



**Research article**

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**Taxonomy of the ant genus *Nesomyrmex* Wheeler  
(Formicidae, Myrmicinae) in the Afrotropical region,  
with a review of current species groups and description of a  
new species of the *N. angulatus* group from Mozambique**

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**Abstract.** This study reviews the taxonomy of the ant genus *Nesomyrmex* Wheeler, 1910 in the Afrotropical region. Previous revisionary studies are discussed and four species groups are proposed on the basis of external morphology. The *N. angulatus* group contains seven species that are widely distributed throughout the whole Afrotropical region, with one species also occurring in the Palaearctic and Malagasy regions. The *N. cataulacoides* group is monotypic, with one morphologically bizarre species found in Equatorial rain forests. The *N. humerosus* group is also monotypic and occurs in East Africa. The last and by far most species-rich group is the *N. simoni* group that contains 17 species, all of which are endemic to South Africa. The four groups are defined for the first time for the region, and an illustrated identification key is provided. Furthermore, the *N. angulatus* group is more thoroughly reviewed. One new species from Mozambique is described, *N. inhaca* sp. nov., and species accounts for the other six are provided. Also, an illustrated identification key to the species of the *N. angulatus* group is presented.

**Keywords.** Ant taxonomy, identification key, Inhaca Island, new species, systematics.

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## Introduction

*Nesomyrmex* Wheeler, 1910 is a moderately-sized genus of myrmicine ants distributed in the tropics and subtropics of the Neotropical, Afrotropical and Malagasy regions (Guénard & Economo 2016). It was traditionally grouped within the tribes Leptothoracini or Formicoxenini, but the most recent phylogeny of the Myrmicinae revealed it to be a member of the Crematogastrini (Ward *et al.* 2015). At present, *Nesomyrmex* contains 55 valid species (Bolton 2016), of which 21 are Neotropical, 25 Afrotropical, and nine Malagasy (Kempf 1959; Brandão 1991; Bolton 2003; Mbanyana & Robertson 2008; Csósz & Fisher 2015). Two extinct species are known from Dominican Amber (De Andrade *et al.* 1999). For most of its existence *Nesomyrmex* was treated by most authorities as either a subgenus or junior synonym of the much more diverse and widespread genus *Leptothorax* Mayr, 1855 (e.g., Wheeler 1922; Kempf 1959; Bolton 1982). Relatively recently, Bolton (2003) revived it from synonymy with *Leptothorax* and raised it to full genus status by providing a thorough generic diagnosis and distinguishing it from other closely related genera. The most recent multi-locus molecular phylogeny of the subfamily Myrmicinae strongly supports the monophyly of the genus (Ward *et al.* 2015). The alpha taxonomy of the genus in the Afrotropical region is in reasonably good condition. Bolton (1982) revised the genus (as part of *Leptothorax*), recognised nine species, and gave a species level key to the then known species. Later, Snelling (1992) described the morphologically bizarre *N. cataulacoides* from Cameroon as a tenth Afrotropical species. More recently, Mbanyana & Robertson (2008) provided an extensive revision of the *Nesomyrmex* of southern Africa. They recognised 20 species from South Africa, of which they described 15 as new, leading to a total of 25 *Nesomyrmex* species for the whole Afrotropical region.

During a recent taxonomic survey of the *Tetramorium* Mayr, 1855 fauna of Mozambique hosted in the ant collection of the Museum of Comparative Zoology, Harvard University Cambridge, the first author encountered a strange looking series from Inhaca Island. After a closer look it became apparent that this series was accidentally mislabelled as *Tetramorium*, but clearly belonged to the genus *Nesomyrmex*. While the genus identity was easily assessed, it was impossible to identify the species on the basis of the currently available taxonomic identification keys (Bolton 1982; Mbanyana & Robertson 2008). A more thorough examination and comparison with reference material revealed that the material belonged to a new, undescribed species. In this study we review the *Nesomyrmex* species groups found in the Afrotropical region and provide an identification key to the four proposed groups herein: *N. angulatus* group, *N. cataulacoides* group, *N. humerosus* group and *N. simoni* group. In addition, we review the *N. angulatus* group and describe the new species as *N. inhaca* sp. nov. In order to aid in identification we also provide an illustrated key to all species of the *N. angulatus* group.

## Abbreviations of depositories

The collection abbreviations given below follow Evenhuis (2016). The material upon which this study is based on is located and/or was examined at the following institutions:

- BMNH = The Natural History Museum, London, U.K.
- HLMD = State Museum of Hesse Darmstadt, Darmstadt, Germany
- MCZC = Museum of Comparative Zoology, Harvard University, Cambridge, U.S.A.
- KSMA = Museum of Arthropods, King Saud University, Riyadh, Kingdom of Saudi Arabia
- SAMC = Iziko Museum of Cape Town, Cape Town, South Africa
- ZFMK = Zoological Research Museum Alexander Koenig, Bonn, Germany

## Material and methods

All new type material and all imaged specimens can be uniquely identified by specimen-level codes affixed to each pin (e.g., CASENT0493062). Digital colour montage images were created using a JVC KY-F75 digital camera and Syncroscopy Auto-Montage software (version 5.0), or a Leica DFC 425 camera in combination with the Leica Application Suite software (version 3.8), or a Canon EOS 7D in

combination with Helicon Focus (version 6.5). All images used in this publication are available online and can be seen on AntWeb (<http://www.antweb.org>). General terminology for ant morphology is based on Bolton (1982, 1994), and for the description of degrees of inclination of pilosity we follow Wilson (1955). When referring to the terminology for the description of surface sculpturing we follow Harris (1979) and Bolton (1982). Morphometric measurements were performed with a Leica M165 equipped with an orthogonal pair of micrometres at magnifications ranging from 63× to 120×. Measurements and indices are presented as minimum and maximum values with arithmetic means in parentheses, and all measurements are expressed in mm to two decimal places. The measurements and indices used in this study follow Hita Garcia & Fisher (2015).

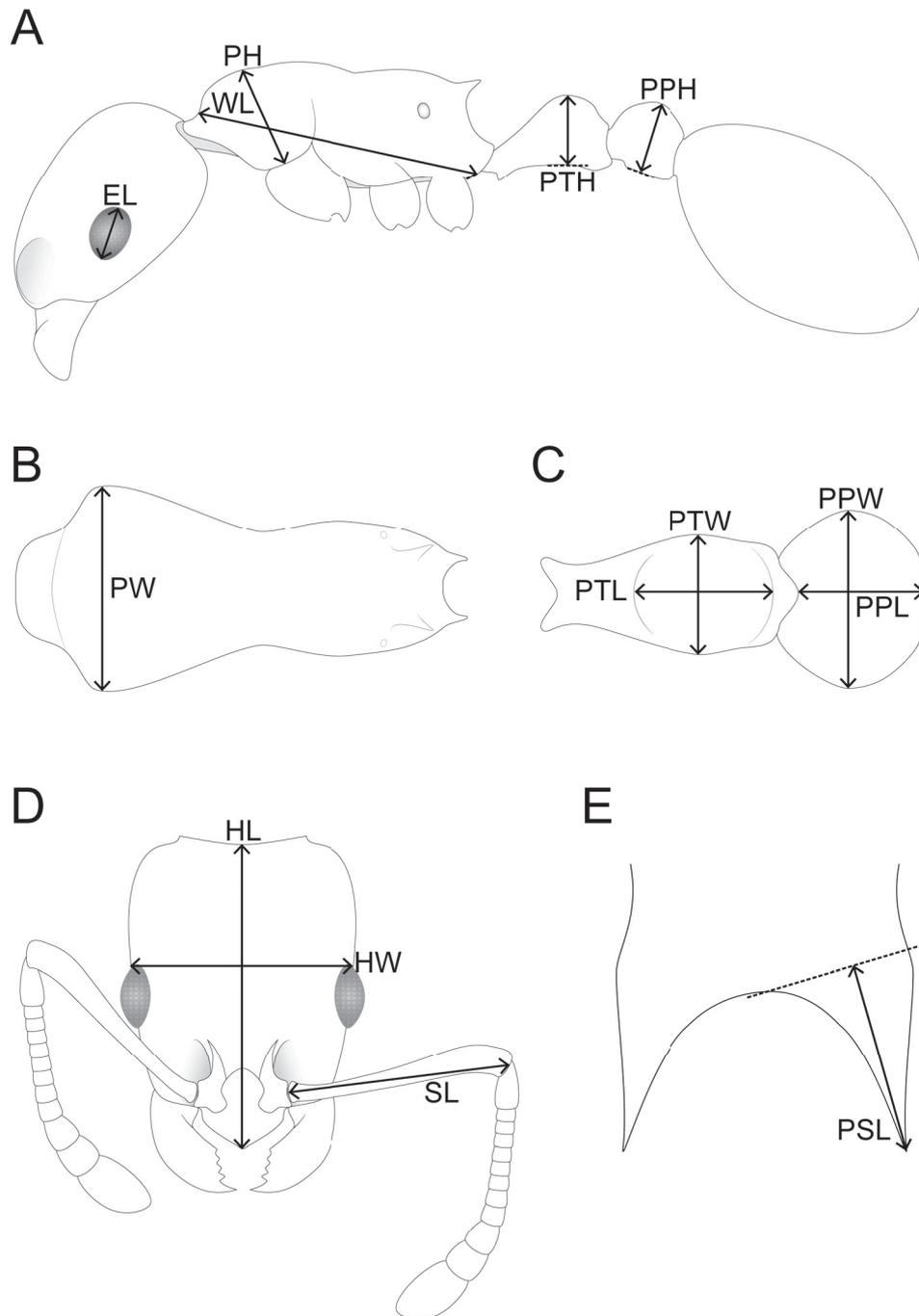
### Measurements:

- EL = Eye length: maximum diameter of compound eye measured in oblique lateral view  
 HL = Head length: maximum distance from the midpoint of the anterior clypeal margin to the midpoint of the posterior margin of the head, measured in full-face view. Impressions on the anterior clypeal margin and the posterior head margin reduce head length  
 HW = Head width: width of the head directly behind the eyes measured in full-face view  
 PH = Pronotal height: maximum height of the pronotum measured in lateral view  
 PPH = Postpetiole height: maximum height of the postpetiole measured in lateral view  
 PPL = Postpetiole length: maximum length of the postpetiole measured in dorsal view  
 PPW = Postpetiole width: maximum width of the postpetiole measured in dorsal view  
 PSL = Propodeal spine length: in dorsofrontal view the tip of the measured spine, its base, and the centre of the propodeal concavity between the spines must all be in focus. Using a dual-axis micrometre the spine length is measured from the tip of the spine to a virtual point at its base where the spine axis meets orthogonally with a line leading to the median point of the concavity.  
 PTH = Petiolar node height: maximum height of the petiolar node measured in lateral view from the highest (median) point of the node to the ventral outline. The measuring line is placed at an orthogonal angle to the ventral outline of the node  
 PTL = Petiolar node length: maximum length of the dorsal face of the petiolar node from the anterodorsal to the posterodorsal angle, measured in dorsal view excluding the peduncle  
 PTW = Petiolar node width: maximum width of the dorsal face of the petiolar node measured in dorsal view  
 PW = Pronotal width: maximum width of the pronotum measured in dorsal view  
 SL = Scape length: maximum scape length excluding basal condyle and neck  
 WL = Weber's length: diagonal length of the mesosoma in lateral view from the posteroventral margin of propodeal lobe to the anterior-most point of pronotal slope, excluding the neck

### Indices:

- OI = Ocular index:  $EL/HW \times 100$   
 CI = Cephalic index:  $HW/HL \times 100$   
 SI = Scape index:  $SL/HW \times 100$   
 DMI = Dorsal mesosoma index:  $PW/WL \times 100$   
 LMI = Lateral mesosoma index:  $PH/WL \times 100$   
 PSLI = Propodeal spine index:  $PSL/HL \times 100$   
 LPeI = Lateral petiole index:  $PTL/PTH \times 100$   
 DPeI = Dorsal petiole index:  $PTW/PTL \times 100$   
 LPpI = Lateral postpetiole index:  $PPL/PPH \times 100$   
 DPpI = Dorsal postpetiole index:  $PPW/PPL \times 100$   
 PPI = Postpetiole index:  $PPW/PTW \times 100$

The distribution data given below is based on the previous revisions or treatments of the genus for the Afrotropical region (Bolton 1982; Snelling 1992; Mbanyana & Robertson 2008), as well as on data mined from AntWeb and other online databases (Bolton 2016; Guénard & Economo 2016).



**Fig. 1.** Schematic line drawings of *Nesomyrmex inhaca* sp. nov., illustrating the measurements used. **A.** Body in profile with measuring lines for EL, WL, PH, PTH and PPH. **B.** Mesosoma in dorsal view with measuring line for PW. **C.** Petiole and postpetiole in dorsal view with measuring lines for PTL, PTW, PPL and PPW. **D.** Head in full-face view with measuring lines for HL, HW and SL. **E.** Dorsocaudal view of the propodeum with measuring line for PSL.

