91 (0.88 ± 0.03), MOD = 0.84–0.98 (0.93 ± 0.05), LOD = 0.58–0.67 (0.63 ± 0.04), IAD = 0.13–0.20 (0.16 ± 0.02), AOD = 0.18–0.22 (0.21 ± 0.02), CAL = 0.20–0.27 (0.23 ± 0.02), CPL = 0.27–0.33 (0.30 ± 0.02), EL = 0.87–0.98 (0.95 ± 0.04), EW = 0.38–0.44 (0.42 ± 0.02), GW = 0.24–0.33 (0.28 ± 0.03), SPL = 0.36–0.42 (0.39 ± 0.02), F1L = 0.09–0.13 (0.11 ± 0.01), F2L = 0.13–0.16 (0.15 ± 0.01), F3L = 0.13–0.16 (0.15 ± 0.01), F2W = 0.11–0.13 (0.12 ± 0.01), MsW = 1.16–1.32 (1.26 ± 0.06), SCL = 0.29–0.33 (0.32 ± 0.02), MNL = 0.16 (0.16 ± 0.00), MPL = 0.20–0.22 (0.21 ± 0.01), MtW = 1.03–1.26 (1.17 ± 0.08).

Coloration. Body black except for the following parts: lower half of clypeus yellow; mandible yellow except for apically reddish; labrum yellow; F1 yellowish brown or brown ventrally; pronotal lobe yellowish brown; tegula yellow translucent; tibiae basally and apically yellow; tibial spur yellow; tarsi yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma pale brown.

Pubescence. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: paraocular area, pronotal dorsum to lobe thinly tomentose. Metasomal terga with sparse, simple and short hairs over entire surface.

Structure and sculpture head. Nearly as long as wide; HW:HL = 1:1.00. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.94:0.68. IOD:OOD:OCD = 1:0.94:0.68. IAD:AOD = 1:1.43. Ocellocular area with dense PP, IS nearly smooth (IS = 0.5–2 d). Paraocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area and clepeus nearly flat, weakly shiny, with moderately dense PP; IS smooth (IS = 0.5–4 d). CPL:CAL = 1:0.76. EW:GW = 1:0.67. Genal area with weak straight ridges. Malar space linear. Occiput not carinate. Hypostomal carinae nearly parallel. Postgena longitudinal
lineolate. Mandible edentate. Antenna short, not reaching metasoma. F2L:F2W = 1:0.76; flagellum nearly flattened ventrally.

**Thorax.** Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum with dense PP over entire surface; IS smooth except for anteriorly weakly tessellate (IS=0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with moderately dense PP over entire surface; IS smooth (IS=0.5–4 d). Metanotum weakly rugulose. Mesepisternum with dense PP over entire surface; IS smooth (IS=0.5–1 d). SCL:MNL:MPL = 1:0.49:0.68. Propodeum: metapostnotum shiny and gently inclined, with short straight ridges occupying anterior half or anterior ⅔, weakly tessellate or nearly smooth on posterior half or posterior ⅓; junction between metapostnotum and posterior surface not carinate, weakly tessellate or nearly smooth; lateral surface weakly rugulose and distinctly tessellated; posterior surface with lateral carina on lower half, without oblique carina. Coxae normal shape, without tubercle. Fore trochanter narrower, longer than wide. Basitibial plate of hind leg weakly carinate marginally. Inner hind tibial spur serrate. Fore wing with three submarginal cells.

**Abdomen.** Disc of T1 without distinct PP and tessellation. Disc of T2–T4 with moderately dense fine PP, T2 without lineolation, T3–T4 with weak lineolation on apical margin. S7 with moderately long, apically rounded median process.

**Genitalia.** Gonobase flat at bottom; gonocoxite smooth; ventral retrorse lobe tongue-like, moderately long but not reaching gonobase, with sparse short hairs ventrally.

**Variation**
Disc of T1 lineolate only on basal area (Fig. 16B) in the specimens (not type series) collected from Hokkaido (northern Japan).

**Distribution**
Japan (Hokkaido, Honshu, Shikoku, Kyushu, Izu-shotô Islands, Tsu-shima Is., northern Ryukyus). Records of *Lasioglossum (Hemihalictus) pallilomum* (Strand, 1914) from the nearby continent, such as the Korean Peninsula, Russian Far East, and China (Ebmer 1978a, 1996, 2006), may correspond to this new species.

**Flight period**
Female: April to November.

Males are collected from June to August based on 46 male specimens in the late Dr Sakagami’s collection (MNHAH). These specimens were collected from the nest of this species (identified as *L. (H.) pallilomum* (Strand, 1914)).

**Flower records**
Commelinaceae: *Commelina communis* L. 
Convolvulaceae: *Ipomoea lacunosa* L. 
Cucurbitaceae: *Cucurbita* sp. 
Elaeagnaceae: *Elaeagnus umbellata* Thunb. var. "umbellata" 
Elatinaceae: *Stellaria* sp. 
Fabaceae: *Astragalus sinicus* L.; *Pueraria lobata* (Willd.) Ohwi; *Trifolium dubium* Sibth. 
Gentianaceae: *Gentiana zollingeri* Fawc. 
Geraniaceae: *Geranium carolinianum* L. 
Lamiaceae: *Clinopodium gracile* (Benth.) Kuntze; *Isodon inensus* (Thunb.) Kudô; *Lamium amplexicaule* L.; *Prunella vulgaris* L. subsp. "asiatica" (Nakai) H.Hara. 
Mazaceae: *Mazus pumilus* (Burm.f.) Steenis. 
Oleaceae: *Ligustrum japonicum* Thunb. 
Oxalidaceae: *Oxalis corniculata* L.; *Oxalis debilis* Kunth subsp. "corymbosa" (DC.) Lourteig. 
Plantaginaceae: *Veronica persica* Poir. 
Polygonaceae: *Fallopia japonica* (Houtt.) Ronse Decr. var. "japonica"; *Persicaria longiseta* (Bruijn) Kitag. 
Portulaceae: *Portulaca oleracea* L. 
Primulaceae: *Lysimachia clethroides* Duby. 
Rubiaceae: *Paederia foetida* L. 
Rutaceae: *Zanthoxylum schinifolium* Siebold & Zucc. 
Solanaceae: *Lycium chinense* Mill.; *Solanum nigrum* L. 
Vitaceae: *Ampelopsis glandulosa* (Wall.) Momiy. var. "heterophylla" (Thunb.) Momiy.; *Vitis ficifolia* Bunge.

**Habitat**

This species has been collected from various environments such as cultivated or urban areas in the lowlands, seaside, mountain areas, and semi-natural grassland. The type locality is shown in Fig. 19B.

**Comments**

In Japan, this species has been identified as *L. (H.) pallilomum* (Strand, 1914), which might be endemic to Taiwan.

*Lasioglossum (Hemihalictus) speculinum* (Cockerell, 1925)

Figs 11, 16C, 17F, 20G

*Halictus perplexans* var. *speculinus* Cockerell, 1925: 11 (holotype: USNM, ♀, typelocality = Preobrageniya Bay, Siberia, Russia, examined).

*Evylaevus (Prosopaliclus) speculinus* – Pesenko 2007b: 111. 
*Lasioglossum (Hemihaliclus) speculinum* – Murao 2017a: 461.

**Diagnosis**

Females are separated from other members of the *sexstrigatus* group occurring in Japan by a combination of the following character states: head relatively longer than wide (length/width ratio 1.07); metasoma entirely black; metasomal terga with white fimbriae on latero-apical margins; and disc of T1 basally to medially with distinct lineolation (Fig. 16C) (Murao 2017a).

**Material examined**

**Holotype**

RUSSIA – Siberia • ♀; Preobrageniya Bay; 12 Jul.; USNM.

