



Monograph

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Revision of the Eurybrachidae XIV. The Australian genera *Olonia* Stål, 1862 and *Stalobrachys* gen. nov. (Hemiptera: Fulgoromorpha)

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Abstract. The Australian genus of Eurybrachidae Stål, 1862 (Hemiptera, Fulgoromorpha) *Olonia* Stål, 1862 is redescribed and reviewed. Seven new species are described: *O. bourgoini* sp. nov. (N Queensland, Chillagoe), *O. danielsi* sp. nov. (N Queensland, Cape York Peninsula), *O. guillaumei* sp. nov. (N Queensland), *O. hochae* sp. nov. (N Queensland, Undara), *O. monteithi* sp. nov. (Queensland), *O. rylandae* sp. nov. (N Queensland, Chillagoe) and *O. soulierae* sp. nov. (Queensland). *Platybrachys nobilis* (Stål, 1863) is transferred to *Olonia* and the new combination *Olonia nobilis* (Stål, 1863) comb. nov., is proposed. *Olonia ornata* Lallemand, 1928 and *O. apicalis* (Walker, 1851) are removed from *Olonia* and transferred to *Platybrachys* Stål, 1859 and *Maeniana* Metcalf, 1952, respectively. Hence, the new combinations *Platybrachys ornata* (Lallemand, 1928) comb. nov. and *Maeniana apicalis* (Walker, 1851) comb. nov. are proposed. The new genus *Stalobrachys* gen. nov. is described to accommodate *Olonia alboapicata* Jacobi, 1928 and the new combination *S. alboapicata* (Jacobi, 1928) gen. et comb. nov. is proposed. The male genitalia are illustrated and photographs of habitus, a distribution map, biological data and an identification key are provided. The genus *Olonia* currently contains twelve species and the genus *Stalobrachys* gen. nov. has one species.

Keywords. Planthopper, *Ipomoea*, Queensland.

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Introduction

Eurybrachidae is a small family of planthoppers (Fulgoromorpha Evans, 1946) with 41 genera and 193 species, representing only 1.7% of the genera and 1.4% of the species of Fulgoromorpha. The family is restricted to the Old World and is distributed in the Afrotropical, Oriental and Australasian regions, with some species present in the southeasternmost part of the Palaearctic region in China; it is not recorded from Madagascar or Taiwan (Metcalf 1956; Bourgoin 2018). Eurybrachidae represent a characteristic

component of the planthopper fauna of Australia, with 14 genera and 48 species which all belong to the tribe Platybrachini Schmidt, 1908 of the subfamily Platybrachinae Schmidt, 1908. This subfamily counts 20 genera distributed in the Afrotropical, Oriental and Australasian regions. A single genus, *Ancyra* White, 1845, is included in the tribe Ancyrini Schmidt, 1908 and is the only representative of the Platybrachinae in the Oriental region. All the other 19 genera of Platybrachinae belong to the tribe Platybrachini and are endemic either to the Afrotropical (5 genera) or to the Australasian region (Metcalf 1956; Fennah 1964; Bourgoïn 2018). The Australian Eurybrachidae are known to feed on trees and shrubs belonging to two families, Myrtaceae (*Eucalyptus* spp.) and Fabaceae (*Acacia* spp.); however, the host plants are still very poorly documented and remain unknown for most of the species (Constant 2006a, 2006b, 2008b; Bourgoïn 2018). When I began the revision of Eurybrachidae (Constant 2004), it appeared necessary to redefine and review all existing genera. The genus *Olonia* was rapidly recognized as problematic, being poorly defined and containing species erroneously placed in it (Constant 2006b). The scrutiny of the types of all species placed in *Olonia* and putatively closely related genera (e.g., *Platybrachys* Stål, 1859, *Lyncilia* Stål, 1863, *Maeniana* Metcalf, 1952), together with the study of numerous unidentified specimens, now allow the completion of a comprehensive revision of the genus *Olonia* as the fourteenth part of the ongoing revision of the family.

The present paper aims to fully revise the taxonomy of *Olonia*, to describe seven new species, to fix the status of *Lyncilia nobilis* Stål, 1863, *Olonia ornata* Lallemand, 1928 and *O. apicalis* (Walker, 1851), and to describe a new genus, *Stalobrachys* gen. nov., to include *Olonia alboapicata* Jacobi, 1928, together with providing complete illustrations of all the species, an identification key, distribution maps and biological data.

Material and methods

The types of all described species have been studied and as much material as possible has been examined. The genitalia of all the males have been checked.

The genitalia were extracted after boiling the abdomen for some minutes in a 10% solution of potassium hydroxide (KOH) at about 100°C. Some drops of saturated alcoholic Chlorazol black solution were added for contrasting (Carayon 1969). The pygofer was separated from the abdomen and the aedeagus dissected with a needle blade for examination in 70% ethanol. All resulting fragments were then placed in glycerine for preservation in a tube attached to the pin of the corresponding specimen.