## Results

Class Hexapoda Blainville, 1816  
Order Hymenoptera Linnaeus, 1758  
Suborder Apocrita Latreille, 1810  
Infraorder Aculeata Latreille, 1802  
Superfamily Formicoidea Latreille, 1804  
Family Formicidae Latreille, 1809  
Subfamily Myrmicinae Lepeletier de Saint-Fargeau, 1835  
Tribe Crematogastrini Forel, 1893  
  
Genus *Nesomyrmex* Wheeler, 1910

### Review of Afrotropical *Nesomyrmex* species groups

#### *Synoptic list of Afrotropical Nesomyrmex species*

##### *Nesomyrmex angulatus* species group

*Nesomyrmex angulatus* (Mayr, 1862) [Algeria, Botswana, Comoros, Djibouti, Egypt, Eritrea, Ethiopia, Ghana, Kenya, Libya, Madagascar, Malawi, Mozambique, Niger, Nigeria, Oman, Saudi Arabia, Seychelles, South Africa, Sudan, Uganda, Tanzania, Tunisia, Yemen, Zimbabwe]  
= *Nesomyrmex angulatus ilgii* (Forel, 1894)  
= *Nesomyrmex latinodis* (Mayr, 1895)  
= *Nesomyrmex angulatus concolor* (Santschi, 1914)

*Nesomyrmex denticulatus* (Mayr, 1901) [South Africa]

*Nesomyrmex evelynae* (Forel, 1916) [Burkina Faso, Central African Republic, Ghana, D.R. Congo, Kenya, Uganda]

*Nesomyrmex grisoni* (Forel, 1916) [Central African Republic, D.R. Congo, Ghana]

*Nesomyrmex inhaca* sp. nov. [Mozambique]

*Nesomyrmex innocens* (Forel, 1913) [D.R. Congo, Kenya]

*Nesomyrmex stramineus* (Arnold, 1948) [South Africa, Swaziland]

##### *Nesomyrmex cataulacoides* species group

*Nesomyrmex cataulacoides* (Snelling, 1992) [Cameroon, Kenya]

##### *Nesomyrmex humerosus* species group

*Nesomyrmex humerosus* (Emery, 1896) [Kenya, Tanzania, Yemen]

##### *Nesomyrmex simoni* species group

*Nesomyrmex antoinetteae* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex braunsi* (Forel, 1912) [South Africa]

*Nesomyrmex cederbergensis* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex entabeni* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex ezantsi* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex inye* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex karooensis* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex koebergensis* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex larsenae* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex mcgregori* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex nanniae* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex njengelanga* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex ruani* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex saasveldensis* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex simoni* (Emery, 1895) [South Africa]

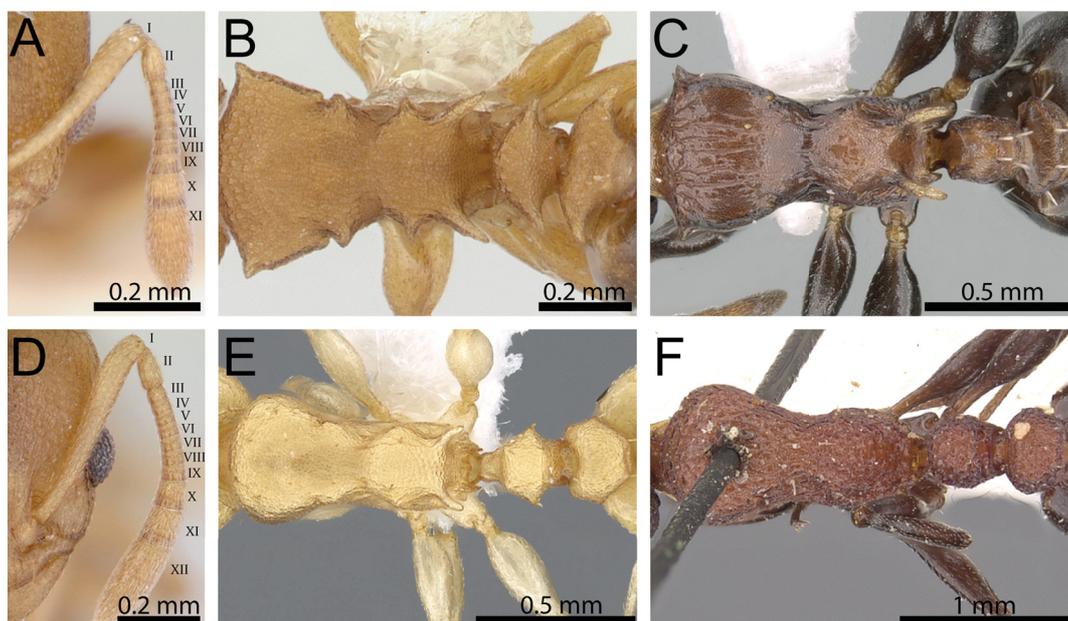
*Nesomyrmex tshiguvhoae* Mbanyana & Robertson, 2008 [South Africa]

*Nesomyrmex vannoorti* Mbanyana & Robertson, 2008 [South Africa]

**Identification key to Afrotropical *Nesomyrmex* species groups (workers)**

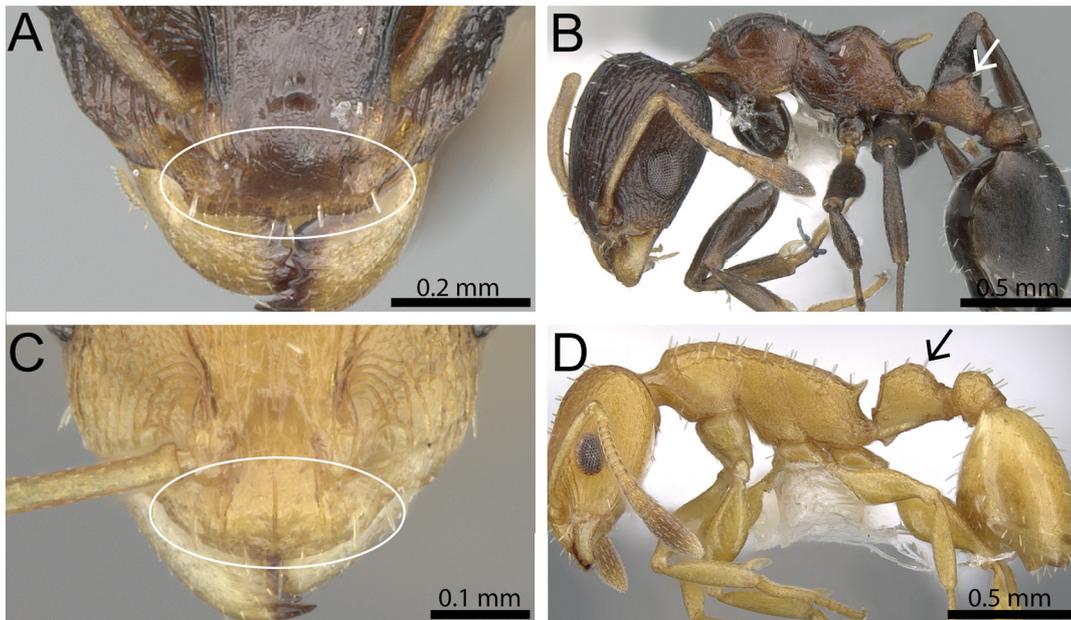
The following newly developed identification key to species groups is loosely based on Bolton (1982) and Mbanyana & Robertson (2008), and also incorporates ideas from Snelling (1992).

1. Antennae with 11 segments (Fig. 2A); petiolar node and postpetiole with conspicuous and very well developed lateral spines (Fig. 2B) .....*N. cataulacoides* group
  - Antennae with 12 segments (Fig. 2D); petiolar node and postpetiole never with conspicuous and very well developed lateral spines as above, at most petiolar node with small, lateral denticles (Fig. 2C, E–F) .....2
2. Anterior clypeal lobe short, flat-margined, and never convex, lobe with a small median triangular projection (Fig. 3A); pronotum anterodorsally sharply marginate, with sharp, dentate corners (Fig. 2C); petiole barrel-shaped with very weakly developed, short and triangular petiolar node (Fig. 3B) .....*N. humerosus* group
  - Anterior clypeal lobe always conspicuously developed, usually convex and variably rounded, sometimes flat, but never with a small median triangular projection (Fig. 3C); pronotum anterodorsally either rounded or weakly marginate but without sharp, dentate corners (Fig. 2E–F); petiole variably shaped, but never as above, usually with very well developed petiolar node (Fig. 3D) .....3

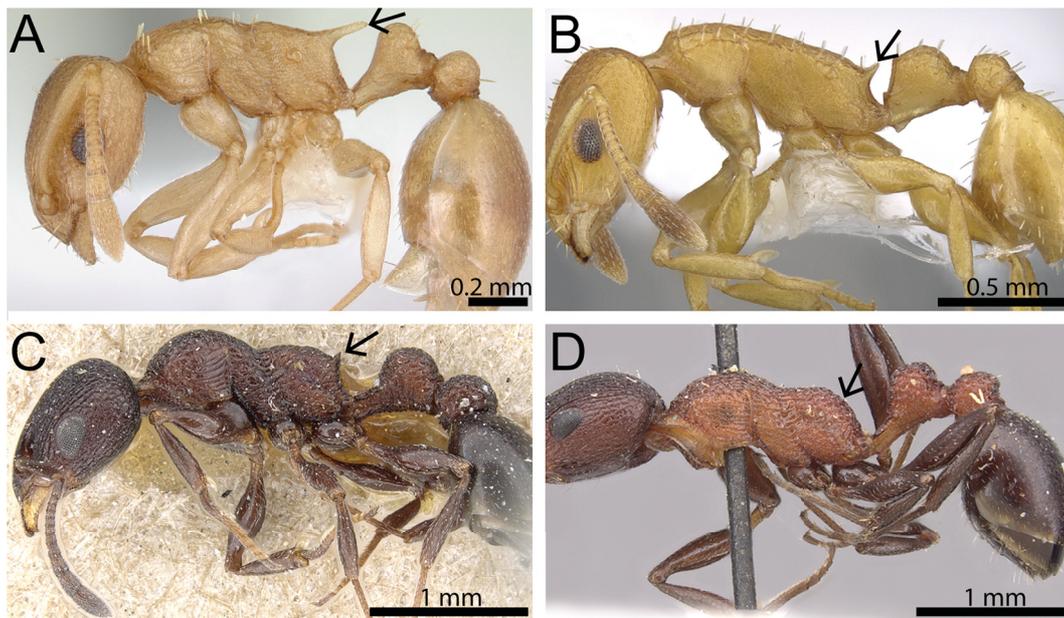


**Fig. 2.** Part of head in full-face view showing the antennae (antennal segments are counted in Roman numerals) and dorsum of mesosoma. **A–B.** *Nesomyrmex cataulacoides* (Snelling, 1992) (CASENT0172593, CASENT0178301). **C.** *N. humerosus* (Emery, 1896) (CASENT0906196). **D.** *N. evelynae* (Forel, 1916) (CASENT0178298). **E.** *N. stramineus* (Arnold, 1948) (CASENT0922011). **F.** *N. braunsi* (Forel, 1912) (CASENT0909209).

3. Propodeal spines present and conspicuous, usually long and narrow, in profile distinctly longer than their basal width (Fig. 4A–B); hairs on dorsum of mesosoma always present, short, and blunt (Fig. 4A–B) ..... *N. angulatus* group  
 – Propodeal spines usually absent (Fig. 4D), if present, then very short and broad, in profile no longer than their basal width (Fig. 4C); hairs on dorsum of mesosoma variably developed: usually long, fine and acute, sometimes absent, and only very rarely short and blunt .....  
 ..... *N. simoni* group



**Fig. 3.** Anterior cephalic dorsum (white ellipse around anterior clypeus) and body in profile. **A–B.** *N. humerosus* (Emery, 1896) (CASENT0906196). **C.** *N. angulatus* (Mayr, 1862) (CASENT0235552). **D.** *N. angulatus* (Mayr, 1862) (CASENT0922010).



**Fig. 4.** Body in profile. **A.** *Nesomyrmex evelynae* (Forel, 1916) (CASENT0178298). **B.** *N. angulatus* (Mayr, 1862) (CASENT0922010). **C.** *N. simoni* (Emery, 1895) (CASENT0904790). **D.** *N. braunsi* (Forel, 1912) (CASENT0909209).

***Nesomyrmex angulatus* species group**

**Definition**

Antennae with 12 segments; anterior clypeal lobe conspicuously convex, rounded and without a small median triangular projection; frontal carinae absent; propodeum always with moderately long to long spines, in profile distinctly longer than their basal width; petiole and postpetiole without large lateral spines, petiole sometimes with small denticles; all dorsal surfaces of body with short, blunt pilosity (with the exception of *N. evelynae*, which lacks standing hairs on most of first gastral tergite).

**Comments**

This group contains seven species that are widely distributed in the Afrotropical region, with one species extending into the Palaearctic and Malagasy regions. All species appear to be arboreal or subarboreal. Species accounts for all group members are provided below in the review of the group. The alpha taxonomy of the group appears straightforward based on the literature (Bolton 1982; Mbanyana & Robertson 2008). However, we find some species delimitations quite problematic (see species accounts below) and it might be necessary to revise the *N. angulatus* species group in the future after the accumulation of more material from additional Afrotropical localities.

***Nesomyrmex cataulacoides* species group**

**Definition**

Antennae with 11 segments; anterior clypeal lobe conspicuously convex and rounded, without a small median triangular projection; frontal carinae absent; propodeal spines very well developed, long and spiniform; petiole and postpetiole each with a pair of large and conspicuous lateral spines; all dorsal surfaces of body without standing pilosity.

**Comments**

The *N. cataulacoides* species group holds only one morphologically bizarre and extraordinary species, which is impossible to confuse with any other *Nesomyrmex* species from the Afrotropical or any other region. The 11-segmented antennae, lack of standing pilosity on all dorsal surfaces, and the extreme spinosity render *N. cataulacoides* immediately recognizable. The possession of spines/teeth on the anterior pronotal corners, the anterior and posterior propodeum, and on both waist segments is unique within the genus. The affinities of *N. cataulacoides* to other *Nesomyrmex* are difficult to ascertain, mostly due to the extreme morphological specializations. So far it is not possible to associate it closely with any other *Nesomyrmex* species or species group. Currently, the species is known only from Cameroon and Kenya. Based on the few collections available, this species is strictly arboreal. The observed disjunctive distribution is most likely a sampling artefact due to the scarcity of collecting in Central African canopies, and we expect that *N. cataulacoides* will be collected in intermediate countries in the future.