**Other material**

JAPAN – Hokkaido • 1 ♀; Ashoro-cho, Ashoro; 43°20′0″ N, 143°40′0″ E; 2 Jul. 2013; O. Tadauchi leg.; ELKU • 1 ♀; Honbetsu-cho, Senbiri; 43°35′0″ N, 143°35′0″ E; 1 Jul. 2013; O. Tadauchi leg.; ELKU • 1 ♀; Ikeda (Tokachi); 14–16 Jul. 1953; Y. Hirashima leg.; ELKU • 2 ♀♀; Sapporo, Barato; 21 Jun. 1973;
M. Matsumoto leg.; MNHAH • 2 ♀; same location as for preceding; 28 Jun. 1973; M. Matsumoto leg.; MNHAH • 1 ♂; same location as for preceding; 22 Jul. 1973; M. Matsumoto leg.; MNHAH • 1 ♂; same location as for preceding; 3 Aug. 1973; M. Matsumoto leg.; MNHAH • 2 ♀; Hamakoshimizu; 19 Jun. 1966; H. Fukuda leg.; MNHAH • 7 ♀; same location as for preceding; 26 Jul. 1966; H. Fukuda leg.; MNHAH • 1 ♂; same location as for preceding; 27 Jul. 1966; H. Fukuda leg.; MNHAH • 7 ♂; same location as for preceding; 27 Sep. 1966; H. Fukuda leg.; MNHAH • 6 ♀; same location as for preceding; 28 Sep. 1966; H. Fukuda leg.; MNHAH • 2 ♀; same location as for preceding; 19 Jun. 1967; H. Fukuda leg.; MNHAH • 4 ♀; same location as for preceding; 6 Jul. 1967; H. Fukuda leg.; MNHAH • 1 ♂; Asahikawa, Inosawa; 25 Jun. 1969; MNHAH • 12 ♀; same location as for preceding; 27 Jun. 1969; MNHAH • 8 ♀; same location as for preceding; 13 Jul. 1969; MNHAH • 2 ♀; same location as for preceding; 26 Aug. 1969; MNHAH • 1 ♂; same location as for preceding; 10 Jun. 1970; MNHAH • 2 ♀; same location as for preceding; 25 Jul. 1970; MNHAH • 1 ♂; same location as for preceding; 11 Aug. 1970 MNHAH • 3 ♀; Sapporo; 16 Jul. 1968; S.F. Sakagami leg.; MNHAH • 3 ♀; same location as for preceding; 23 Jul. 1968; S.F. Sakagami leg.; MNHAH • 1 ♂; same location as for preceding; 3 Aug. 1968; S.F. Sakagami leg.; MNHAH • 1 ♂; Tottori Pref.; 9 Jul. 1974; M. Ishikawa leg.; MNHAH • 1 ♂; Sapporo, Hokkaido University Campus; 6 Jun. 1959; S.F. Sakagami leg.; MNHAH • 1 ♂; same location as for preceding; 11 Jun. 1959; S.F. Sakagami leg.; MNHAH • 1 ♂; same location as for preceding; 26 Jun. 1959 S.F. Sakagami leg.; MNHAH • 3 ♀; same location as for preceding; 30 Jun. 1959; S.F. Sakagami leg.; MNHAH • 1 ♂; same location as for preceding; 5 Jul. 1959; S.F. Sakagami leg.; MNHAH • 1 ♂; same location as for preceding; 6 Jul. 1959; S.F. Sakagami leg.; MNHAH • 3 ♀; same location as for preceding; 14 Jul. 1959; S.F. Sakagami leg.; MNHAH • 2 ♀; same location as for preceding; 2 Sep. 1959; S.F. Sakagami leg.; MNHAH • 4 ♀; same location as for preceding; 9 Sep. 1959; S.F. Sakagami leg.; MNHAH. – Honshu • 1 ♀; Aomori Pref., Misawa, Amagamori; 21 Jul. 1986; M. Yamada leg.; MNHAH • 1 ♂; Ibaraki Pref., Tsukuba, Kouyadai; 24 Jul. 1989; T. Matsumura leg.; MNHAH • 2 ♀; Tottori Pref., Seihaku-gun, Daisen-cho; 16 Jul. 2004; T. Sugimoto leg.; cMur • 1 ♀; Okayama Pref., Soujiya-shi; 5 Jul. 2006; Y. Kenmotsu leg.; ELKU • 1 ♂; Hiroshima Pref., Asida river; 17 May 1997; S. Nakamura leg.; ELKU. – Izu Islands • 2 ♀; Hachijo Is., Okago-Sokoto; 27 May 1964; Y. Hirashima and M. Shiga leg.; ELKU • 2 ♀; same location as for preceding; 23 Jul. 2004; R. Murao and T. Sugimoto leg.; cMur • 1 ♀; Kumamoto Pref., Kikuchi-gun, Ohzu; 2 Jul. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 14 Jul. 2003; M. Ishida leg.; AETU • 2 ♀; Kumamoto Pref., Aso-gun, Ichinomiyama-machi, Tateyamabokuya; 22 Jul. 2004; R. Murao leg.; cMur • 1 ♀; Kumamoto Pref., Aso-gun, Aso-machi, Yamadaseibubokuya; 22 Jul. 2004; T. Sugimoto leg.; cMur.

Redescription

Female

Measurements (n = 5, unit mm). BL = 4.50–5.00 (4.85 ± 0.21), WL = 4.38–4.63 (4.50 ± 0.13), HL = 1.48–1.61 (1.55 ± 0.05), HW = 1.39–1.48 (1.45 ± 0.04), IOD = 0.26–0.31 (0.28 ± 0.02), OOD = 0.26–0.29 (0.28 ± 0.01), OCD = 0.16–0.19 (0.18 ± 0.01), UOD = 0.90–1.00 (0.96 ± 0.04), MOD = 1.06–1.13 (1.08 ± 0.03), LOD = 0.74–0.84 (0.80 ± 0.04), IAD = 0.11–0.15 (0.13 ± 0.01), AOD = 0.27–0.29 (0.28 ± 0.01), CAL = 0.27–0.31 (0.30 ± 0.02), CPL = 0.27–0.34 (0.32 ± 0.03), EL = 1.65–1.75 (1.70 ± 0.05), EW = 0.35–0.39 (0.37 ± 0.02), GW = 0.23–0.35 (0.31 ± 0.05), SPL = 0.56–0.61 (0.59 ± 0.03), F1L = 0.08–0.11 (0.09 ± 0.01), F2L = 0.08–0.29 (0.08 ± 0.00), F3L = 0.08 (0.08 ± 0.00), F2W = 0.11–0.15 (0.13 ± 0.01), MsW = 1.70–1.85 (1.75 ± 0.06), SCL = 0.35–0.43 (0.38 ± 0.03), MNL = 0.15–0.20 (0.19 ± 0.02), MPL = 0.20–0.28 (0.25 ± 0.03), MtW = 1.70–2.00 (1.83 ± 0.11).

Coloration. Body black except for the following parts: mandible reddish brown apically; F8–F10 or F6–F10 yellowish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma blackish brown.
PUBESCENCE. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotal dorsum to lobe and metanotum densely tomentose; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa. Disc of T1 with sparse short hairs on medial and posterior areas. Discs of T2–T4 with moderately dense short hairs over entire surface.

STRUCTURE AND SCULPTURE HEAD. Slightly longer than wide; HW:HL = 1:1.07. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.89:0.74. IOD:OOD:OCD = 1:1:0.66. IAD:AOD = 1:2.18. Ocellocular area densely punctate, IS smooth (IS = 0.5–1.5 d). Paraocular area and frons weakly shiny, with slightly reticulate PP. Supraocular area slightly convex, weakly shiny, with moderately dense PP, IS nearly smooth (IS = 0.5–4 d). CPL:CAL = 1:0.94. Clypeus nearly flat, with dense PP on upper half and larger shallow PP on lower half; IS smooth over entire surface (IS = 0.5–1 d on upper half). EW:GW = 1:0.84. Genal area to postgena with straight ridges. Malar space linear. Occiput not carinate. Hypostomal carinae nearly parallel. Mandible bidentate. Labrum (Fig. 11C): basal area approximately 1.8 × as wide as long; distal process approximately 0.6 × as long as basal area, tongue-like, and without lateral projection; distal keel pointed apically. Antenna short, not reaching metasoma. F2L:F2W = 1:1.56; flagellum nearly flattened ventrally.

THORAX. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 11D) with dense PP over entire surface; IS weakly tessellate on anterior half, and nearly smooth on posterior half (IS = 0.5–3 d); parapsidal line a narrow groove. Mesoscutellum with sparse PP on submedian area and denser PP on marginal area; IS smooth over entire surface (IS = 1–4 d on submedian area, and 0.5–1.5 d on marginal area). Metanotum weakly rugulose. Mesepisternum with reticulate PP over entire surface. SCL:MNL:MPL = 1:0.49:0.64. Propodeum: metapostnotum (Fig. 11E) weakly shiny and gently inclined, with nearly straight ridges reaching posterior area; junction between metapostnotum and posterior surface not carinate, weakly tessellate; lateral surface weakly rugulose; posterior surface with lateral carina on lower half, without oblique carina. Coxae normal shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur with 3–4 slender teeth as in Fig. 20G (n = 6). Fore wing with three submarginal cells.

ABDOMEN. Disc of T1 with moderately dense distinct PP on medial area and with lineolation on anterior and medial areas (lineolation reaching punctate zone on medial area, but interrupted on submedial patch) (Fig. 16C). Disc of T2–T3 nearly smooth on medial area, and very weakly lineolate on remaining part. Disc of T4 with very weak lineolation over entire surface.