For routine identification, the abdomen was removed and the genitalia examined directly. In this case, the dry abdomen was placed in a gelatin capsule or glued on cardboard, pinned under the specimen.

Posterior wings were also mounted for a large number of specimens; they have been glued on white cardboard or transparent plastic rectangles and then pinned under the specimen.

The description of the female genitalia follows Bourgoïn (1993) with some additions from the studies of Soulier-Perkins (1997) and Soulier-Perkins & Bourgoïn (1998) on the family Lophopidae; the description of the wing venation follows Bourgoïn *et al.* (2015).

The measurements were taken as in Constant (2004) and the following abbreviations are used:

- BF = maximum breadth of the frons
- BT = maximum breadth of the thorax
- BTg = maximum breadth of the tegmen

- BV = maximum breadth of the vertex
BW = maximum breadth of the posterior wing
LF = length of the frons in midline
LM = length of the mesonotum in midline
LP = length of the pronotum in midline
LT = total length from anterior margin of vertex to apex of tegmina
LTg = maximum length of the tegmen
LV = length of the vertex in midline
LW = maximum length of the posterior wing

The metatibiotarsal formula gives the number of spines on the (side of metatibia) apex of the metatibia/apex of first metatarsus/apex of second metatarsus.

For each picture a number of photographs were taken with a Canon 700D camera equipped with a Tamron 90 mm macro lens and stacked with Combine Z software. They were optimized with Adobe Photoshop CS3. Observations were done with a Leica MZ8 stereo microscope. The distribution maps were produced with SimpleMappr (Shorthouse 2010), which provides a delimitation of bioregions similar to that proposed by the Interim Biogeographic Regionalisation for Australia, Version 7 (see <http://www.environment.gov.au/land/nrs/science/ibra>). For the transcription of the labels of the types, the wording on each single label is given *verbatim* placed within quotes, with supplementary information not on the label given in square brackets where appropriate. In the results section, species are treated in alphabetical order.

Acronyms used for the collections are as follows:

- AMS = Australian Museum, Sydney, New South Wales, Australia
ANIC = Australian National Insect Collection, CSIRO, Canberra, ACT, Australia
ASCU = Agricultural Scientific Collections Unit, Orange Agricult. Inst., Orange, NSW, Australia
BMNH = The Natural History Museum, London, United Kingdom
BPBM = Bernice P. Bishop Museum, Honolulu, Hawaii, USA
CAS = California Academy of Sciences, San Francisco, California, USA
INHS = Illinois Natural History Survey, Prairie Research Institute, Champaign, Illinois, USA
MNHN = Muséum national d'Histoire naturelle, Paris, France
MVMA = Museum of Victoria, Melbourne, Victoria, Australia
NHMW = Naturhistorisches Museum, Vienna, Austria
NHRS = Naturhistoriska Riksmuseet, Stockholm, Sweden
QDPI = Queensland Department of Primary Industries, Indooroopilly, Queensland, Australia
QM = Queensland Museum, South Brisbane, Queensland, Australia
QPIM = Department of Primary Industries, Mareeba, Queensland, Australia
RBINS = Royal Belgian Institute of Natural Sciences, Brussels, Belgium
RMNH = Nationaal Natuurhistorisch Museum ('Naturalis'), Leiden, the Netherlands
SAM = South Australian Museum, Adelaide, South Australia, Australia
TAMU = Texas A & M University, College Station, Texas, USA
USNM = National Museum of Natural History, Washington DC, USA
ZMHB = Museum für Naturkunde der Humboldt-Universität, Berlin, Germany
ZMPA = Polish Academy of Sciences, Museum of the Institute of Zoology, Warsaw, Poland

Results

Class Hexapoda Blainville, 1816
Order Hemiptera Linnaeus, 1758
Suborder Auchenorrhyncha Duméril, 1806
Infraorder Fulgoromorpha Evans, 1946
Superfamily Fulgoroidea Latreille, 1807
Family Eurybrachidae Stål, 1862
Subfamily Platybrachinae Schmidt, 1908
Tribe Platybrachini Schmidt, 1908

Genus *Olonia* Stål, 1862
Figs 1–3

Olonia Stål, 1862: 488 (description in key; list of included species; type species: *Eurybrachys rubicunda* Walker, 1851, by subsequent designation in Distant 1906: 206).

Lyncilia Stål, 1863: 248 (description) syn. nov. (type species: *Lyncilia nobilis* Stål, 1863, by monotypy).