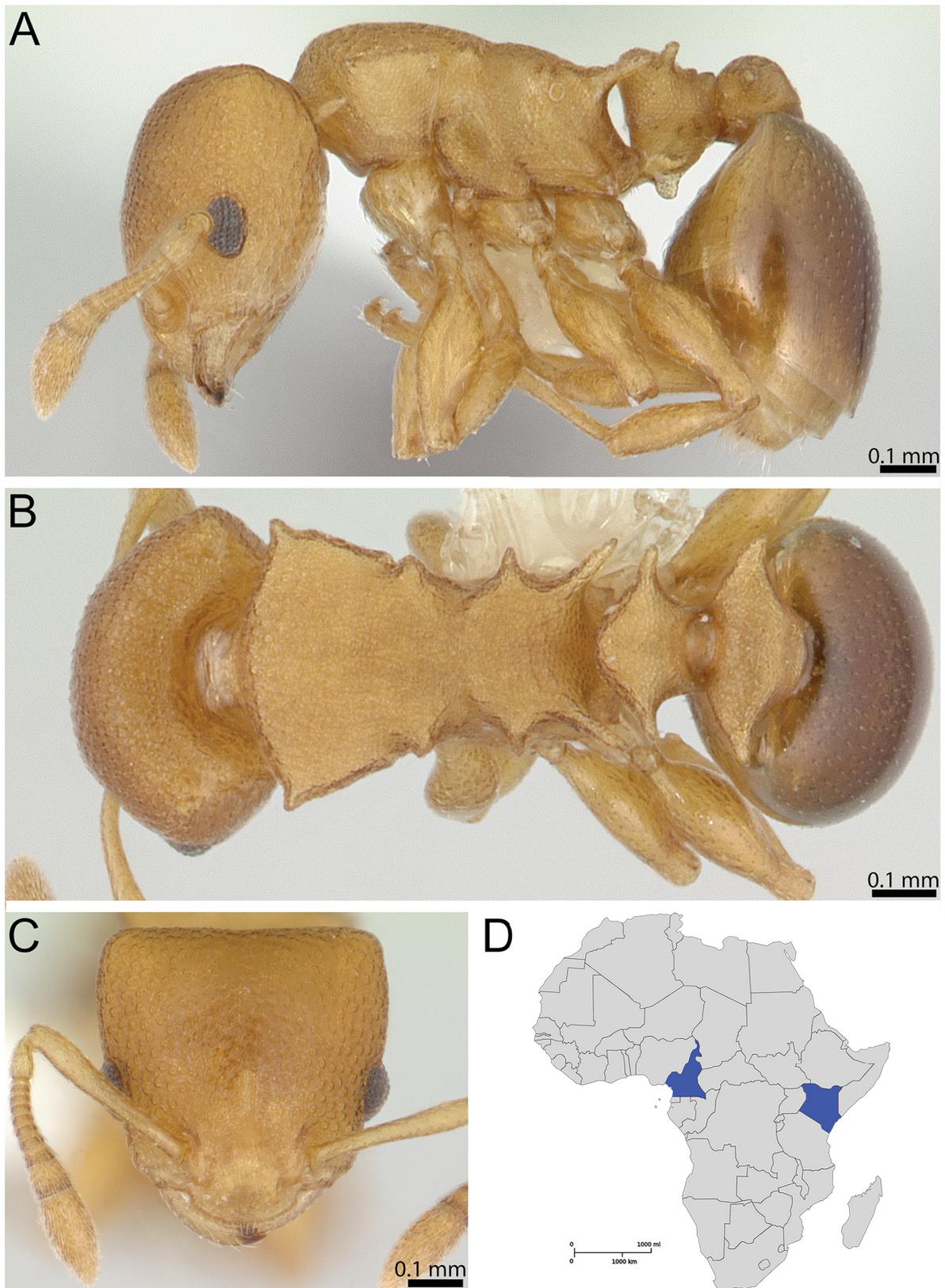
***Nesomyrmex humerosus* species group**

**Definition**

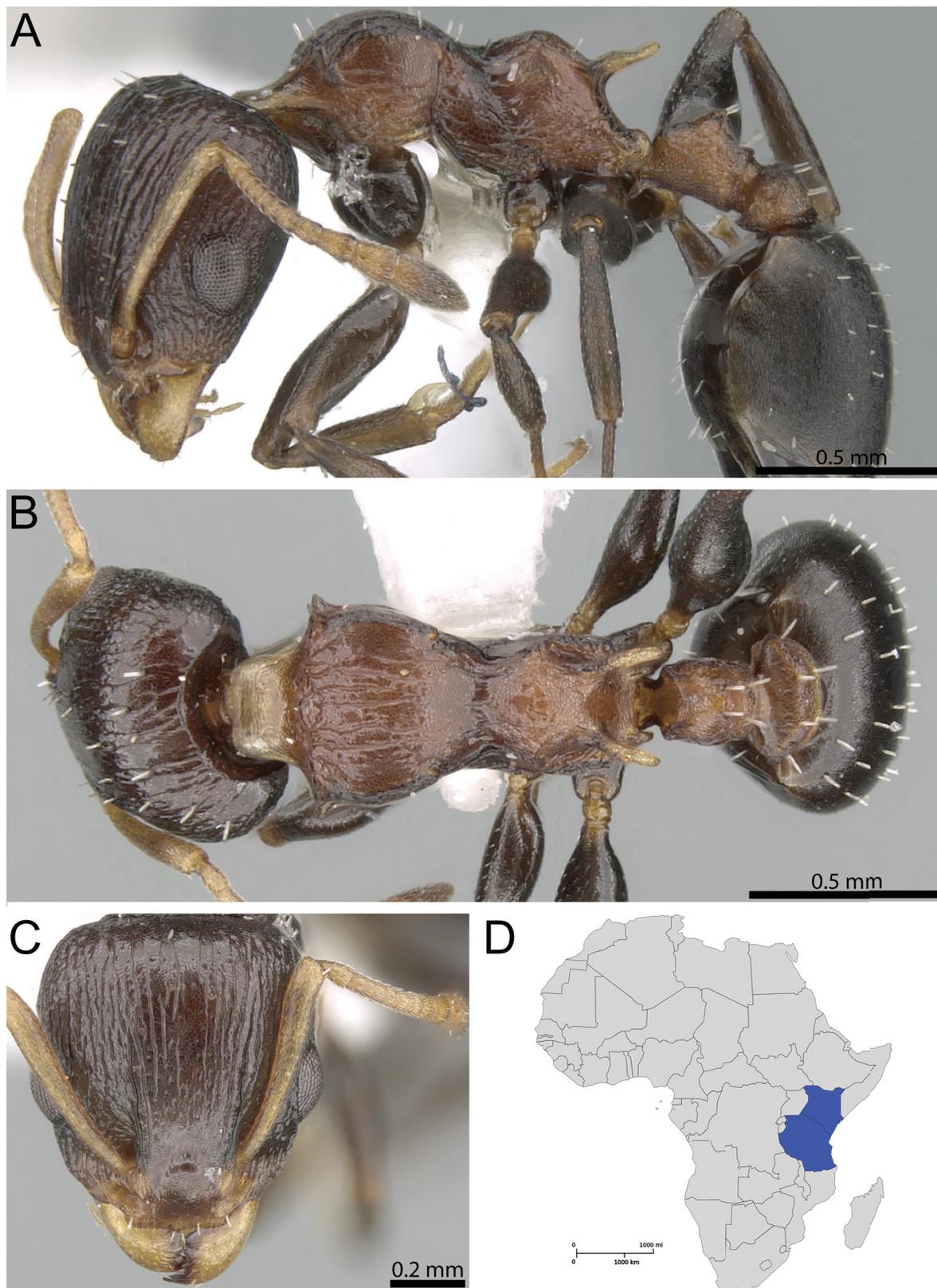
Antennae with 12 segments; anterior clypeal lobe short, flat-margined, and never convex, with small median triangular projection; frontal carinae present, but weakly developed; propodeal spines very well developed, long and spiniform; petiole and postpetiole without lateral spines; all dorsal surfaces of body with short, blunt pilosity.

**Comments**

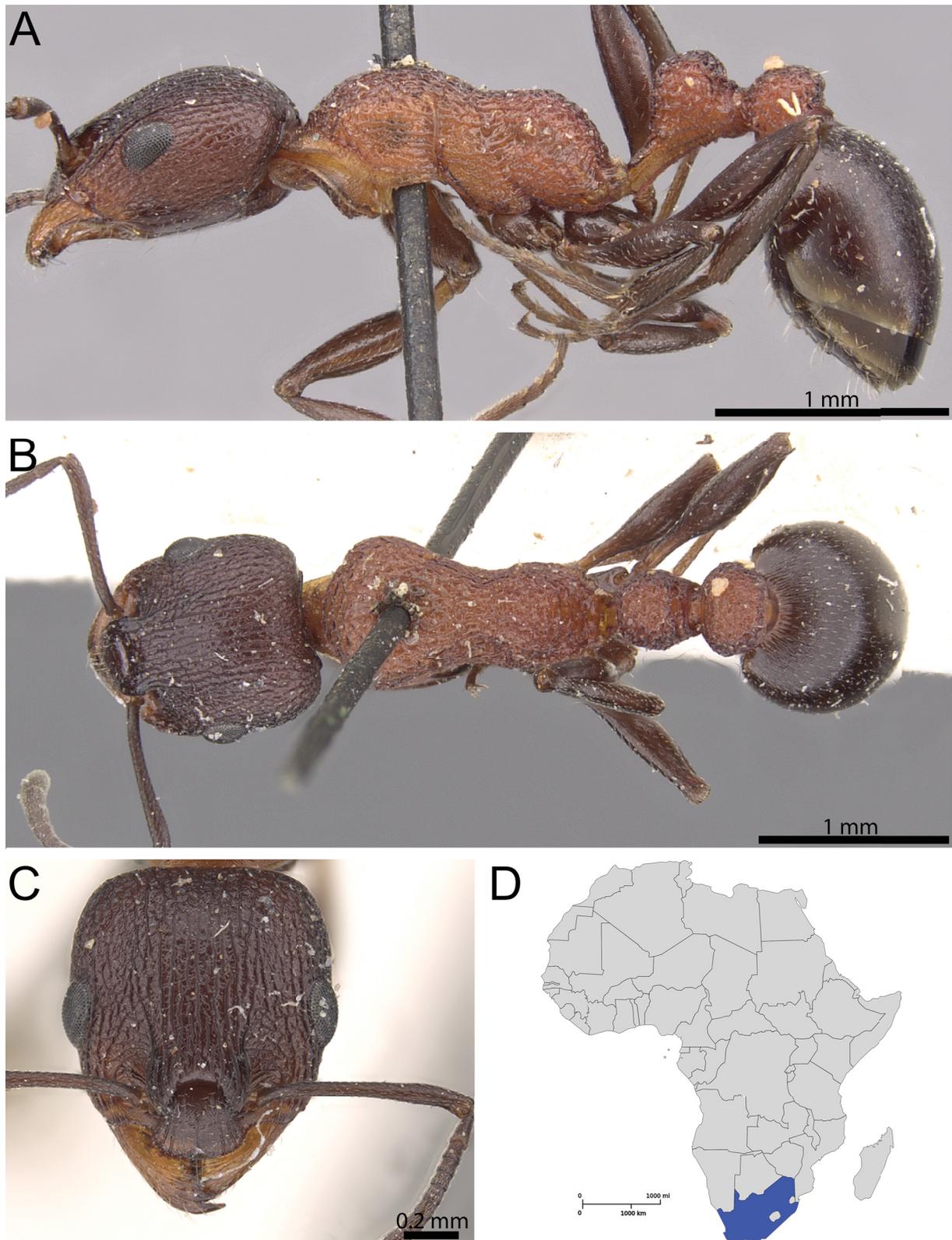
This group contains only one species, which is morphologically quite unique in the Afrotropical region. Of special importance are the short, flat-margined anterior clypeal lobe with a small median triangular projection and the barrel-shaped petiolar node with its small, triangular node. These characters are in slightly modified ways also seen in several Neotropical and Malagasy species, while they are absent in



**Fig. 5.** *Nesomyrmex cataulacoides* (Snelling, 1992) (CASENT0178300). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.



**Fig. 6.** *Nesomyrmex humerosus* (Emery, 1896) (CASENT0906196). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.



**Fig. 7.** *Nesomyrmex braunsi* (Forel, 1912) (CASENT0909209). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.

the other Afrotropical species groups. However, the fact that *N. humerosus* shares these characters with species from other regions does not necessarily mean that *N. humerosus* is more closely related to them. It could also be an independent African lineage and similarities with species from other regions might be based on convergence. Despite the fact that *N. humerosus* does not resemble most species from the *N. simoni* group, it still shares characters with some species, such as the large eyes and the shape of the dorsal mesosomal outline, and it could be that *N. humerosus* is just an aberrant *N. simoni* group member. At present, it is not possible to deduce the biogeographical and phylogenetic affinities of this peculiar species. Currently, *N. humerosus* is known to occur in Kenya, Tanzania and Yemen. It is a rather rarely collected species and our scarce knowledge is based on just four collection events. Based on a sample collected in Kenya by the first author, it seems to live on vegetation, but it was also sampled from the ground in Tanzania and Yemen. It is possible that the species also occurs in other East African countries, such as Somalia and Mozambique, which are greatly under-sampled.

#### *Nesomyrmex simoni* species group

##### Definition

Antennae with 12 segments; anterior clypeal lobe conspicuously convex and rounded, without a small median triangular projection; frontal carinae absent; propodeum usually unarmed, rarely with short teeth, in profile no longer than their basal width; petiole and postpetiole without lateral spines; usually all dorsal surfaces of body with (mostly) long, fine or (rarely) short, blunt standing pilosity, sometimes pilosity reduced on a few body parts, but never completely absent from all dorsal surfaces.