Male
Not examined in the present study.

Variation
Disc of T1 distinctly lineolate over entire surface in three specimens collected from Hachijo Is., Japan.

Distribution
Japan (Hokkaido, Honshu, Kyushu, Izu-shotō Islands), Korean Peninsula, Russian Far East, China.

Flight period
Female: June to September.

Males have been collected from July to August in the Korean Peninsula (Ebmer 1978b).

Flower records
The specimens examined in this paper were collected on the flowers of 26 species in 13 families as follows. Apiaceae: Aegopodium podagraria L. Araliaceae: Aralia cordata Thunb. Asteraceae:

Fig. 11. Lasioglossum (Hemihalictus) speculinum (Cockerell, 1925), ♀. A. Lateral habitus. B. Head in frontal view. C. Labrum. D. Mesoscutum. E. Metapostnotum. Scale bars: A = 3 mm; B, D–E, = 0.5 mm; C = 0.2 mm.

**Lasioglossum (Hemihalictus) sphecodicolor** Sakagami & Tadauchi, 1995  
Figs 12, 16D, 18A, 20H


**Diagnosis**

Females are similar to *L. (H.) eidmanni* (Blüthgen, 1930). According to Sakagami & Tadauchi (1995), this species is only separated from *L. (H.) eidmanni* by the metasomal terga without distinct apical fimbriae. In contrast, in *L. (H.) eidmanni*, the metasomal terga have more or less dense and well-developed apical fimbriae.

**Material examined**

**Paratypes**


**Other material**

JAPAN – Honshu • 1 ♀; Akita Pref., Omonogawa-machi, Fukai; 7 Jun. 1978; K. Baba and N. Kato leg.; ELKU • 1 ♀; Iwate Pref., Morioka, Kuriyagawa; 2 Jun. 1964; Y. Maeta leg.; ELKU • 1 ♀; same location as for preceding; 23 Jun. 1964; Y. Maeta leg.; ELKU • 1 ♀; Miyagi Pref., Onoda-cho, Arasawa; 29 Jun. 2000; K. Gōukon leg.; cGou • 1 ♀; Miyagi Pref., Kawatabi; 22 Jun. 1986; K. Gōukon leg.; cGou • 1 ♀; Yamagata Pref., Oguni-machi, Tamagawa; 24 Jun. 1981; K. Baba leg.; ELKU • 1 ♀; Fukushima Pref., Kōriyama, Nakayama; 23 May 1975; O. Tadauchi leg.; ELKU • 1 ♀; Tochigi Pref., Nasu-machi, Moriko; 22 May 1975; O. Tadauchi leg.; ELKU • 1 ♀; Gunma Pref., Nagano-hara, Asamabokujyō; 19 Jul. 1967; T. and H. Suda leg.; ELKU • 1 ♀; Niigata Pref., Miomote; 13 Jun. 1977; K. Baba leg.; ELKU • 1 ♀; same location as for preceding; 10 Jul. 1981; K. Baba leg.; ELKU • 1 ♀; Niigata Pref., Kurokawa; 9 Aug. 1970; K. Baba leg.; ELKU • 2 ♀♂; same location as for preceding; 8 Jul. 1981; K. Baba leg.; ELKU • 1 ♀; same location as for preceding; 23 May 1985; K. Baba leg.; ELKU • 1 ♀; same location as for preceding; 16 Jul. 1985; K. Baba leg.; ELKU • 1 ♀; Niigata Pref., Asahi, Koage; 15 Jun. 1985; K. Baba leg.; ELKU • 2 ♀♂; Niigata Pref., Senami; 17 Jun. 1977; K. Baba leg.; ELKU • 1 ♀; same location as for preceding; 23 Jun. 1985; K. Baba leg.; ELKU • 1 ♀; Niigata Pref., Shibata-shi, Kawahigashi; 22 May 1977; K. Baba leg.; ELKU • 1 ♀; Niigata Pref., Yuzawa, Misumata; 19 Jun. 1977; K. Baba leg.; ELKU • 1 ♀; Niigata Pref., Yuzawa-machi, Hiuchi-toge; 20 Jun. 1982; K. Baba leg.; ELKU • 3 ♀♂; Niigata Pref., Asahi, Waseda; 23 May 1985; K. Baba leg.; ELKU • 1 ♀; Yamanashi Pref., near Kawaguchi-ko, Misaka-touge; 25 Aug. 1971; H. Suda leg.; ELKU • 3 ♀♂; Nagano Pref., Aburakisawa; 5 Sep. 2014; S. Sawada leg.; cMur • 1 ♀; Ishikawa Pref., Shishiku; 3 Sep. 1972; I. Togashi leg.; ELKU. – Kyushu • 2 ♂♂; Miyazaki Pref., Ebino-Kobayashi-shi; 2 Jun. 2002; K. Mitai leg.; cMur.

**Distribution**

Japan (Hokkaido, Honshu, Izu-shotō Islands, Shikoku, Kyushu, Tsu-shima Is., northern Ryukyus).

MURAO R., Redefinition and revision of the *Lasioglossum sexstrigatus* group in Japan
**Flight period**

Female: April to October.

Male: August to October. The flight records of male are based on the collection data of the original description of this species (Sakagami & Tadauchi 1995).

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**Fig. 12.** *Lasioglossum (Hemihalictus) sphecodicolor* Sakagami & Tadauchi, 1995. **A.** ♀, dorsal habitus. **B.** ♀, head in frontal view. **C.** ♂, head in frontal view. **D.** ♀, labrum. **E.** ♀, mesoscutum. **F.** ♀, metapostnotum. Scale bars: A = 3 mm; B–C = 1 mm; D = 0.2 mm; E–F = 0.5 mm.
Flower records

Astilbe odontophylla Miq. (Saxifragaceae), Deutzia crenata Siebold & Zucc. (Hydrangeaceae), Hydrangea serrata (Thunb.) Ser. var. serrata (Hydrangeaceae), Persicaria manshuricola Kitag. (Polygonaceae), and Taraxacum of cinale Weber ex F.H.Wigg. (Asteraceae).

Lasioglossum (Hemihalictus) subsimplex sp. nov.
urn:lsid:zoobank.org:act:D4408B7F-6A84-4BB2-8B5D-8B9FDF35E031
Figs 13, 16E, 18B

Diagnosis

Females are similar to L. (H.) simplicior but are separated from them by the lineolation of T1 interrupted in part (Fig. 16E). In contrast, in L. (H.) simplicior, the lineolation of T1 is present across the entire surface (Fig. 15E).

Etymology

The specific name is derived from this species’ similarity to L. (H.) simplicior (Cockerell, 1931).

Material examined

Holotype
JAPAN – Kyushu • ♀; Oita Pref., Kusu-gun, Kokonoe-machi, Chojyabaru; 33°7’6.773″ N, 131°13’49.331″ E; 1050 m a.s.l.; 13 Aug. 2010; Y. Murao leg.; ELKU.
[Verbatim label: JAPAN: Kyushu / Chojyabaru / Kokonoe-machi / Kusu-gun / Oita Pref. / 13. VIII. 2010 / Yumi Murao leg. / Lasioglossum (Hemihalictus) subsimplex Murao]