##### Comments

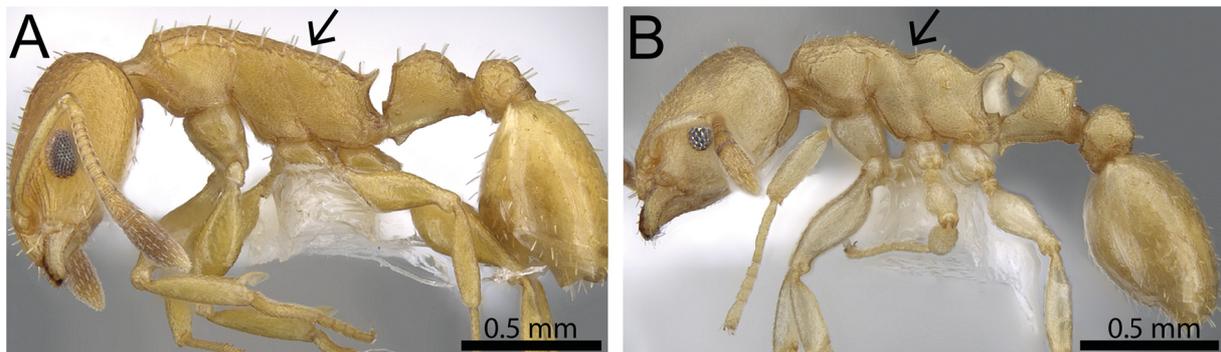
The 17 species of this group are all endemic to South Africa. In contrast to the members of most other Afrotropical groups, all *N. simoni* group species nest and live on the ground. Mbanyana & Robertson (2008) revised this group extensively, provided a sound and functional species identification key, and presented detailed descriptions of all species. Consequently, in this study we do not go into further details concerning the *N. simoni* group and refer to Mbanyana & Robertson (2008).

#### Review of the *Nesomyrmex angulatus* species group

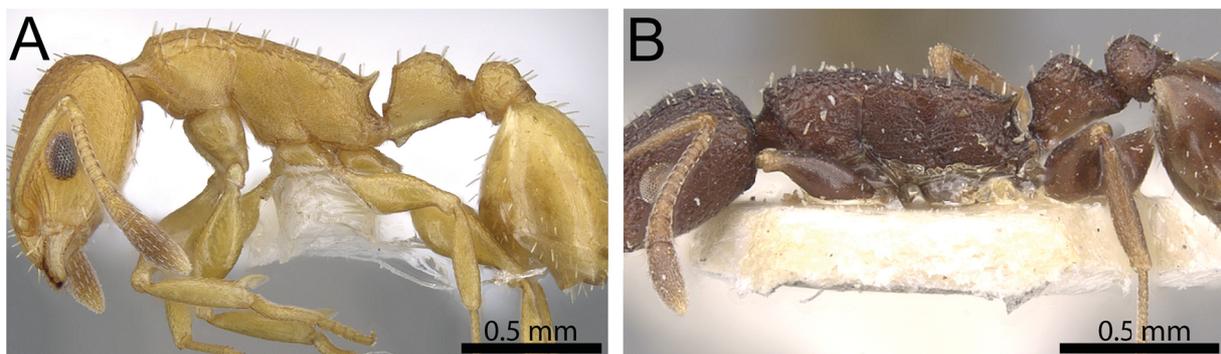
##### Identification key to Afrotropical species of the *N. angulatus* species group (workers)

The following key is partly based on Bolton (1982) and Mbanyana & Robertson (2008).

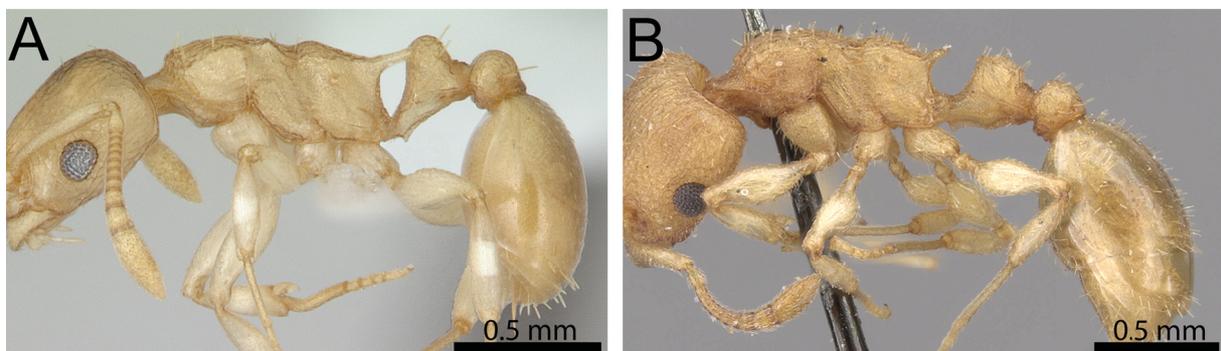
1. In profile mesosomal dorsum forming a single, uninterrupted flat surface without any trace of metanotal groove; petiolar peduncle short (Fig. 8A) .....2  
– In profile mesosomal dorsum always with conspicuously impressed metanotal groove; petiolar peduncle long (Fig. 8B) .....3
2. Body colour yellow to very light brown (Fig. 9A) .....*N. angulatus* (Mayr, 1862)  
– Body colour very dark brown to black (Fig. 9B) .....*N. grisoni* (Forel, 1916)
3. First gastral tergite lacking standing hairs except for single transverse row on posterior end of tergite (Fig. 10A) .....*N. evelynae* (Forel, 1916)  
– First gastral tergite with standing hairs evenly distributed throughout (Fig. 10B) .....4
4. Antennal scapes conspicuously longer (SI 95–98); in dorsal view sides of petiolar node straight to weakly rounded, not laterally denticulate (Fig. 11A); dorsum of propodeum without standing hairs (Fig. 11B) .....*N. inhaca* sp. nov.  
– Antennal scapes conspicuously shorter (SI 67–77); in dorsal view petiolar node laterally denticulate (Fig. 11C); dorsum of propodeum with short standing hairs (Fig. 11D) .....5



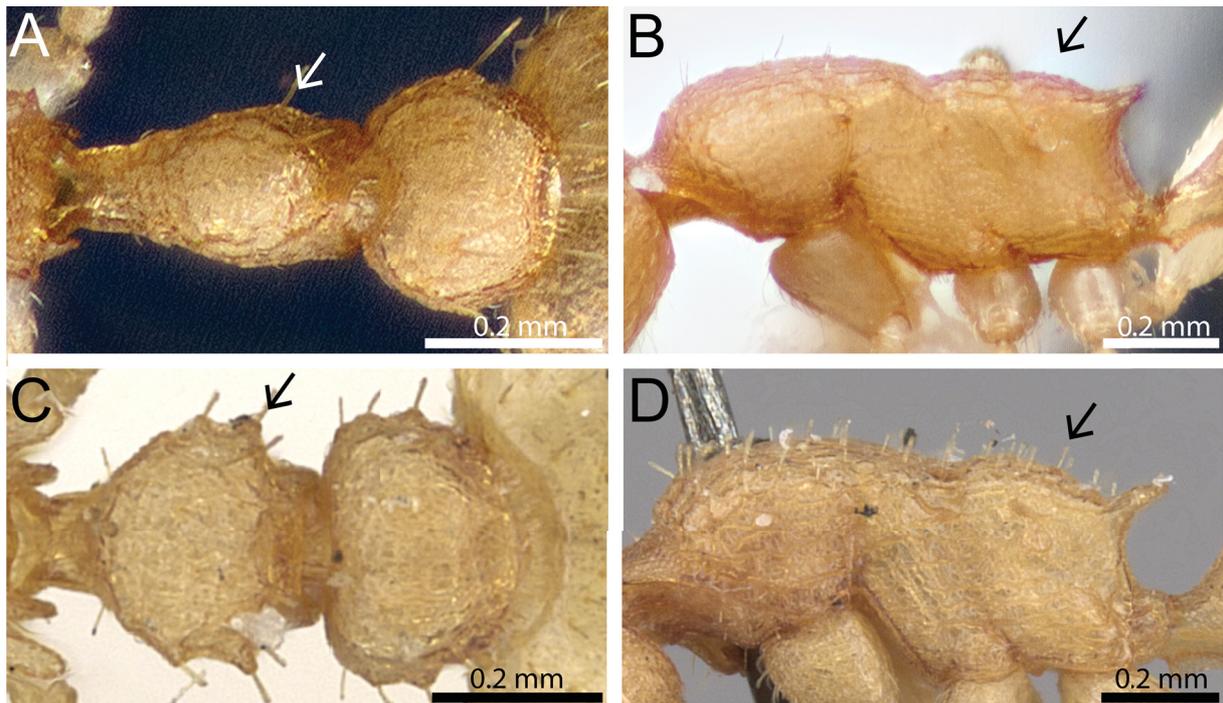
**Fig. 8.** Body in profile. **A.** *Nesomyrmex angulatus* (Mayr, 1862) (CASENT0922010). **B.** *N. stramineus* (Arnold, 1948) (CASENT0922011).



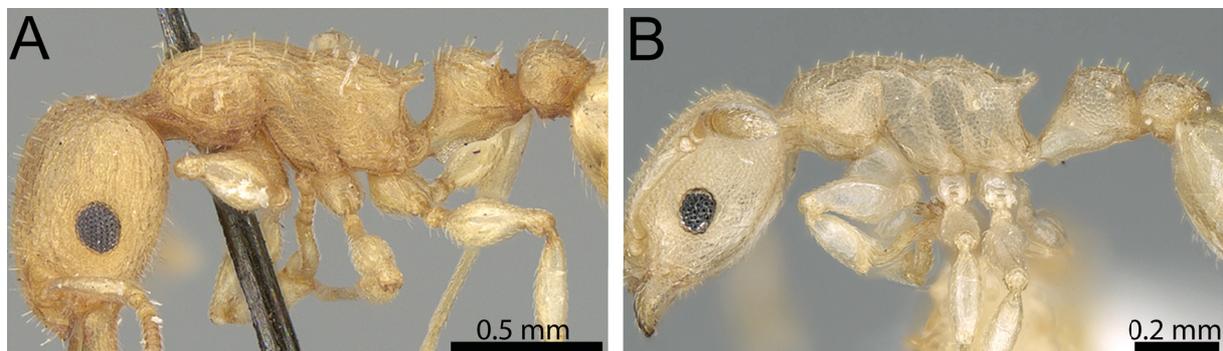
**Fig. 9.** Body in profile. **A.** *Nesomyrmex angulatus* (Mayr, 1862) (CASENT0922010). **B.** *N. grisoni* (Forel, 1916) (CASENT0908994).



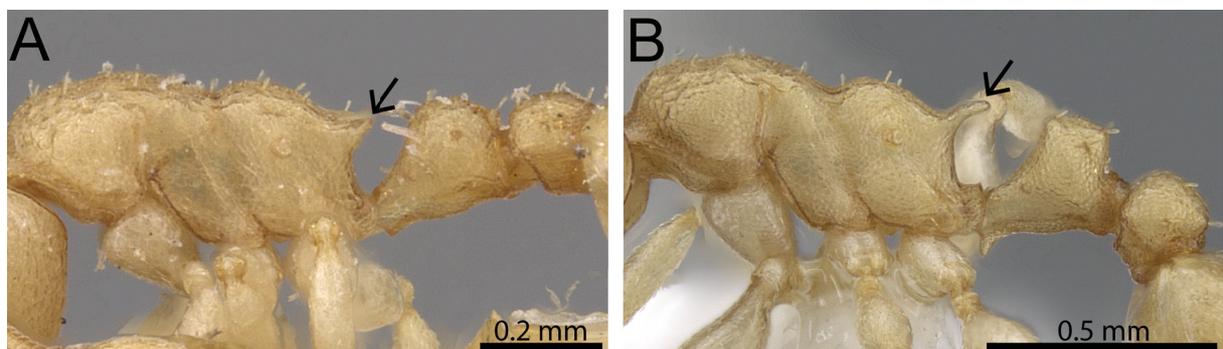
**Fig. 10.** Body in profile. **A.** *Nesomyrmex evelynae* (Forel, 1916) (CASENT0413068). **B.** *N. denticulatus* (Mayr, 1901) (CASENT0908992).



**Fig. 11.** Waist segments in dorsal view and mesosoma in profile. **A–B.** *Nesomyrmex inhaca* sp. nov. (CASENT0790016). **C.** *N. denticulatus* (Mayr, 1901) (CASENT0914923). **D.** *N. denticulatus* (Mayr, 1901) (CASENT0908992).



**Fig. 12.** Head, mesosoma, and waist segments in profile. **A.** *Nesomyrmex denticulatus* (Mayr, 1901) (CASENT0914923). **B.** *N. innocens* (Forel, 1913) (CASENT0906195).



**Fig. 13.** Mesosoma and waist segments in profile. **A.** *Nesomyrmex innocens* (Forel, 1913) (CASENT0908995). **B.** *N. stramineus* (Arnold, 1948) (CASENT0922011).

5. Eyes larger, with 10–12 ommatidia in longest row (Fig. 12A); subpetiolar process with a conspicuous tooth anteriorly, followed by a long cuticular flange running back to the postpetiolar junction (Fig. 12A) ..... *N. denticulatus* (Mayr, 1901)  
 – Eyes smaller, with 7–9 ommatidia in longest row (Fig. 12B); subpetiolar process with or without a conspicuously developed tooth anteriorly, but without a long cuticular flange running back to the postpetiolar junction (Fig. 12B) ..... 6
6. Propodeal spines shorter and thicker, elongate-triangular and only weakly longer than their basal width; in profile petiolar node nodiform, appearing approximately as long as high (Fig. 13A) ..... *N. innocens* (Forel, 1913)  
 – Propodeal spines longer and thinner, several times longer than their basal width; in profile petiolar node high, rectangular nodiform, appearing around twice as high as long (Fig. 13B) ..... *N. stramineus* (Arnold, 1948)

*Nesomyrmex angulatus* (Mayr 1862)

Figs 3C–D, 4B, 8A, 9A, 14

*Leptothorax angulatus* Mayr, 1862: 739 (w.), Egypt.

*Leptothorax angulatus* var. *concolor* Santschi, 1914: 107, fig. 15 (w.), Kenya.

*Leptothorax angulatus* r. *ilgii* Forel, 1894: 82 (w.), Ethiopia.

*Leptothorax latinodis* Mayr, 1895: 130 (w.), Mozambique.

*Leptothorax angulatus* – Santschi 1914: 107 (q.).

*Leptothorax (Goniothorax) angulatus* – Emery 1896: 58 (footnote).

*Nesomyrmex angulatus* – Bolton 2003: 272. See also: Mbanyana & Robertson 2008: 38. (Current subspecies: nominal plus *lybica*.)

*Leptothorax (Goniothorax) angulatus* r. *ilgii* – Santschi 1912: 148 (q.). — W.M. Wheeler 1922: 891. (Junior synonym of *angulatus*: Bolton 1982: 324.)

*Leptothorax (Goniothorax) latinodis* – Emery 1896: 58 (footnote). (Junior synonym of *angulatus*: Bolton 1982: 324.)

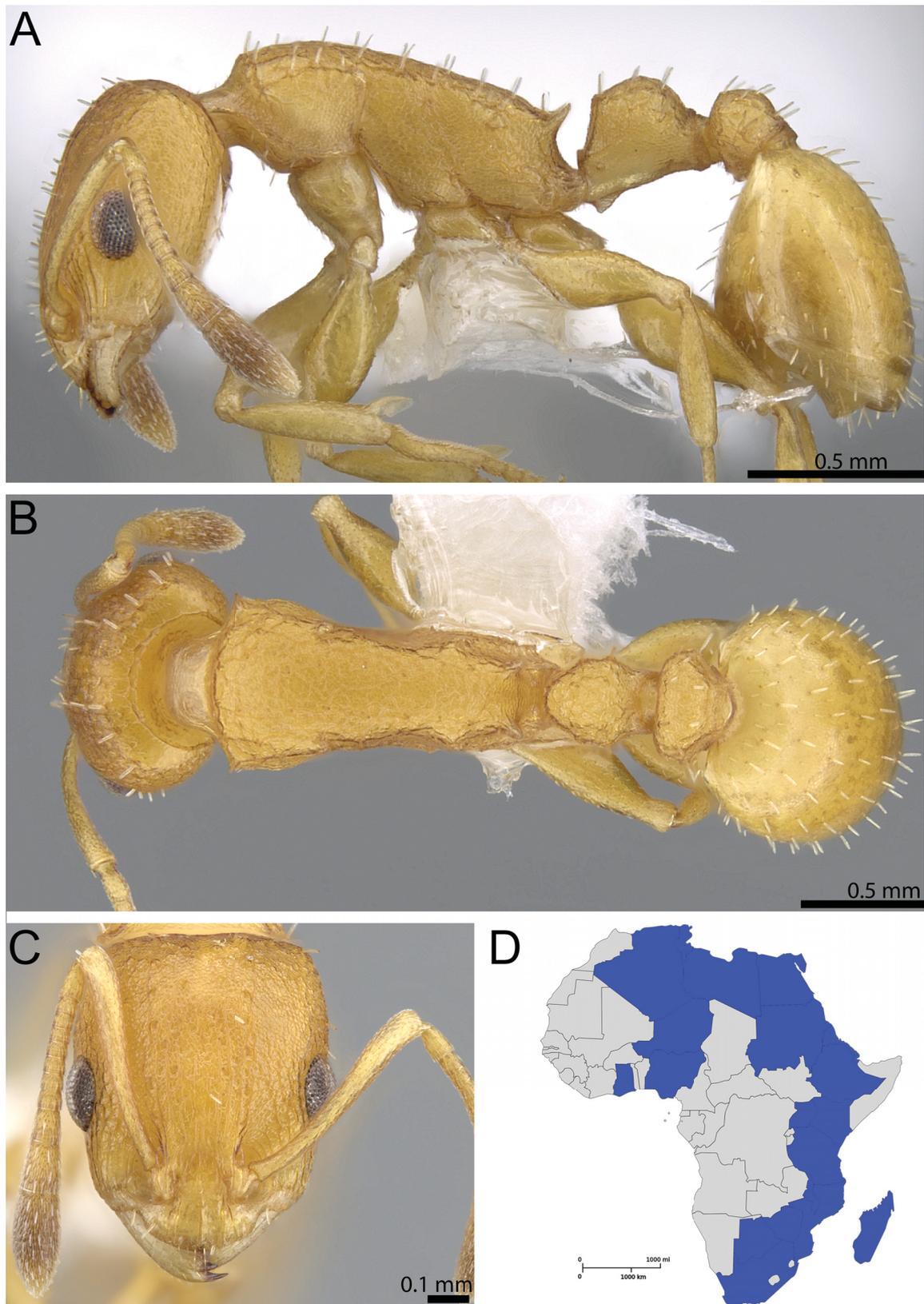
*Leptothorax angulatus* var. *concolor* – Emery 1915: 16 (q.m.). (Junior synonym of *angulatus*: Bolton 1982: 324.)

**Diagnosis**

The following character combination distinguishes *N. angulatus* from the other members of the group: in profile mesosomal dorsum forming a single, uninterrupted flat surface without any trace of metanotal groove; petiolar peduncle short; body colour yellow to very light brown.

**Diagnostic comments**

*Nesomyrmex angulatus* together with *N. grisoni* are easily separable from the other members of the group on the basis of the dorsal mesosomal outline, which is an uninterrupted, flat surface without any trace of a metanotal groove. The separation of *N. angulatus* from *N. grisoni* is less clear though. As Bolton (1982) stated in his revision, the only differentiating character is body colour, which is yellowish in *N. angulatus* and dark brown to black in *N. grisoni*. In general, body coloration is a rather weak diagnostic character and extremely variable in many ant species, and it is likely that both species are actually conspecific and the differently coloured forms represent geographic or ecological variants. The latter seems probable if one considers that *N. angulatus* is predominantly an arid-adapted species, while *N. grisoni* appears to prefer humid rain forests. Nevertheless, at the moment we hesitate to synonymise the two species and prefer to keep them separate for the following reasons. First, while there is a lot of material of *N. angulatus* in many museums, there is not much of *N. grisoni*, making comparative



**Fig. 14.** *Nesomyrmex angulatus* (Mayr, 1862) (CASENT0922010). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.

analyses challenging. Secondly, and more importantly, we are not fully convinced that all the material currently listed and identified as *N. angulatus* represents the same species. On the basis of some recent collections from Kenya and Mozambique we were able to observe a lot of morphological variation within and between localities. As already noted by Bolton (1982), the shape of the petiolar node seems to be especially variable. Consequently, we cannot rule out the possibility of dealing with a complex of more or less cryptic species. At the same time it is possible that *N. angulatus* is not only a very successful and widespread, but also an extremely variable species. The solution to this problem is not the aim of this study, since it requires the accumulation of an extensive amount of material from all over Africa, the Arabian Peninsula, and the Malagasy region.

### **Biology**

*Nesomyrmex angulatus* was collected from a variety of habitat types, such as tropical dry forest, coastal scrub, mangrove forest, savannah, and Acacia woodland. In general it seems as if the species prefers comparatively arid environments. In addition, it is predominantly found on the trunk of trees or the lower vegetation, rarely on the ground, and it nests in pre-existing cavities of dead wood (Bolton 1982; Mbanyana & Robertson 2008).

### **Distribution**

This species has by far the widest distribution range within the *N. angulatus* group, and likely represents the most widespread *Nesomyrmex* species on a global scale. It is found in the majority of African countries, as well as on the Arabian Peninsula and in most of the Malagasy region.

### *Nesomyrmex denticulatus* (Mayr, 1901) Figs 10B, 11C–D, 12A, 15

*Leptothorax denticulatus* Mayr, 1901: 5 (w.q.), South Africa.

*Leptothorax (Goniothorax) denticulatus* – W.M. Wheeler 1922: 891.

*Nesomyrmex denticulatus* – Bolton 2003: 272 (see also Bolton 1982: 328; Mbanyana & Robertson 2008: 38).

### **Diagnosis**

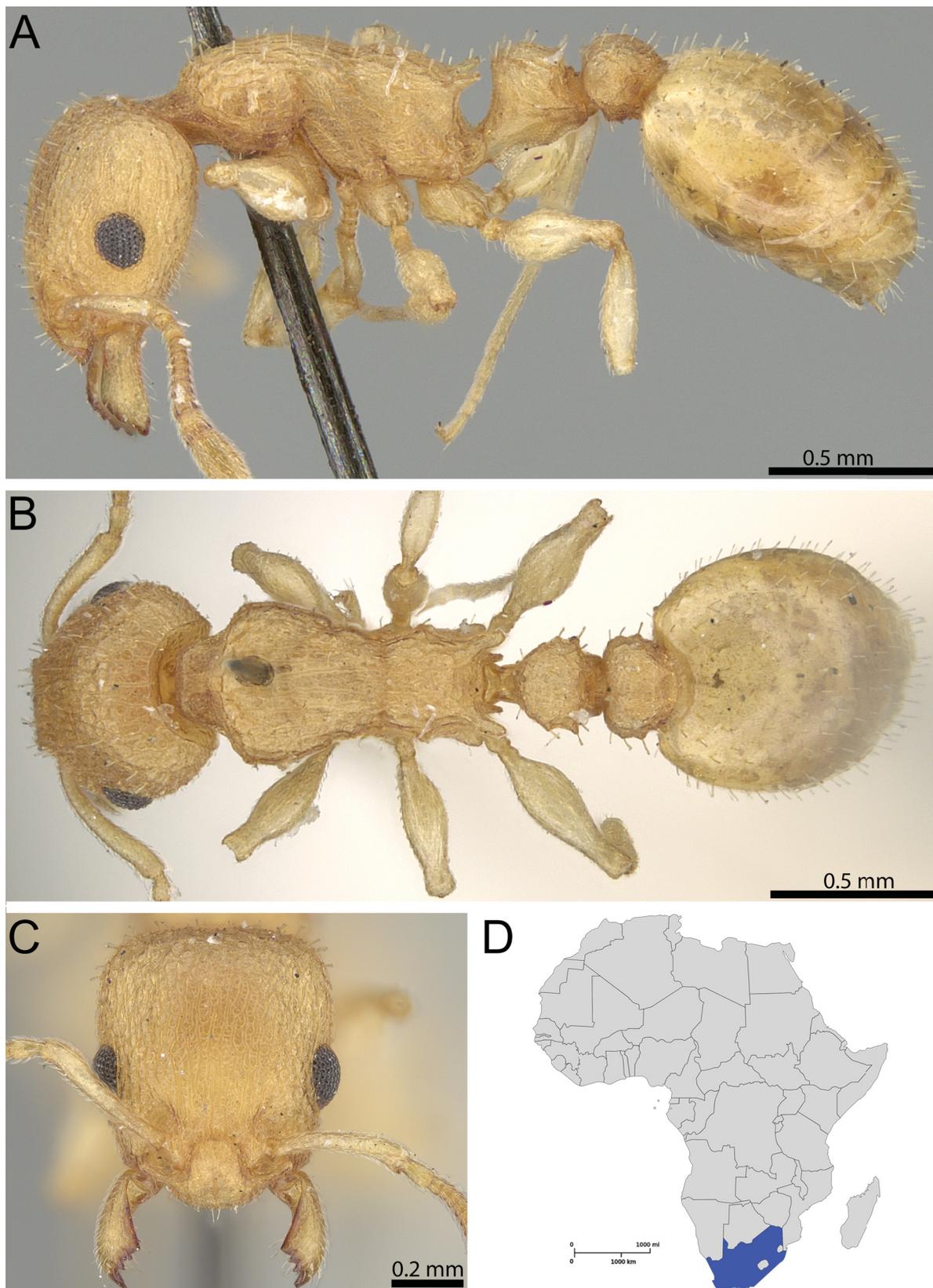
The following character combination separates *N. denticulatus* from the remainder of the group: eyes with 10–12 ommatidia in longest row; in profile mesosomal dorsum with conspicuously impressed metanotal groove; in dorsal view petiolar node laterally denticulate; subpetiolar process with a conspicuous tooth anteriorly followed by a long cuticular flange which runs back to the postpetiolar junction; dorsum of propodeum with standing hairs; first gastral tergite with standing hairs evenly distributed throughout.

### **Diagnostic comments**

The three species, *N. denticulatus*, *N. innocens* and *N. stramineus*, are morphologically very similar and can be well separated from the other species by the laterally denticulate petiolar node. The separation of these three can be challenging though. *Nesomyrmex denticulatus* is larger in general body size, has larger eyes with more ommatidia, and a subpetiolar process with a conspicuous tooth anteriorly, followed by a long cuticular flange which runs back to the postpetiolar junction, and slightly denser pilosity.

### **Biology**

This species usually nests in cavities of branches on trees and bushes previously excavated by wood-boring beetles, lepidopteran larvae or termites (Mbanyana & Robertson 2008). It is found in a variety



**Fig. 15.** *Nesomyrmex denticulatus* (Mayr, 1901) (CASENT0914923). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.

of more arid habitats, such as late succession Fynbos, Succulent Karoo with large bushes, along edges of Southern Afrotropical Forest, and possibly also Albany Thicket (Mbanyana & Robertson 2008).

### **Distribution**

*Nesomyrmex denticulatus* is only known from South Africa, where it seems to be relatively common in the Western and Eastern Cape regions.

*Nesomyrmex evelynae* (Forel, 1916)  
Figs 2D, 4A, 10A, 16

*Leptothorax (Goniothorax) evelynae* Forel, 1916: 423 (w.q.), D.R. Congo.