Paratypes
JAPAN – Hokkaido • 4 ♀♂; Antaroma-Aizankei (Ishikari); 26 Jul. 1952; T. Shirôzu leg.; ELKU. – Honshu • 1 ♀; Ishikawa Pref., Negami-machi, Yamaguchi; 29 Aug. 1996; I. Togashi leg.; ELKU. – Kyushu • 1 ♀; Oita Pref., Kokonoe-machi, Chojyabaru; 33°7’6.773″ N, 131°13’49.331″ E; 1050 m a.s.l.; 13 Aug. 2010; R. Murao leg.; cMur • 2 ♀♂; Oita Pref., Kusu-gun, Kokonoe-machi, Handa-kogen; 33°7’58.702″ N, 131°14’2.412″ E; 9 Jul. 2013; R. Murao leg.; ELKU • 1 ♀; Oita Pref., Takeda, Nagayu; 28 Apr. 2009; O. Tadauchi leg.; ELKU • 1 ♀; Oita Pref., Mt Kuju; 9 May 2009; O. Tadauchi leg.; ELKU • 2 ♀♂; same location as for preceding; 23 May 2009; O. Tadauchi leg.; ELKU • 2 ♀♂; Oita Pref., Shimohanda; 12 Apr. 1975; O. Tadauchi leg.; ELKU • 1 ♀; Miyaji-Bochu (Mt Aso); 23 Jun. 1959; Y. Hirashima leg.; ELKU • 1 ♀; Kumamoto Pref., Aso-machi, Aso; 28 May 2001; A. Yamada leg.; ELKU • 1 ♀; same location as for preceding; 16 Jun. 2001; A. Yamada leg.; ELKU • 1 ♀; Miyazaki Pref., Takachiho-cho, Gokasyo-Sobosan; 27 Apr. 2003; F. Kodoi leg.; ELKU • 2 ♀♂; Miyakonojo-Hyuga, Naka; 13 Apr. 1958; Y. Hirashima leg.; ELKU • 2 ♀♂; Makizono (Satsuma); 11 Apr. 1959; Y. Hirashima leg.; ELKU • 2 ♀♂; Kagoshima; 4 Apr. 1949; Y. Hirashima leg.; ELKU • 1 ♀; Sata (Osumi) Magome-Hetsuka; 24 May 1952; T. Esaki and Y. Hirashima leg.; ELKU • 1 ♀; Kagoshima Pref., Kii-re-machi, Kiire; 31 Mar. 1975; O. Tadauchi leg.; ELKU • 1 ♀; Kagoshima Pref., Miyakonojo; 29 Mar. 1959; Y. Maeta leg.; ELKU. – Ryukyu • 1 ♀; Kagoshima Pref., Yaku-shima, Onoaida; 40–200 m a.s.l.; 27 Jul. 1982; S. Ikudome leg.; KWC.

SOUTH KOREA – Jeollabuk-do • 1 ♀; Namweon-gun, Sannae-myon, Sanlyong-ri; 14 May 1991; O. Tadauchi leg.; ELKU. – Gyeongsangnam-do • 1 ♀; Hamyang-gun, Macheongmeon, Samjeong-ri; 12 May 1991; T. Saigusa leg.; ELKU.
**Description**

**Female**

**Measurements** (n = 5, unit mm). BL = 5.00–5.63 (5.45 ± 0.27), WL = 4.63–5.00 (4.80 ± 0.14), HL = 1.55–1.65 (1.59 ± 0.04), HW = 1.61–1.74 (1.69 ± 0.05), IOD = 0.27–0.29 (0.28 ± 0.01), OOD = 0.26–0.29 (0.28 ± 0.01), OCD = 0.19 (0.19 ± 0.00), UOD = 1.00–1.06 (1.02 ± 0.03), MOD = 1.16–1.26 (1.21 ± 0.04), LOD = 0.87–0.94 (0.89 ± 0.03), IAD = 0.13–0.15 (0.14 ± 0.01), AOD = 0.29–0.31 (0.30 ± 0.01), CAL = 0.27–0.29 (0.29 ± 0.01), CPL = 0.32–0.34 (0.34 ± 0.01), EL = 1.85–1.90 (1.88 ± 0.03), EW = 0.42–0.48 (0.46 ± 0.03), GW = 0.29–0.32 (0.31 ± 0.02), SPL = 0.63–0.68 (0.65 ± 0.02), F1L = 0.08–0.10 (0.09 ± 0.01), F2L = 0.08 (0.08 ± 0.00), F3L = 0.08 (0.08 ± 0.00), F2W = 0.13–0.15 (0.14 ± 0.01), MsW = 1.85–2.10 (1.98 ± 0.09), SCL = 0.40–0.45 (0.42 ± 0.03), MNL = 0.23–0.25 (0.25 ± 0.01), MPL = 0.28–0.30 (0.28 ± 0.01), MtW = 1.95–2.15 (2.04 ± 0.07).

**Coloration.** Body black except for the following parts: mandible reddish brown apically; F4–F10 yellowish brown (holotype and 18 paratypes) or brown (remaining paratypes) ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma brown.

**Pubescence.** Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotal dorsum to lobe and metanotum moderately densely tomentose; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa. Disc of T1 with sparse short hairs on medial area. Discs of T2–T4 with moderately dense short hairs over entire surface.

**Structure and Sculpture.**

**Head.** Wider than long; HW:HL = 1:0.94. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.84:0.74. IOD:OOD:OCD = 1:0.99:0.69. IAD:AOD = 1:2.11. Ocellocular area densely pustulate, IS nearly smooth (IS = 0.5–2 d). Paraocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area slightly convex, dimly shiny, with dense PP; IS distinctly tessellate (IS = 0.5–2 d). CPL:CAL = 1:0.86. Clypeus nearly flat, with dense PP on upper half and larger yellowish brown on lower half; IS nearly smooth over entire surface (IS = 0.5–1 d on upper half). EW:GW = 1:0.68. Genal area to postgena with straight ridges. Malar space linear. Occiput not carinate. Hypostomal carinae nearly parallel. Mandible bidentate. Labrum (Fig. 13C): basal area approximately 2 × as wide as long; distal process approximately 0.7 × as long as basal area, narrow, and without lateral projection; distal keel narrow, pointed apically. Antenna short, not reaching metasoma. F2L:F2W = 1:1.76; flaggellum nearly flattened ventrally.

**Thorax.** Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 13D) with dense PP over entire surface; IS distinctly tessellate nearly over entire surface (but weakly tessellate on posterior margin) (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with moderately dense PP over entire surface; IS weakly tessellate over entire surface (IS = 0.5–3 d in paratypes). Metanotum weakly rugulose. Mesoepisternum with reticulate PP on upper area and weak rugulae on lower area. SCL:MNL:MPL = 1:0.58:0.67. Propodeum: metapostnotum (Fig. 13E) weakly shiny and gently inclined, with straight ridges reaching to near posterior margin; junction between metapostnotum and posterior surface not carinate, weakly tessellate; lateral surface weakly rugulose and distinctly tessellate; posterior surface with lateral carina on lower half, without oblique carina. Coxae normal shape, without tubercle. Fore trochanter narrower, longer than wide. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur with 3–4 slender teeth (n = 27). Fore wing with three submarginal cells.

**Abdomen.** Disc of T1 with weak lineolation interrupted on medial area (Fig. 16E), and with sparse fine PP on medial area; submedian patch distinct and nearly smooth. Disc of T2–T3 with very weak lineolation on apical half, and without lineolation on basal half. T4 very weakly lineolate over entire surface.
Male
Unknown.

Distribution
Japan (Hokkaido, Honshu, Kyushu, northern Ryukyus), Korean Peninsula.

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Fig. 13. *Lasioglossum (Hemihalictus) subsimplicior* sp. nov., ♀. A. Lateral habitus. B. Head in frontal view. C. Labrum. D. Mesoscutum. E. Metapostnotum. Scale bars: A = 3 mm; B = 1 mm; C = 0.25 mm; D–E = 0.5 mm.
Flight period
Female: late March to August.

Flower records

Brassica rapa L. var. oleifera DC. (Brassicaceae), 
Eleutherococcus sp. (Araliaceae), 
Erigeron annuus (L.) Pers. (Asteraceae), 
Ixeirdium dentatum (Thunb.) Tzvelev subsp. dentatum (Asteraceae).

Habitat
This species has been collected mainly from mountainous areas in western Japan. One of the collecting sites in Japan is shown in Fig. 19A.

Lasioglossum (Hemihalictus) tadauchii Murao, 2012
Fig. 18D


Diagnosis
Females are similar to L. (H.) taeniolellum. According to Murao (2012), this species is separated from L. (H.) taeniolellum by the postgena having distinct lineolation over entire surface, the distal process of labrum without lateral projection (Murao 2012: fig. 5), and T1 with short hairs and fine PP on disc (Murao 2012: fig. 7). In contrast, in L. (H.) taeniolellum, the lineolation on postgena does not reach the apical margin, the distal process of labrum with horn-like lateral projection (Fig. 14D), and T1 nearly smooth. Male unknown.

Distribution
Japan (central Ryukyus: Amami-Ohshima Is.).

Flight records
Female: April to October.

Flower records
Two species in two families were reported as floral records by Murao (2012).

Habitat
This species has been collected from around subtropical forests in mountainous area (Murao 2012). It may prefer humid environments.

Lasioglossum (Hemihalictus) taeniolellum (Vachal, 1903)
Figs 2B, 14, 16F, 18C, 20I


Halictus subfamiliaris Strand, 1910: 191 (holotype: ZMHB, ♀, type locality = Japan, examined).

Synonymy by Ebmer (1978b).

Diagnosis

Females are similar to *L. (H.) tadauchii*, but are separated from them by the lineolation on postgena not reaching the apical margin, the distal process of the labrum with a horn-like lateral projection (Fig. 14D), and T1 nearly smooth. In contrast, in *L. (H.) tadauchii*, the postgena have a distinct lineolation across the entire surface, the distal process of the labrum is without a lateral projection (Murao 2012: fig. 5), and T1 has short hairs and fine PP on the disc (Murao 2012: fig. 7).