*Nesomyrmex evelynae* – Bolton 2003: 272 (see also Bolton 1982: 328).

### **Diagnosis**

The following character combination distinguishes *N. evelynae* from the other species of the group: in profile mesosomal dorsum with conspicuously impressed metanotal groove; petiolar peduncle long; dorsum of propodeum without standing hairs; first gastral tergite lacking standing hairs except for single transverse row on posterior end of tergite.

### **Diagnostic comments**

The recognition of this species within the *N. angulatus* group is fairly straightforward, since it is the only one that lacks standing hairs on most of the first gastral tergite while all other group species have standing hairs evenly distributed throughout this tergite. It also lacks standing hairs on the propodeum, a character shared only with *N. inhaca* sp. nov., whereas the other five species have short, standing pilosity on the propodeum.

### **Biology**

It prefers rain forests where it lives in the canopy stratum. Based on canopy fogging samples from Kenya available to the first author, it appears that this species is found commonly on trees, even though in small individual numbers, suggesting smaller colony sizes.

### **Distribution**

*Nesomyrmex evelynae* is found in Equatorial Africa ranging from Burkina Faso and Ghana in the west through the Central African Republic and the D.R. Congo to Uganda and Kenya in the east. The known distribution is disjunctive since *N. evelynae* is not known from the countries between Ghana and the Central African Republic and the D.R. Congo. We consider this more of a sampling artefact though, and expect the species to be collected from the countries in between in future sampling projects.

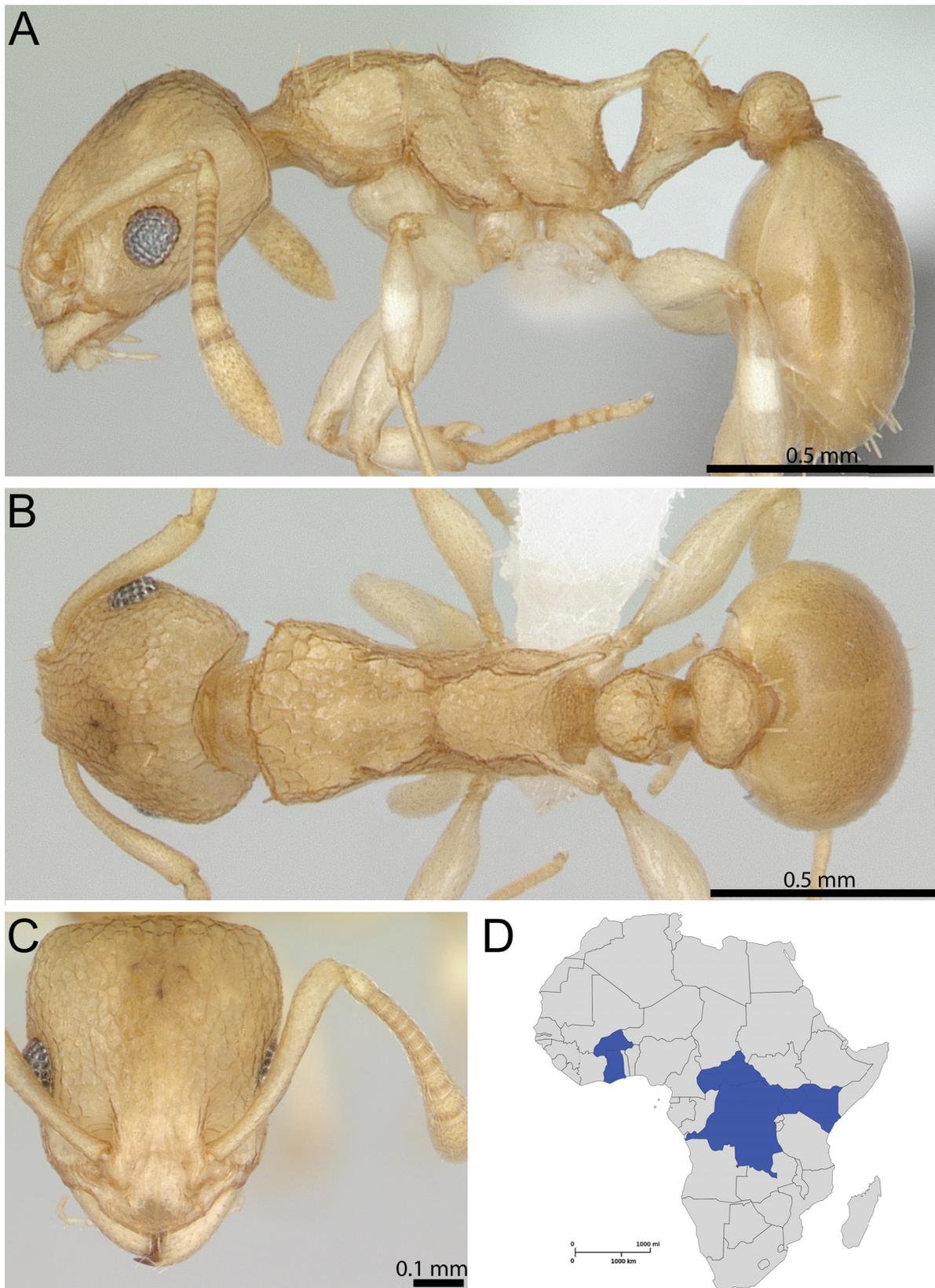
*Nesomyrmex grisoni* (Forel, 1916)  
Figs 9B, 17

*Leptothorax (Goniothorax) grisoni* Forel, 1916: 425 (w.q.). D.R. Congo.

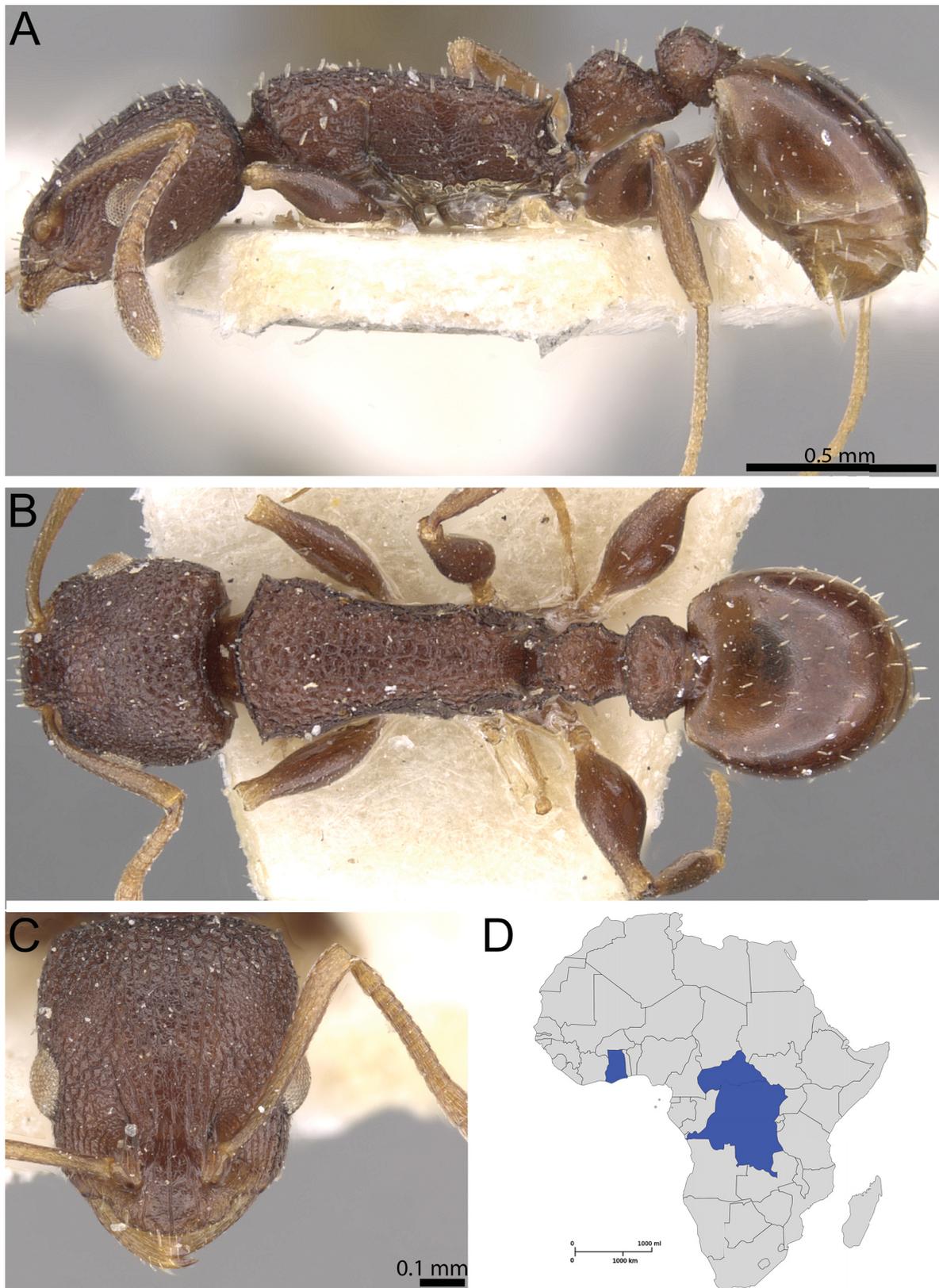
*Nesomyrmex grisoni* – Bolton 2003: 272 (see also Bolton 1982: 329).

### **Diagnosis**

The following character combination distinguishes *N. grisoni* from the other species of the group: in profile mesosomal dorsum forming a single, uninterrupted flat surface without any trace of metanotal groove; petiolar peduncle short; body colour dark brown to black.



**Fig. 16.** *Nesomyrmex evelynae* (Forel, 1916) (CASENT0413068). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.



**Fig. 17.** *Nesomyrmex grisoni* (Forel, 1916) (CASENT0908994). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.

### Diagnostic comments

As mentioned above, *N. grisoni* and *N. angulatus* are straightforwardly distinguishable from the remainder of the group. At the same time they are morphologically very close to each other and only separable on the basis of body colour. For a more in-depth discussion we refer to the species account of *N. angulatus*.

### Biology

Very little information about the natural history of this species is available. It seems to live on vegetation in rain forest habitats.

### Distribution

*Nesomyrmex grisoni* is only known from the Central African Republic, the D.R. Congo and Ghana.

*Nesomyrmex inhaca* sp. nov.

[urn:lsid:zoobank.org:act:C33B5A40-C3C0-4860-BD38-C99AC2D774A3](https://doi.org/10.2307/2393133)

Figs 11A–B, 18

### Diagnosis

The following character combination distinguishes *N. inhaca* sp. nov. from the other members of the *N. angulatus* group: in profile mesosomal dorsum with conspicuously impressed metanotal groove; in dorsal view petiolar node not laterally denticulate; dorsum of propodeum without standing hairs; first gastral tergite with standing hairs evenly distributed throughout.

### Etymology

The new species is named after the type locality, Inhaca Island, to the southeast of Mozambique. The species epithet is a noun in apposition and thus invariant.

### Type material

#### Holotype

MOZAMBIQUE: pinned worker, Maputo, Inhaca Island, 26°2'9" S, 32°54'17" E, 1 m, secondary forest, low vegetation, 21 Jun. 1992, G.D. Alpert leg. (MCZC: MCZ-ENT00593557).

#### Paratypes

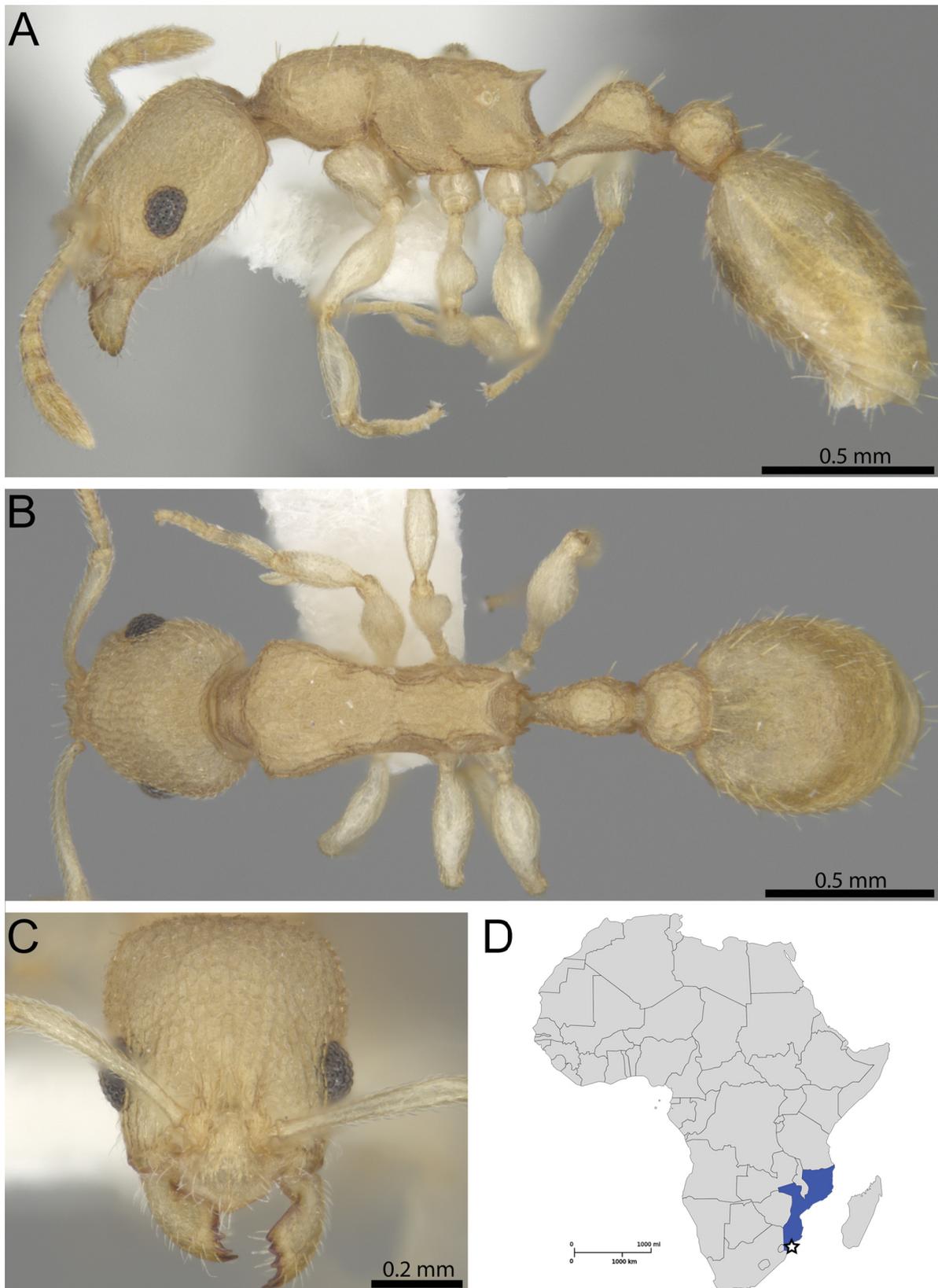
MOZAMBIQUE: 6 pinned workers, same collection data as holotype (BMNH: CASENT0790018; HLMD: HLMD-Hym-2395; KSMA: CASENT0790019; MCZC: CASENT0790017; SAMC: CASENT0790022; ZFMK: CASENT0790020).

### Worker measurements (n=7)

HL 0.69–0.73 (0.71); HW 0.50–0.53 (0.51); SL 0.47–0.51 (0.50); EL 0.16–0.17 (0.17); PH 0.26–0.28 (0.27); PW 0.41–0.42 (0.41); WL 0.85–0.88 (0.87); PSL 0.12–0.13 (0.12); PTL 0.14–0.16 (0.15); PTH 0.21–0.22 (0.22); PTW 0.19–0.20 (0.19); PPL 0.18–0.20 (0.19); PPH 0.21–0.22 (0.21); PPW 0.27–0.28 (0.28); CI 71–73 (72); SI 95–98 (96); OI 30–33 (32); DMI 47–49 (48); LMI 31–32 (32); PSLI 16–18 (17); LPeI 64–70 (69); DPeI 127–141 (131); LPpI 86–93 (89); DPpI 140–147 (145); PPI 138–147 (143).

### Worker description

HEAD. Masticatory margin of mandible with five teeth, decreasing in size from largest, acute apical tooth to smallest basal denticle; clypeus arched-convex to almost triangular, anterior margin with slightly darker, lamellate, flattened ridge all-around; head in full-face view appearing relatively narrow, much



**Fig. 18.** *Nesomyrmex inhaca* sp. nov., holotype (MCZ-ENT00593557). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution and type locality (black and white star symbol).

longer than broad (CI 71–73), sides of head approximately straight, gently broadening behind eye level, narrowest directly behind posterior eye margin, and widest halfway between posterior eye margin and posterior head margin; posterior head margin concave medially; frontal carinae and antennal scrobes absent; antennal scapes moderately long, not reaching posterior head margin (SI 95–98). Eyes relatively large (OI 29–33), with eight to nine ommatidia in the longest row.

**MESOSOMA.** In lateral view mesosomal outline relatively low (LMI 31–32) and flat with conspicuously impressed metanotal groove; promesonotal suture present laterally and completely absent dorsally; pronotum moderately marginate between lateral and dorsal mesosoma, anterodorsal corners not denticulate; propodeum armed with short to moderately long propodeal spines (PSLI 16–18), in profile spines distinctly longer than their basal width; propodeal lobes low and rounded.

**WAIST SEGMENTS AND GASTER.** Petiolar peduncle long, anteriorly with a small tooth-like subpetiolar process; in profile petiolar node relatively low and globular, between 1.4 and 1.5 times as high as long (LPeI 64–70); anterior face smoothly merging with peduncle and petiolar dorsum without any angles, posterior face slightly better developed; node in dorsal view about 1.3 to 1.4 times as wide as long (DPeI 127–141); in dorsal view petiolar node not laterally denticulate; in profile postpetiole globular, about 1.1 to 1.2 times as high as long (LPpI 86–93); in dorsal view about 1.4 and 1.5 times as wide as long (DPpI 140–147); postpetiole in dorsal view around 1.4 to 1.5 times as wide as petiolar node (PPI 138–147).

**SCULPTURE.** Mandibles shagreened to partly smooth and shiny, sometimes with very weak, superficial, irregular, longitudinal rugulae; median clypeal carina present and conspicuous, usually accompanied by one or two lateral, longitudinal, and slightly weaker rugae on each side; cephalic dorsum posteriorly and laterally strongly reticulate-rugose, medially more irregularly longitudinally rugose, ground sculpture conspicuously reticulate-punctulate; mesosoma laterally and dorsally with distinct reticulate-punctulate ground sculpture, lateral mesosoma conspicuously reticulate-rugose, dorsum reticulate-rugose with some irregular, longitudinal elements medially; legs unsculptured, smooth and shining; petiole and postpetiole with irregular reticulate-rugose sculpture superimposed on reticulate-punctulate ground sculpture; sculpture of first gastral tergite variable, some specimens only with reticulate-punctulate ground sculpture, other specimens irregularly rugose/rugulose on top of reticulate-punctulate ground sculpture.

**PILOSITY AND PUBESCENCE.** Head, mesosoma, waist segments and gaster dorsally with sparse, erect, blunt, and moderately long pilosity, hairs shorter on head and mesosoma than on waist segments and gaster; head laterally and ventrally with short appressed to decumbent pubescence; pubescence on mesosoma and waist segments sparse to absent; first gastral tergite with short to moderately long, appressed to decumbent pubescence.

**COLORATION.** Body uniformly yellowish to light brown, in a few specimens legs slightly lighter yellow, almost white.

### **Diagnostic comments**

Within the members of the *N. angulatus* species group, *N. inhaca* sp. nov. cannot be mistaken for *N. evelynae* since the latter is devoid of standing hairs on the first gastral tergite except for a single transverse row on the posterior end of the tergite, and also has very long propodeal spines and a relatively high petiolar node. *Nesomyrmex inhaca* sp. nov. has short, standing hairs evenly distributed throughout the first gastral tergite, shorter propodeal spines and a much lower petiolar node. The species pair *N. angulatus* and *N. grisoni* both do not possess a trace of a metanotal groove, which is obviously present in *N. inhaca* sp. nov. and thus it is not likely these species will be confused. Nevertheless, *N. inhaca* sp. nov. is morphologically closer to the trio of species *N. denticulatus*, *N. innocens* and *N. stramineus*. These

three are, however, easily separable since they possess a petiolar node with distinct lateral denticles and have standing hairs on the propodeal dorsum, while *N. inhaca* sp. nov. lacks both the lateral petiolar denticles and the standing hairs on the propodeum. Moreover, *N. inhaca* sp. nov. has apparently longer antennal scapes based on the much higher SI (94–95) compared to the other three species (SI 67–77). This is partly due to the fact that *N. inhaca* sp. nov. has indeed relatively longer antennal scapes, but the comparatively narrow head with low values of HW contributes to these high SI values. Therefore, we suggest being cautious with scape length as the single diagnostic character.

### Intraspecific variation

Since the description is based on just one collection event, the observed intraspecific variation seen in the worker caste is very low. However, the sculpture on the first gastral tergite shows some variability, as described above.

### Biology

*Nesomyrmex inhaca* sp. nov. was sampled from low vegetation in secondary forest at an elevation of 1 m. Apart from this, nothing is known of its natural history.

### Distribution

The new species is so far only known from one collection event on Inhaca Island in the southeast of Mozambique. Despite this apparently restricted distribution to just one island, we are reluctant to declare *N. inhaca* sp. nov. as endemic to Inhaca Island. With the noticeable exception of the area around Gorongosa, most of Mozambique remains severely under-sampled and our knowledge of local ant communities and species distributions is very poor to non-existent. Consequently, it is possible that *N. inhaca* sp. nov. will also be found on the mainland.

### *Nesomyrmex innocens* (Forel, 1913)

Figs 12B, 13A, 19

*Tetramorium (Leptothorax) innocens* Forel, 1913: 317 (w.), D.R. Congo.

*Leptothorax innocens* – Forel 1916: 425.

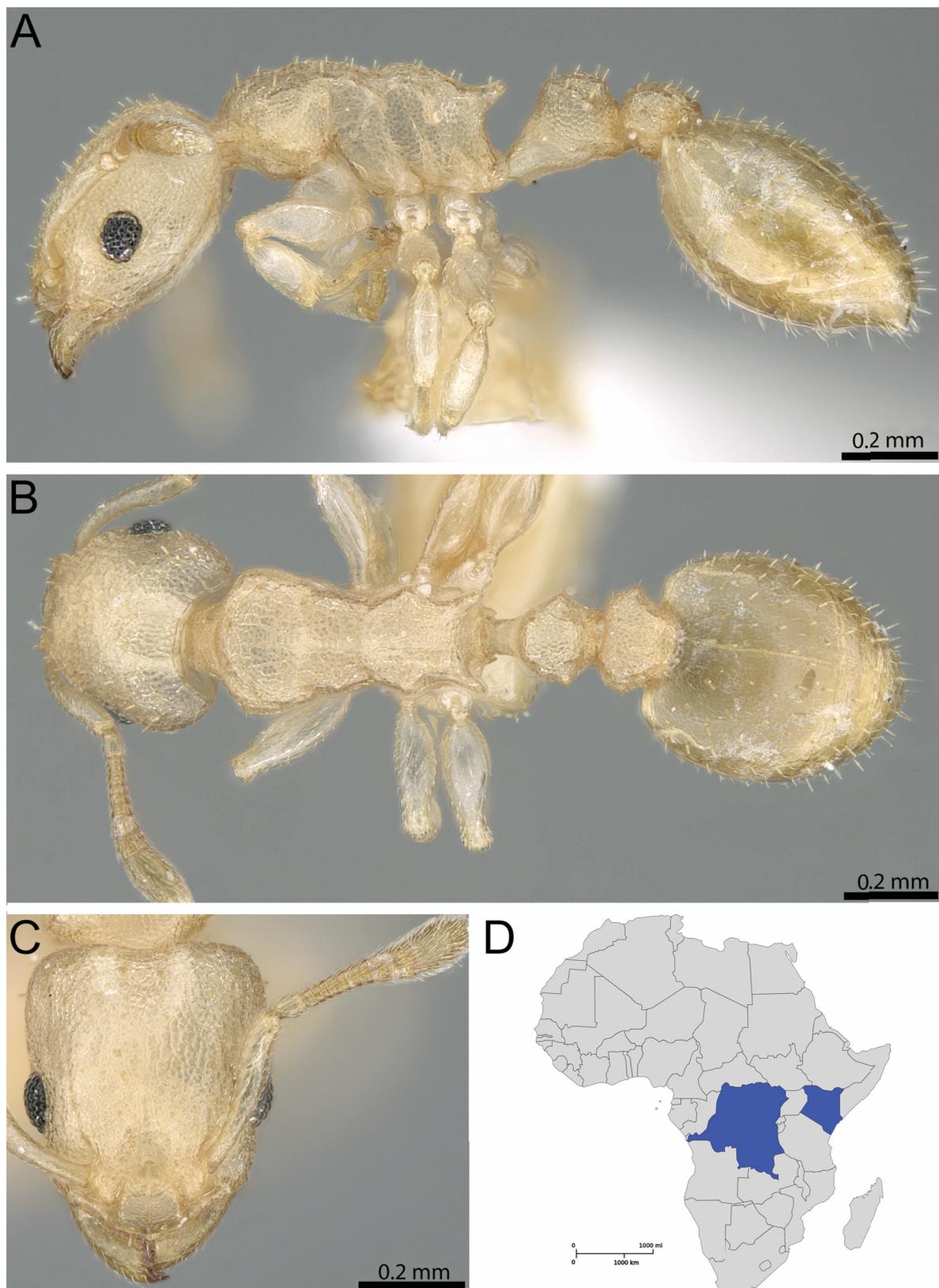
*Nesomyrmex innocens* – Bolton 2003: 272 (see also Bolton 1982: 330).

### Diagnosis

The following character combination distinguishes *N. innocens* from the other members of the group: eyes with 7–9 ommatidia in longest row; in profile mesosomal dorsum with conspicuously impressed metanotal groove; propodeal spines short and thick, elongate-triangular and only weakly longer than their basal width; in profile petiolar node nodiform, appearing approximately as long as high; in dorsal view petiolar node laterally denticulate; subpetiolar process without a long cuticular flange running back to the postpetiolar junction; dorsum of propodeum with standing hairs; first gastral tergite with standing hairs evenly distributed throughout.

### Diagnostic comments

As noted in the description of *N. denticulatus*, the latter, *N. innocens* and *N. stramineus* are morphologically relatively close. *Nesomyrmex innocens* and *N. stramineus* differ from *N. denticulatus* by generally smaller body size, smaller eyes with less ommatidia, and a subpetiolar process without a long cuticular flange running back to the postpetiolar junction. The separation of *N. innocens* and *N. stramineus* is a bit more difficult, as already mentioned by Bolton (1982). *Nesomyrmex innocens* has shorter and thicker propodeal spines and a lower and thicker petiolar node compared to *N. stramineus*. It is not clear at the



**Fig. 19.** *Nesomyrmex innocens* (Forel, 1913) (CASENT0906195). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.

moment whether or not these character states are sufficient to maintain their heterospecificity in the long term. Bolton (1982) had some doubts about this, too, and it is possible that they represent geographical varieties of the same species. However, at present, based on the scarcity of the material, especially of *N. innocens*, we treat them as two different species.