Fig. 14. *Lasioglossum (Hemihalictus) taeniolellum* (Vachal, 1903). **A.** ♀, lateral habitus. **B.** ♀, head in frontal view. **C.** ♂, head in frontal view. **D.** ♀, labrum. **E.** ♀, mesoscutum. **F.** ♀, metapostnotum. Scale bars: A = 3 mm; B–C, E–F = 0.5 mm; D = 0.25 mm.
Material examined

JAPAN – Hokkaido • 1 ♀; Ashoro-cho, Ashoro; 43°20′0″ N, 143°40′0″ E; 2 Jul. 2013; O. Tadauchi leg.; ELKU • 1 ♀; Nishiashoro (Tokachi); 8 Aug. 1955; Y. Hirashima leg.; ELKU • 1 ♀; Ashoro-gun, Metou; 19 Jun. 1957; M. Takahashi leg.; ELKU • 1 ♀; Sapporo, Hokkaido University Campus; 3 Sep. 1969; S.F. Sakagami leg.; MNHAH • 1 ♀; Ebetsubuto; 8 May 1974; M. Ishikawa leg.; MNHAH. – Honshu • 13 ♀♂; Iwate Pref., Morioka, Kuriyagawa; 16 May 1964; Y. Maeta leg.; ELKU • 4 ♀♂; same location as for preceding; 2 Jun. 1964; Y. Maeta leg.; ELKU • 1 ♀; Miyagi Pref., Sendai, Tsuchitoi; 10 May 1977; K. Gôukon leg.; MNHAH • 1 ♀; Ibaraki Pref., Tsukuba, Koyadai; 22 Jun. 1989; T. Matsumura leg.; MNHAH • 1 ♀; Saitama Pref., Urawa; 24 Aug. 1968; T. Nambu leg.; ELKU • 3 ♀♂; Yamanashi Pref., Nakagawa; 9 Apr. 1962; T. Saigusa leg; commented by Dr Hiroshima as “compared with the type of Halictus taeniolellus Vachal ♀” the under label; ELKU • 1 ♀; Kyoto Pref., Expt. Internal. Forest, Kyoto Univ.; 10 Aug. 1985; T. Kakutani leg.; MNHAH. – Kyushu • 4 ♀♂; Fukuoka Pref., Fukuoka-shi, Higashi-ku, Hakozaki, Kyushu Univ.; 10 Jul. 2004; T. Sugimoto leg.; ELKU • 18 ♀♂; same location as for preceding; 18 Apr. 2006; R. Murao leg.; cMur • 1 ♀; same location as for preceding; 15 Jul. 2006; R. Murao leg.; cMur • 14 ♀♂, 5 ♀♂; same location as for preceding; 5 Jul. 2009; R. Murao leg.; cMur • 1 ♀; same location as for preceding; 29 Mar. 2012; R. Murao leg.; ELKU • 2 ♀♂; Kumamoto Pref., Aso-gun, Aso-machi; 20 Apr. 2001; A. Yamada and M. Nomura leg.; AETU • 1 ♀; same location as for preceding; 20 May 2001; A. Yamada leg.; AETU • 2 ♀♂; same location as for preceding; 28 May 2001; A. Yamada and M. Nomura leg.; AETU • 7 ♀♂; same location as for preceding; 7 Jun. 2001; A. Yamada and M. Nomura leg.; AETU • 3 ♀♂; same location as for preceding; 16 Jun. 2001; A. Yamada and M. Nomura leg.; AETU • 1 ♀; same location as for preceding; 2 Jul. 2001; A. Yamada leg.; AETU • 1 ♀; same location as for preceding; 17 Sep. 2001; M. Nomura leg.; AETU • 2 ♀♂; same location as for preceding; 20 Oct. 2001; M. Nomura leg.; AETU • 1 ♀; same location as for preceding; 20 Nov. 2001; M. Nomura leg.; AETU • 3 ♀♂; Kumamoto Pref., Aso-gun, Choyo-son; 13 Jun. 2003; D. Kozai and M. Ishida leg.; AETU • 2 ♀♂; same location as for preceding; 21 Jun. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 26 Jun. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 7 Jun. 2001; A. Yamada and M. Nomura leg.; AETU • 3 ♀♂; same location as for preceding; 2 Jul. 2003; M. Ishida leg.; AETU • 1 ♀; Kumamoto Pref., Aso-gun, Choyo-son, Setaura; 9 Jul. 1996; M. Yamada leg.; AETU • 1 ♀; Kumamoto Pref., Aso-gun, Ozu-machi–Choyo-son; 7–9 Oct. 2002; K. Mitai leg.; ELKU • 1 ♀; Kumamoto Pref., Kikuchi-gun, Ozu-machi; 2 Apr. 2002; T. Sugimoto leg.; AETU • 1 ♀; same location as for preceding; 25 Apr. 2002; Y. Terada leg.; AETU • 1 ♀; same location as for preceding; 21 Jul. 2002; Y. Terada leg.; AETU • 3 ♀♂; same location as for preceding; 18 Apr. 2003; M. Ishida leg.; AETU • 2 ♀♂; same location as for preceding; 28 Apr. 2003; M. Ishida and D. Kozai leg.; AETU • 1 ♀; same location as for preceding; 9 May 2003; M. Ishida leg.; AETU • 3 ♀♂; same location as for preceding; 20 May 2003; D. Kozai and M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 29 May 2003; M. Ishida leg.; AETU • 3 ♀♂; same location as for preceding; 5 Jun. 2003; D. Kozai leg.; AETU • 6 ♀♂; same location as for preceding; 13 Jun. 2003; M. Ishida and D. Kozai leg.; AETU • 1 ♀; same location as for preceding; 21 Jun. 2003; D. Kozai leg.; AETU • 2 ♀♂; same location as for preceding; 7 Jun. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 26 Jul. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 18 Jul. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 23 Jul. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 13 Jun. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 26 Jun. 2003; M. Ishida leg.; AETU • 1 ♀; same location as for preceding; 30 Oct. 2003; M. Ishida leg.; AETU • 2 ♀♂; Kumamoto Pref., Aso-gun, Nishihara-mura; 22 Apr. 2000; R. Murao leg.; AETU • 4 ♀♂; same location as for preceding; 29 Apr. 2000; R. Murao and M. Murase leg.; AETU • 1 ♀; same location as for preceding; 12 May 2000; M. Murase leg.; AETU • 2 ♀♂; same location as for preceding; 20 May 2000; R. Murao and M. Murase leg.; AETU • 4 ♀♂; same location as for preceding; 29 May 2000; R. Murao and M. Murase leg.; AETU • 1 ♀; same location as for preceding; 11 Jun. 2000; R. Murao leg.; AETU • 1 ♀; same location as for preceding; 14 Jun. 2000; M. Murase leg.; AETU • 1 ♀; same location as for preceding; 28 Aug. 2000; M. Murase leg.; AETU • 1 ♀; same location as for preceding; 1 Oct. 2000; M. Murase
leg.; AETU • 1 ♀; same location as for preceding; 6 Nov. 2000; M. Murase leg.; AETU • 2 ♀♀; same location as for preceding; 8 Dec. 2000; M. Murase leg., AETU • 8 ♀♀; Kumamoto Pref., Kikuchi-gun, Kikujo-machi; 22 Apr. 2000; R. Murao and M. Murase leg.; AETU • 8 ♀♀; same location as for preceding; 29 Apr. 2000; R. Murao and M. Murase leg.; AETU • 6 ♀♀; same location as for preceding; 12 May 2000; R. Murao and M. Murase leg.; AETU • 7 ♀♀; same location as for preceding; 20 May 2000; R. Murao and M. Murase leg.; AETU • 8 ♀♀; same location as for preceding; 29 May 2000; R. Murao and M. Murase leg.; AETU • 5 ♀♀; same location as for preceding; 14 Jun. 2000; M. Murase leg.; AETU • 5 ♀♀; same location as for preceding; 26 Jun. 2000; R. Murao and M. Murase leg.; AETU • 1 ♀; same location as for preceding; 6 Jul. 2000; M. Murase leg.; AETU • 1 ♀; same location as for preceding; 16 Aug. 2000; M. Murase leg.; AETU • 1 ♀; same location as for preceding; 28 Aug. 2000; R. Murao leg.; AETU • 1 ♀; same location as for preceding; 8 Sep. 2000; M. Murase leg.; AETU • 2 ♀♀; same location as for preceding; 19 Sep. 2000; M. Murase leg.; AETU • 5 ♀♀; same location as for preceding; 10 Oct. 2000; R. Murao and M. Murase leg.; AETU • 9 ♀♀; same location as for preceding; 19 Oct. 2000; R. Murao and M. Murase leg.; AETU • 5 ♀♀; same location as for preceding; 27 Oct. 2000; R. Murao and M. Murase leg.; AETU • 1 ♀; same location as for preceding; 6 Nov. 2000; M. Murase leg.; AETU.