### **Biology**

Based on the limited data available, *N. innocens* nests in the stem of trees.

### **Distribution and biology**

This species is only known from very few specimens, collected from the D.R. Congo and Kenya.

#### *Nesomyrmex stramineus* (Arnold, 1948)

Figs 2D, 8B, 13B, 20

*Limnomyrmex stramineus* Arnold, 1948: 223 (w.), South Africa.

*Leptothorax stramineus* – Brown 1971: 4.

*Nesomyrmex stramineus* – Bolton 2003: 272 (see also Bolton 1982: 332; Mbanyana & Robertson 2008: 38).

### **Diagnosis**

The following character combination distinguishes *N. stramineus* from the other species of the group: eyes with 7–9 ommatidia in longest row; in profile mesosomal dorsum with conspicuously impressed metanotal groove; propodeal spines relatively long and thin, several times longer than their basal width; in profile petiolar node high, rectangular nodiform, appearing around twice as high as long; in dorsal view petiolar node laterally denticulate; subpetiolar process without a long cuticular flange running back to the postpetiolar junction; dorsum of propodeum with standing hairs; first gastral tergite with standing hairs evenly distributed throughout.

### **Diagnostic comments**

As pointed out above, *N. stramineus* is very close to *N. denticulatus* and *N. innocens*. For more details on its differentiation from them we refer to the species account of *N. innocens*.

### **Biology**

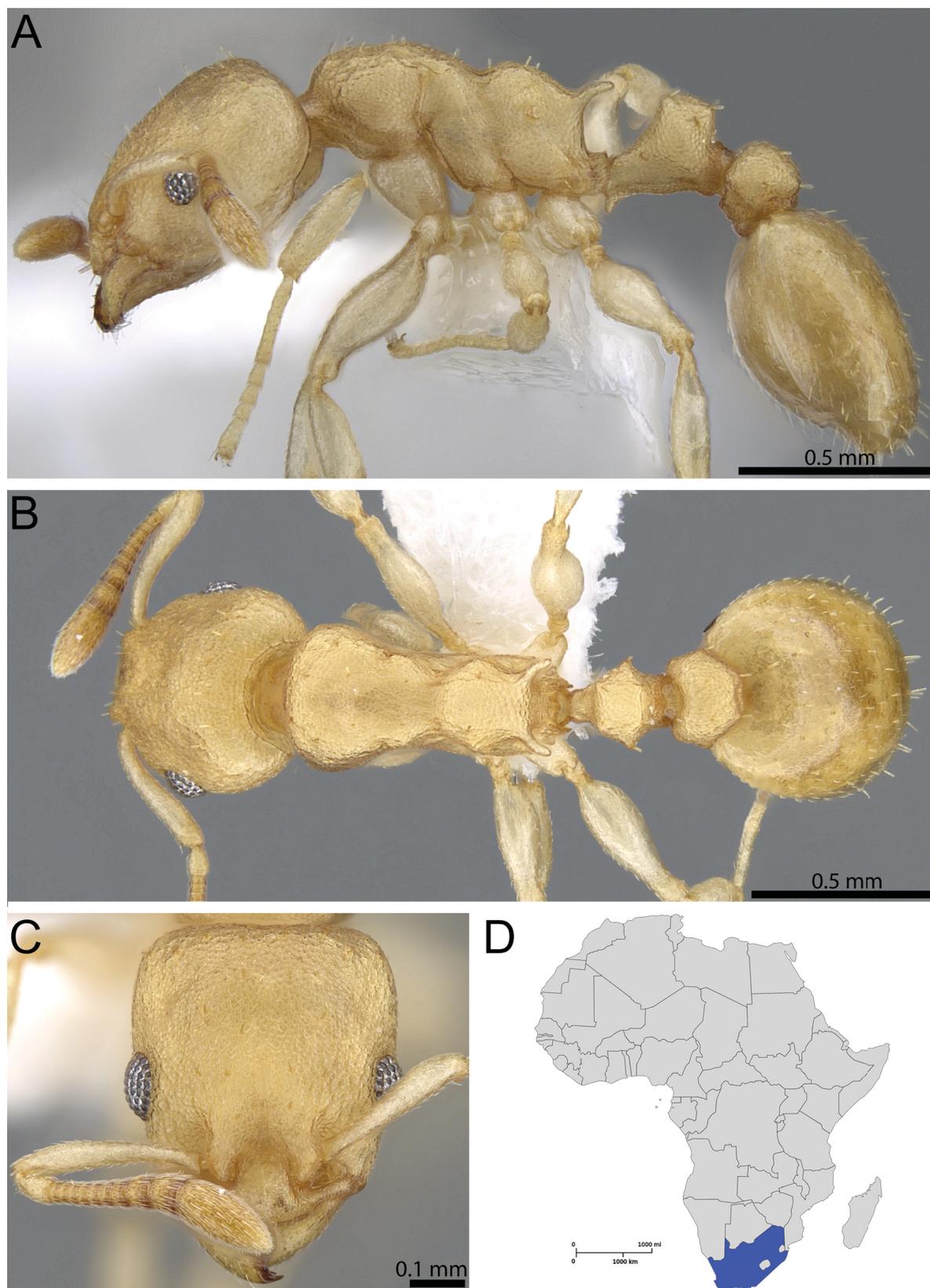
*Nesomyrmex stramineus* was sampled in savannah woodlands and Afromontane forests, where it lives in dead wood on trees (Mbanyana & Robertson 2008).

### **Distribution**

This species occurs only in South Africa and Swaziland. It seems to be rather rare since it is only known from a few collection events.

### **Discussion**

As already mentioned above, initially Bolton (1982) revised the Afrotropical *Nesomyrmex* fauna for the whole region, and more recently Mbanyana & Robertson (2008) for South Africa. Despite the existence of these revisions, there were no clear delimitations of *Nesomyrmex* species groups for the region prior to this study. Bolton (1982) discussed several assemblages of species that shared some morphological characters, but he did not formally define any groups or complexes. In their revision of South African *Nesomyrmex*, Mbanyana & Robertson (2008) provide brief species group definitions for the two groups encountered in South Africa: the *N. angulatus* and *N. simoni* groups. However, they only cover the South African species and omit the remainder of the Afrotropical fauna. In this study, we provide a first



**Fig. 20.** *Nesomyrmex stramineus* (Arnold, 1948) (CASENT0922011). **A.** Body in profile view. **B.** Body in dorsal view. **C.** Head in full-face view. **D.** Map of Africa and Madagascar showing currently known distribution.

assessment of the entire Afrotropical fauna, propose species groups, and provide illustrated identification tools for all groups and most species with the exception of the species of the *N. simoni* group, which was extensively revised in Mbanyana & Robertson (2008). Nevertheless, the species groups proposed here are based on morphology alone, and should be treated as taxonomic convenience groups rather than phylogenetic entities. It is still likely that some or all represent monophyletic clades, but this can only be assessed in detail through a thorough comprehensive phylogenetic study that combines morphological with molecular data.

## Acknowledgements

We would like to thank Jignasha Rana, Stefan Cover, Edward O. Wilson, Kathleen Horton and Patrick McCormack, all from MCZ, for their general assistance and hospitality during the stay of the first author in the famous ant room. Also, we are thankful to Gavin Broad, Peter G. Hawkes and an anonymous reviewer for editing, reviewing and improving a prior version of the manuscript. Furthermore, we highly appreciate Brian L. Fisher's (California Academy of Sciences, San Francisco, USA) on-going efforts to make the majority of ant type images available to the ant community through AntWeb. FHG was granted an Ernst Mayr Travel Grant in Animal Systematics from the MCZ to visit the MCZ ant collection during which the discovery of *N. inhaca* sp. nov. occurred.

## References

- Arnold G. 1948. New species of African Hymenoptera. No. 8. *Occasional Papers of the National Museum of Southern Rhodesia* 2: 213–250.
- Bolton B. 1982. A revision of six minor genera of Myrmicinae (Hymenoptera: Formicidae) in the Ethiopian zoogeographical region. *Bulletin of the British Museum (Natural History), Entomology* 43: 245–307.
- Bolton B. 1994. *Identification Guide to the Ant Genera of the World*. Harvard University Press, Cambridge, Massachusetts.
- Bolton B. 2003. Synopsis and classification of Formicidae. *Memoirs of the American Entomological Institute* 71: 1–370.
- Bolton B. 2016. AntCat: An online catalog of the ants of the world. Available from <http://antcat.org> [accessed 26 Jan. 2016].
- Brown W.L. 1971. Characters and synonymies among the genera of ants. Part IV. Some genera of subfamily Myrmicinae (Hymenoptera: Formicidae). *Breviora* 365: 1–5.
- Brandão C.R.F. 1991. Adendos ao catálogo abreviado das formigas da região Neotropical (Hymenoptera: Formicidae). *Revista Brasileira de Entomologia* 35: 319–412. <http://dx.doi.org/10.5281/zenodo.24565>
- Csösz S. & Fisher B.L. 2015. Diagnostic survey of Malagasy *Nesomyrmex* species-groups and revision of *hafahafa* group species via morphology based cluster delimitation protocol. *Zookeys* 526: 19–59. <http://dx.doi.org/10.3897/zookeys.526.6037>
- De Andrade M.L., Baroni Urbani C., Brandão C.R.F. & Wagensberg J. 1999. Two new species of *Leptothorax* “*Nesomyrmex*” fossils in Dominican amber (Hymenoptera: Formicidae). *Beiträge zur Entomologie* 49: 133–140.
- Emery C. 1896. Studi sulle formiche della fauna neotropica. XVII–XXV. *Bollettino della Società Entomologica Italiana* 28: 33–107.
- Emery C. 1915. Formiche raccolte nell'Eritrea dal Prof. F. Silvestri. *Bollettino del Laboratorio di Zoologia Generale e Agraria della Reale Scuola Superiore d'Agricoltura* 10: 3–26.

- Evenhuis N.L. 2016. The insect and spider collections of the world website. Available from <http://hbs.bishopmuseum.org/codens> [accessed 26 Jan. 2016].
- Forel A. 1894. Abessinische und andere afrikanische Ameisen, gesammelt von Herrn Ingenieur Alfred Ilg, von Herrn Dr. Liengme, von Herrn Pfarrer Missionar P. Berthoud, Herrn Dr. Arth. Müller etc. *Mitteilungen der Schweizerischen Entomologischen Gesellschaft* 9: 64–100. Available from <http://biodiversitylibrary.org/page/10410405> [accessed 24 Nov. 2016].
- Forel A. 1913. Formicides du Congo Belge. Recoltés par MM. Bequaert, Luja, etc. *Revue de Zoologie Africaine* 2: 306–351.
- Forel A. 1916. Fourmis du Congo et d'autres provenances récoltées par MM. Hermann Kohl, Luja, Mayné, etc. *Revue Suisse de Zoologie* 24: 397–460.
- Guénard B. & Economo E.P. 2016. *Antmaps*. Available from <http://www.antmaps.org/> [accessed 26 Jan. 2016].
- Harris R.A. 1979. A glossary of surface sculpturing. *California Department of Food and Agriculture, Bureau of Entomology* 28: 1–31.
- Hita Garcia F. & Fisher B.L. 2015. Taxonomy of the hyper-diverse ant genus *Tetramorium* Mayr in the Malagasy region (Hymenoptera, Formicidae, Myrmicinae) – first record of the *T. setigerum* species group and additions to the Malagasy species groups with an updated illustrated identification key. *Zookeys* 512: 121–153. <http://dx.doi.org/10.3897/zookeys.512.9860>
- Kempf W.W. 1959. A synopsis of the New World species belonging to the *Nesomyrmex*-group of the ant genus *Leptothorax* Mayr (Hymenoptera: Formicidae). *Studia Entomologica* 2: 391–432.
- Mayr G. 1862. Myrmecologische Studien. *Verhandlungen der Kaiserlich-Königlichen Zoologisch-Botanischen Gesellschaft in Wien* 12: 649–776.
- Mayr G. 1895. Afrikanische Formiciden. *Annalen des Kaiserlich-Königlichen Naturhistorischen Museums in Wien* 10: 124–154.
- Mayr G. 1901. Südafrikanische Formiciden, gesammelt von Dr. Hans Brauns. *Annalen des Kaiserlich-Königlichen Naturhistorischen Museums in Wien* 16: 1–30.
- Mbanyana N. & Robertson H.G. 2008. Review of the ant genus *Nesomyrmex* (Hymenoptera: Formicidae: Myrmicinae) in southern Africa. *African Natural History* 4: 35–55.
- Santschi F. 1912. Quelques nouvelles variétés de fourmis africaines. *Bulletin de la Société d'Histoire Naturelle de l'Afrique du Nord* 3: 147–149.
- Santschi F. 1914. *Voyage de Ch. Alluaud et R. Jeannel en Afrique Orientale, 1911–1912. Résultats scientifiques. Insectes Hyménoptères. II. Formicidae*. Librairie A. Schulz, Paris.
- Snelling R.R. 1992. Two unusual new myrmicine ants from Cameroon (Hymenoptera: Formicidae). *Psyche* 99: 95–101. <http://dx.doi.org/10.1155/1992/23808>
- Ward P.S., Brady S.G., Fisher B.L. & Schultz T.R. 2015. The evolution of myrmicine ants: phylogeny and biogeography of a hyperdiverse ant clade (Hymenoptera: Formicidae). *Systematic Entomology* 40: 61–81. <http://dx.doi.org/10.1111/syen.12090>
- Wheeler W.M. 1922. Ants collected by the American Museum Congo Expedition. A contribution to the myrmecology of Africa. *Bulletin of the American Museum of Natural History* 45: 1–1055.
- Wilson E.O. 1955. A monographic revision of the ant genus *Lasius*. *Bulletin of the Museum of Comparative Zoology* 113: 1–201.

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