Redescription

Female

MEASUREMENTS (n=5, unit mm). BL = 5.38–5.50 (5.35 ± 0.21), WL = 4.38–4.88 (4.55 ± 0.21), HL = 1.42–1.48 (1.45 ± 0.02), HW = 1.55–1.61 (1.57 ± 0.03), IOD = 0.29–0.32 (0.30 ± 0.01), OOD = 0.24–0.29 (0.26 ± 0.02), OCD = 0.18–0.21 (0.19 ± 0.01), UOD = 0.97–1.00 (0.97 ± 0.01), MOD = 1.13–1.16 (1.15 ± 0.02), LOD = 0.84–0.90 (0.87 ± 0.02), IAD = 0.15–0.16 (0.15 ± 0.01), AOD = 0.29–0.31 (0.30 ± 0.01), CAL = 0.26–0.27 (0.26 ± 0.01), CPL = 0.29–0.32 (0.31 ± 0.02), EL = 1.70–1.75 (1.72 ± 0.03), EW = 0.39–0.45 (0.42 ± 0.03), GW = 0.61–0.65 (0.63 ± 0.01), SPL = 0.61–0.65 (0.63 ± 0.01), F1L = 0.08–0.10 (0.09 ± 0.01), F2L = 0.08 (0.08 ± 0.00), F3L = 0.08 (0.08 ± 0.00), F2W = 0.11–0.13 (0.13 ± 0.01), Msw = 1.80–1.85 (1.83 ± 0.03), SCL = 0.38–0.40 (0.40 ± 0.01), MNL = 0.20–0.23 (0.22 ± 0.01), MPL = 0.25–0.30 (0.28 ± 0.02), MtW = 1.85–2.05 (1.90 ± 0.09).

COLORATION. Body black except for the following parts: mandible reddish brown apically; all flagellar segments brown or F5–F10 yellowish brown ventrally; tegula yellowish brown translucent; tibial spur yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma yellowish brown.

PUBESCENCE. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: pronotal dorsum to lobe and metanotum moderately densely tomentose; posterior surface of propodeum sparsely tomentose; hind trochanter, femur, and tibia mixed with plumose hairs, forming scopa. Disc of T1 without short hairs on medial area. Discs of T2–T4 with moderately dense short hairs over entire surface.

STRUCTURE AND SCULPTURE HEAD. Wider than long; HW:HL = 1:0.92. Vertex rounded in frontal view. MOD : UOD : LOD = 1:0.85:0.76. IOD : OOD : OCD = 1:0.87:0.65. IAD : AOD = 1:1.96. Ocellocellar area densely punctate, IS nearly smooth (IS = 0.5–2 d). Paracocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area slightly convex, dimly shiny, with dense PP; IS distinctly tessellate (IS = 0.5–1.5 d). CPL : CAL = 1:0.85. Clypeus nearly flat, with reticulate PP nearly over entire surface, its PP gradually sparser toward lower area; IS nearly smooth on lower area (IS = 0.5–2 d on lower area). EW : GW = 1:0.71. Genal area with weak straight ridges. Malar space linear. Occiput not carinate. Hypostomal carinae nearly parallel. Postgena weakly tessellate, sometimes nearly smooth in part. Mandible bidentate, Labrum (Fig. 14D): basal area approximately 2.1 × as wide as long; distal process approximately 0.7 × as long as basal area, narrow, and with horn-like lateral projection; distal keel narrow, pointed apically. Antenna short, not reaching metasoma. F2L : F2W = 1:1.56; flagellum nearly flattened ventrally.
THORAX. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum (Fig. 14E) with dense PP over entire surface; IS distinctly tessellate over entire surface (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with sparse PP on submedian area and denser PP on marginal area; IS nearly smooth over entire surface (IS = 1–3 d on submedian area, and 0.5–1 d on marginal area). Metanotum weakly rugulose. Mesepisternum with reticulate PP over entire surface. SCL:MNL:MPL = 1:0.56:0.71. Propodeum: metapostnotum (Fig. 14F) weakly shiny and gently inclined, with short straight ridges occupying only anterior half, with weak tessellation on posterior half; junction between metapostnotum and posterior surface not carinate, weakly tessellate; lateral surface weakly rugulose and distinctly tessellate; posterior surface with lateral carina on lower half, without oblique carina. Coxae normal shape, without tubercle. Fore trochanter narrow, longer than posterior. Basitibial plate of hind leg carinate marginally. Inner hind tibial spur with 2–5 slender teeth as in Fig. 20I (n = 13). Fore wing with three submarginal cells.

ABDOMEN. Disc of T1 nearly smooth over entire surface (Fig. 16F). Disc of T2 very weakly lineolate in part (only anterior or both anterior and posterior area). T3–T4 very weakly lineolate over entire surface.

First description of male

Measurements (n = 5, unit mm). BL = 3.92–5.23 (4.68 ± 0.56), WL = 3.23–4.00 (3.71 ± 0.27), HL = 1.18–1.44 (1.32 ± 0.10), HW = 1.27–1.51 (1.41 ± 0.09), IOD = 0.22–0.31 (0.28 ± 0.03), OOD = 0.24–0.27 (0.25 ± 0.01), OCD = 0.18–0.22 (0.20 ± 0.02), UOD = 0.84–0.98 (0.92 ± 0.05), MOD = 0.89–1.07 (1.00 ± 0.06), LOD = 0.60–0.76 (0.70 ± 0.06), IAD = 0.18–0.22 (0.20 ± 0.02), AOD = 0.18–0.22 (0.20 ± 0.02), CAL = 0.20–0.27 (0.24 ± 0.02), CPL = 0.29–0.31 (0.30 ± 0.01), EL = 0.87–1.09 (0.98 ± 0.07), EW = 0.40–0.47 (0.43 ± 0.03), GW = 0.27–0.33 (0.29 ± 0.03), SPL = 0.31–0.38 (0.35 ± 0.03), F1L = 0.09–0.11 (0.11 ± 0.01), F2L = 0.18–0.22 (0.21 ± 0.02), F3L = 0.18–0.24 (0.22 ± 0.02), F2W = 0.11–0.13 (0.12 ± 0.01), MsW = 1.19–1.48 (1.34 ± 0.10), SCL = 0.29–0.38 (0.33 ± 0.03), MNL = 0.13–0.18 (0.16 ± 0.02), MPL = 0.22–0.24 (0.23 ± 0.01), MtW = 1.03–1.29 (1.19 ± 0.02).

COLORATION. Body black except for the following parts: lower half of clypeus yellow; mandible except for apically and labrum yellow; mandible apically reddish; all flagellar segments yellowish brown ventrally; pronotal lobe yellow or yellowish brown; tegula yellow translucent; tibiae basally and apically yellow; tibial spur yellow; tarsi yellow; metasomal terga broadly yellowish brown translucent apically. Wings transparent, veins and stigma yellowish brown.

PUBESCENCE. Body hairs whitish, and covered with erect and sparse straight or fine branched hairs except for the following parts: lower paraocular area, supraclypeal area, pronotal dorsum to lobe and metanotum thinly tomentose. Metasomal terga with sparse, simple and short hairs over entire surface.

STRUCTURE AND SCULPTURE HEAD. Wider than long; HW:HL = 1:0.93. Vertex rounded in frontal view. MOD:UOD:LOD = 1:0.92:0.70. IOD:OOD:OCD = 1:0.90:0.71. IAD:AOD = 1:1.00. Ocellocular area with reticulate PP. Paraocular area and frons weakly shiny, with shallow reticulate PP. Supraclypeal area nearly flat, weakly shiny, with moderately dense PP over entire surface; IS nearly smooth (IS = 0.5–3 d). Clypeus weakly shiny, with dense PP over entire surface; IS nearly smooth (IS = 0.5–1 d). CPL:CAL = 1:0.78. EW:GW = 1:0.68. Genal area with weak straight ridges. Malar space linear. Occiput not carinate. Hypostomal carinae nearly parallel. Postgena longitudinal lineolate. Mandible edentate. Antenna short, not reaching metasoma. F2L:F2W = 1:0.58; flagellum nearly flattened ventrally.

THORAX. Dorsolateral angle of pronotum obtuse; lateral surface without ridges; lateral lobe rounded. Tegula ovoid, nearly smooth. Mesoscutum with dense PP over entire surface; IS smooth except for anteriorly weakly tessellate (IS = 0.5–2 d); parapsidal line a narrow groove. Mesoscutellum with sparse PP over entire surface; IS smooth (IS = 1–5 d). Metanotum weakly rugulose. Mesepisternum with moderately dense PP over entire surface; IS smooth (IS = 1–3 d). SCL:MNL:MPL = 1:0.49:0.68. Propodeum: metapostnotum shiny and gently inclined, with short straight ridges occupying only
anterior half, weakly tessellate or nearly smooth on posterior half; junction between metapostnotum and posterior surface not carinate, weakly tessellate or nearly smooth; lateral surface weakly rugulose and distinctly tessellate; posterior surface with lateral carina on lower half, without oblique carina. Coxae normal shape, without tubercle. Fore trochanter narrow, longer than wide. Basitibial plate of hind leg weakly carinate marginally. Inner hind tibial spur without tooth. Fore wing with three submarginal cells.

**ABDOMEN.** Disc of T1 without distinct PP and tessellation. Disc of T2–T4 with sparse fine PP, T2 without lineolation, T3 with weak lineolation on apical half, and T4 with weak lineolate over entire surface. S7 with moderately long, apically truncate or rounded median process.

**GENITALIA.** Gonobase flat at bottom; gonocoxite smooth; ventral retrorse lobe tongue-like, moderately long but not reaching gonobase, with sparse short hairs ventrally.

**Distribution**
Japan (Hokkaido, Honshu, Izu-shotō Islands, Shikoku, Kyushu, Tsu-shima Is., northern Ryukyus), Korean Peninsula.

**Flight period**
Female: April to December.

Males have been collected from August to September in the Korean Peninsula (Ebmer 1978b). In Japan, males have been collected at one site (Hakozaki Campus, Kyushu University, Japan, Fig. 19C) from June to July.

**Flower records**

**Habitat**
This species has been collected mainly in cultivated or urban lowland areas and semi-natural grassland in Kyushu, western Japan. One of the collection sites in Japan is shown in Fig. 19C.
Fig. 16. 1st metasomal tergum, ♀. A–B. Lasioglossum (Hemihalictus) spectrum sp. nov. C. L. (H.) speculinum (Cockerell, 1925). D. L. (H.) sphecodicolor Sakagami & Tadauchi, 1995. E. L. (H.) subsimplicior sp. nov. F. L. (H.) taeniolellum (Vachal, 1903). Scale bars: 1 mm.
**Unknown species in Japan**

*Lasioglossum (Hemihalictus)* sp.

**Material examined**

JAPAN – Kyushu • 2 ♀; Fukuoka Pref., Soeda-machi, Kyushu Univ., Hikosan Exp. St.; 33°28′48.746″ N, 130°54′55.452″ E; 13 Jun. 2013; R. Murao leg.; ELKU • 2 ♂; Kumamoto Pref., Aso-shi, Aso-machi, Matoishi wilderness; 37°27′15″ N, 128°1′10″ E; 5 Aug. 2013; R. Murao leg.; ELKU.

**Comments**

This unknown species has been identified as *Lasioglossum (Hemihalictus) sexstrigatus* (Schenck, 1869) in Japan. According to DNA analysis in the present study, the pair-wise sequence divergence between Japanese and European specimens was quite clear (7.3–7.9% between *L. sexstrigatus* and *L.* sp. in Table 1). Morphologically, females of this species are slightly different from *L. (H.) sexstrigatus* as the mesoscutum has a weak tessellation over the entire surface (in contrast, in *L. (H.) sexstrigatus*, the mesoscutum is nearly smooth on the posterior half).

In the present study, I could not conclude that this unknown species is either undescribed or a known species, but this will be examined in further studies.

---

**Fig. 19.** Habitat in Japan. A. Chojyabaru, Kokonoe-machi, Kusu-gun, Oita Pref., Kyushu, Japan. 
Key to species of the Lasioglossum (Hemihalictus) sexstrigatus group in Japan

Females

1. Metasoma generally reddish over entire surface (Fig. 12A) ...............................................................

   \[L. (H.) sphecodicolor\] Sakagami & Tadauchi, 1995

   – Metasoma generally black over entire surface ................................................................................ .. 2

2. Disc of T1 with lineolation (e.g., Fig. 15A–B) .................................................................................. 3

   – Disc of T1 without lineolation (e.g., Fig. 15C) ............................................................................. ... 12

3. Lineolation of T1 nearly over entire surface (Fig. 15A, D–F) .......................................................... 4

   – Lineolation of T1 over part of surface (Figs 15B, 16A–C, E) ........................................................... 7

4. Ridges of metapostnotum short, reaching anterior half (Fig. 9F) (distribution: northern to central
   Ryukyus) .........................................................................

   \[L. (H.) smilodon\] Ebmer & Sakagami, 1994

   – Ridges of metapostnotum long, reaching posterior margin (Figs 4F, 7F, 8F) ................................... 5

5. IS of mesoscutum nearly smooth on posterior margin; T1 with silky dull luster...............................

   \[L. (H.) ohei\] Hirashima & Sakagami, 1966

   – IS of mesoscutum with distinct tessellation over entire surface; T1 with enamel-like luster ........... 6

6. Head longer than wide or slightly longer than wide (HL/HW ratio = 1.02–1.07, n = 5) (restricted to
   coastal sand dunes) .............................................................................

   \[L. (H.) frigidum\] Sakagami & Ebmer, 1996

   – Head wider than long or nearly as long as wide (HL/HW ratio = 0.94–1.0, n = 5) (distribution: Izu-
   shotô Islands) ...................................................................................

   \[L. (H.) simplicior\] Cockerell, 1931

7. Ridges of metapostnotum short, reaching anterior half (Fig. 5F) (distribution: Ryukyus) ............... 8

   – Ridges of metapostnotum long, reaching posterior margin or anterior ¾ (Figs 10F, 11E, 13E) ....... 10

8. Disc of T1 with sparse short hairs (Murao 2012: fig. 7).....\[L. (H.) tadauchii\] Murao, 2012 (in part)

   – Disc of T1 without distinct hairs (Fig. 15B)..................................................................................... 9

9. Disc of T1 with very weak lineolation on only basally...............................................................

   \[L. (H.) amamiense\] Ebmer & Sakagami, 1994 (in part)

   – Disc of T1 with very weak lineolation interrupted on medial area (Fig. 15B)...............................

   \[L. (H.) ikudomei\] Murao sp. nov.
10. IS of mesoscutum medially to posteriorly smooth (Fig. 11D) ............................................................
   – IS of mesoscutum medially to posteriorly tessellate (Figs 10E, 13D) .............................................

11. Metapostnotum dimly shiny, with distinct tessellation among ridges (Fig. 10F) ...................................
   – Metapostnotum shinier, without distinct tessellation among ridges (Fig. 13E) ......................

12. Disc of T1 with distinct hairs and PP (Fig. 15C) .............................................................................
   – Disc of T1 without hairs and PP (Fig. 16F) ..................................................................................

13. IS of mesoscutum medially to posteriorly nearly smooth (Fig. 6D) ............................................... 
   – IS of mesoscutum medially to posteriorly distinctly tessellate (Murao 2012: fig. 3) (distribution:
   central Ryukyus) ..................................................................................................................

14. Distal process of labrum with horn-like lateral projection (Fig. 14D) ............................................
   – Distal process of labrum without lateral projection (e.g., Fig. 4D) ..............................................

15. Ridges of metapostnotum short, reaching anterior half (Murao et al. 2010: fig. 1N) (distribution:
   central Ryukyus) ..................................................................................................................
   – Ridges of metapostnotum long, reaching posterior margin ......................................................

Males
1. Metasoma generally reddish over entire surface as in female (Fig. 12A) ........................................
   – Metasoma generally black over entire surface ........................................................................

2. Clypeus nearly black, without distinct yellow spot on lower half (Figs 5C, 9C) .....................
   – Clypeus with a distinct yellow spot on lower half (Figs 4C, 7C, 8C, 10C, 14C) ............

3. F2 long, over 1.5 × F1 ........................................................................................................
   – F2 short, under 1.5 × F1 ........................................................................................................

4. F2 long, over 1.5 × F1 ........................................................................................................
   – F2 short, approximately or under 1.5 × F1 ..............................................................................

5. Head wider than long, HL/HW ratio 0.90–0.96 ........................................................................
   – Head longer than wide, HL/HW ratio 1.04–1.07 (restricted to coastal sand dunes) ............

6. F2 nearly as long as F1; labrum without basal elevation ............................................................
   – F2 1.4–1.5 × F1; labrum with a distinct basal elevation (Fig. 2B) ..............................................

Discussion

The morphological differences among species of the *sexstrigatus* group often depend on subtle characteristics such as the sculpture of the mesoscutum or T1 and the length of metapostnotal ridges, etc., as described in the diagnosis of each species and the above Key. These characteristics are useful only in female specimens. Generally, in *Lasioglossum*, the interspecific differences are clearer in males than in the female, particularly using male genitalia. However, the interspecific differences in males of the *sexstrigatus* group is unclear even when comparing male genitalia. In addition, it is even more difficult to identify the species, owing to the male cephalic polymorphism. Among the European *Lasioglossum*, *L. (H.) pleurospeculum* Herrmann, 2001, *L. (H.) sabulosum* (Warncke, 1986), and *L. (H.) sexstrigatus* belong to the *sexstrigatus* group (Herrmann & Doczkal 1999; Herrmann 2001). According to Herrmann & Doczkal (1999) and Herrmann (2001), the tooth of the inner hind tibial spur in females is one of the useful characters that distinguishes *L. (H.) sexstrigatus* and the other two species: short teeth in *L. (H.) sexstrigatus*, long teeth in *L. (H.) pleurospeculum* and *L. (H.) sabulosum*. However, for the Japanese species, there is no interspecific difference in the female inner hind tibial spur (Fig. 20). In the present study, I examined part of the DNA barcoding region of the cytochrome oxidase subunit I gene of mitochondrial DNA gene for some species of the *sexstrigatus* group. The interspecies sequence divergences were larger than intraspecies divergences as in Table 1. Thus, DNA barcodes are quite useful in judging the interspecific differences in the *sexstrigatus* group. The *sexstrigatus* group is diverse in eastern Asia, and its diversity is unclear in the Oriental Region. Accumulation and utilisation of DNA data will be a very effective tool in the progress of taxonomic study of the *sexstrigatus* group in the Oriental Region.

Acknowledgments

I would like to express my thanks to Dr Yulia Astafurova (Zoological Institute, Russian Academy of Sciences, St Petersburg, Russia), Mr Fritz Gusenleitner (Biologiezentrum/Oberösterreichisches Landesmuseum, Linz, Austria), Mr Maximilian Schwarz (Linz, Austria), Mr Andreas Werner Ebmer (Puchenau, Austria), Dr Andreas Taeger (Senckenberg Deutsches Entomologisches Institut, Münchenberg, Germany), Mr Andrew Liston (Senckenberg Deutsches Entomologisches Institut, Münchenberg, Germany) and Dr Frank Koch (Museum für Naturkunde an der Humboldt Universität zu Berlin, Germany) for their help in my examining their Palaearctic and Oriental *Lasioglossum* collections. I would especially like to thank Prof. Emeritus Dr Osamu Tadauchi (Fukuoka, Japan), Prof. Emeritus Dr Yasuo Maeta (Shimane, Japan), Mr Katsuo Gōukon (Miyagi, Japan), Dr Shuichi Ikudome (Kagoshima Women’s College, Japan), Dr Ryoichi Miyanaga (Shimane University, Japan), Dr Toshiharu Mita (Kyushu University, Japan), and Mr Makio Iwata (Kumamoto, Japan), Mr Keiichi Otsui (Fukuoka, Japan) for the loan of valuable specimens and literature. I also thank Editage (www.editage.com) for the English language editing. This research was supported by the JSPS Institutional Program for Young Researcher Overseas Visits.

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Blüthgen P. 1934. Schwedisch-chinesische wissenschaftliche Expedition nach den nordwestlichen Provinzen Chinas unter Leitung von Dr. Sven Hedin und Prof. Sü Ping-chang: Insekten, gesammelt vom


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https://doi.org/10.1007/BF01731581


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Appendix 1

Taxa scored for cladistic analysis.

Ingroups

1. *Lasioglossum clypearis* group: *L. (H.) buccale* (Pérez, 1903); *L. (H.) punctatissimum* (Schenck, 1853).

2. *L. limbellus* group: *L. (H.) brevicorne* (Schenck, 1870); *L. (H.) limbellum* (Morawitz, 1876).


6. *L. pauperatum* group: *L. (H.) pauperatum* (Brullé, 1832); *L. (H.) transitorium* (Schenck, 1870).

7. *L. semilucens* group: *L. (H.) kuroshio* Takahashi & Sakagami, 1993; *L. (H.) lucidulum* (Schenck, 1861); *L. (H.) metis* Ebmer, 2002; *L. (H.) minutissimum* (Kirby, 1802); *L. (H.) pumilum* Sakagami & Tadauchi, 1995; *L. (H.) quadrinotatulum* (Schenck, 1861); *L. (H.) sakagamii* Ebmer, 1978; *L. (H.) sulcatulum longifacies* Sakagami & Tadauchi, 1995; *L. (H.) transpositum* (Cockerell, 1925); *L. (H.) zunaga* Sakagami & Tadauchi, 1995.


9. *L. villosulum* group: *L. (H.) villosulum* (Kirby, 1802); *L. (H.) truncaticolle* (Morawitz, 1877).

10. *L. alphenum* group (= *L. (Sudila)*): *L. (H.) alphenum* (Cameron, 1897); *L. (H.) aulacophorum* (Strand, 1913); *L. (H.) bidentatum* (Cameron, 1898).

Outgroup

*Dialictus*: *L. (D.) aeratum* (Kirby, 1802); *L. (D.) leucopus* (Kirby, 1802).
Appendix 2
Characters scored for cladistic analysis.

**Color**
1. Body color: (0) black, (1) metallic green.

**Head**
2. Head length / width ratio in female: (0) under 0.95, (1) 0.96–1.09, (2) over 1.10, (3) over 1.50.
3. Genal process of male head: (0) present as variation, (1) absent.
4. Yellow spot on male clypeus: (0) present and distinct, (1) present, but indistinct, (2) absent.
5. Supraclypeus: (0) nearly flat, (1) distinctly swelled.
6. Shape of male clypeus: (0) normal, (1) extremely long, (2) incurved on lower margin, (3) bidentate as variation.
7. Malar area: (0) linear, (1) distinctly longer than wide.
8. Length of male antenna: (0) long, attaining to metasoma, (1) short, not attaining to metasoma.
9. F1 : F2 in male: (0) F2 1.3–1.6 × F1, (1) F2 1.7–2.0 × F1, (2) F2 as long as F1.
10. Color of male labrum: (0) black, (1) yellow.
11. Basal elevation of male labrum: (0) present, not depressed centrally, (1) present, depressed centrally, (2) absent.
12. Distal process of female labrum: (0) narrow, (1) moderately broad.
13. Distal process of male labrum: (0) present, (1) absent.

**Mesosoma**
14. Mesepisternum tubercle: (0) present, (1) absent.
15. Oblique carina of propodeum: (0) present, (1) absent.
16. Hairs on posterior surface of propodeum: (0) sparse, (1) dense and tomentose.
17. Coxa in male: (0) normal, (1) modified.
18. Basitibial plate of hind leg in male: (0) present with weak carina marginally, (1) absent.
19. Inner hind tibial spur in male: (0) serrate, (1) dentate.

**Metasoma**
20. Color: (0) black, (1) reddish.
21. Apical fimbriae on female metasomal terga: (0) present, (1) absent.
22. Hairs on male metasomal sterna: (0) normal, without hair tuft, (1) present, with thin hair tuft on S4–S5, (2) present, with thin hair tuft on S3–S5.

23. Median process of male S8: (0) present, well developed (over S7), (1) present, less developed (not over S7), (2) absent.

**Male genitalia**

24. Gonobase ventral arm: (0) connected, (1) not connected.

25. Sculpture of gonocoxite: (0) smooth, (1) lineolate or reticulate.

26. Shape of gonostylus: (0) small and bud-like, (1) small and elongate, (2) large and rounded.

27. Gonostylus hairs: (0) simple, (1) simple, mixed with fine branched hairs.

28. Length of ventral retrorse lobe: (0) long, attaining to gonobase, (1) short or moderately long, not attaining to gonobase.

29. Sculpture of ventral retrorse lobe: (0) smooth, (1) lineolate.

30. Ventral retrorse lobe hairs: (0) sparse to dense short hairs over entire surface, (1) a few short hairs, (2) moderately dense setae apically.

31. Angle of penis valve in lateral view: (0) gently curved, (1) angulate.

32. Cleft of penis valve: (0) present, (1) absent.
Appendix 3 (continued on next page)

Data matrix for cladistic analysis.

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### Appendix 3 (continued)

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Appendix 4
List of species of *Lasioglossum* Cockerell, 1897 used in the DNA analysis.

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