Olonia – Stål 1863: 250 (description of a new species). — Atkinson 1886: 13 (incomplete English translation of key to genera of Eurybrachidae in Stål 1862). — Karsch 1890: 60 (compared with *Metoponitys* Karsch, 1890); 1895: 215 (compared with *Aspidonitys* Karsch, 1895). — Melichar 1903: 67 (placed in a group of eurybrachid genera without a spine under the eye). — Distant 1906: 206 (type species designation, new species), 207 (compared to *Yarrana* Distant, 1906). — Kirkaldy 1907: 105 (listed without comment). — Schmidt 1908: 243 (placed in Platybrachini (= current Platybrachinae)). — Hacker 1924: 40 (compared with *Platybrachys* Stål, 1859; notes on *O. viridiventris* Stål, 1863 (now in *Hackerobrachys* Constant, 2006)). — Metcalf 1936: 131 (senior homonym of *Olonia* Muir, 1925); 1938: 294 (senior homonym of *Olonia* Muir, 1925); 1947: 163 (listed as typical genus of the Australian fauna); 1956: 63 (catalogued). — Fennah 1964: 159 (in key to genera of Platybrachini), 162 (compared with *Maon* Fennah, 1964). — Constant 2005a: 41 (mentioned in historical review of *Metoponitys* Karsch, 1890); 2006a: 47 (mentioned as needing revision); 2006b: 31 (compared with *Hackerobrachys* Constant, 2006 and *Fletcherobrachys* Constant, 2006).

Lyncilia – Karsch 1895: 215 (compared with *Aspidonitys* Karsch, 1895). — Schmidt 1908: 243 (placed in the Platybrachini (= current Platybrachinae)). — Metcalf 1956: 68 (catalogued). — Fennah 1964 (synonymized under *Platybrachys* Stål, 1859 (erroneous)).

non *Olonia* – Muir 1925: 161 (new genus of Cixiidae (junior homonym of *Olonia* Stål, 1862)). — Metcalf 1936: 131 (*Muirolonia* as replacement name of *Olonia* Muir, 1925 in Cixiidae); 1938: 294 (*Muirolonia* as replacement name of *Olonia* Muir, 1925 (*Muirolonia* erroneously stated as being a new genus)).

Diagnosis

Rather small-sized (6–12 mm), dark brown variegated with black and white, tegmina usually with a white marking along costal margin on nodal line and posterior wings usually with a white marking near externoapical angle, sometimes orange on disc. The genus can be recognized by the following set of characters:

- (1) gonostyli strongly sclerified and fused basally on about $\frac{1}{3}$ of length
- (2) gonostyli divided into a centroventral and a laterodorsal part

- (3) gonostyli with laterodorsal part with large lateral process projecting laterally and bearing dorsoapical, articulate, spoon-shaped process
- (4) aedeagus strongly reduced with dorsal portion of phallobase projecting dorsally as a spine or hook

The genus shares the condition of possessing a spoon-shaped process on the gonostyli with the Australian genera *Chewobrachys* Constant, 2008, *Fletcherobrachys* Constant, 2006, *Hackerobrachys* Constant, 2006, *Maeniana* Metcalf, 1952, *Nirus* Jacobi, 1928 and *Stalobrachys* gen. nov. but differs

– from *Chewobrachys* by its smaller size (6–12 mm; 12–16 mm in *Chewobrachys*), the dark brown colour (greyish brown in *Chewobrachys*) and the male gonostyli divided into a centroventral and a laterodorsal part (not divided in *Chewobrachys*)

– from *Fletcherobrachys* by the bright red abdomen (green to orange in *Fletcherobrachys*), the hind wings brown or with discal orange marking (basal half white in *Fletcherobrachys*), the absence of sexual dimorphism on median tibiae (with externodistal process in females of *Fletcherobrachys*) and the male gonostyli fused only on basal $\frac{1}{3}$ and divided into a centroventral and a laterodorsal part (fused along most of their length and not divided in *Fletcherobrachys*)

– from *Hackerobrachys* by the frons being brown and slightly convex (bright yellow to red and strongly convex in *Hackerobrachys*), the posterior wings with some white and/or orange markings and rounded apically (uniformly brown with apex subquadrate in *Hackerobrachys*) and the male gonostyli fused only on basal $\frac{1}{3}$ and divided into a centroventral and a laterodorsal part (fused along most of their length and not divided in *Hackerobrachys*)

– from *Maeniana* by the pygofer having the posterior margin strongly sinuate (with elongate laterodorsal process projecting posteriorly in *Maeniana*), the gonostyli fused on about $\frac{1}{3}$ of their length (not or very shortly fused in *Maeniana*), and the laterodorsal part of the gonostyli with a strong spine or hook and with the lateral process bearing a spoon-shaped process (laterodorsal part elongate and laminate, without spine or hook, and bearing a spoon-shaped process apically in *Maeniana*)

– from *Nirus* by the frons being slightly convex (concave in *Nirus*), the pygofer broader on the dorsal $\frac{2}{3}$ (on ventral half in *Nirus*), and the laterodorsal part of the gonostyli with a strong spine or hook and with the lateral process bearing a spoon-shaped process (laterodorsal part of gonostyli large and laterally compressed, not strongly sclerified and without a spine or hook in *Nirus*)

– from *Stalobrachys* gen. nov. by the narrower posterior wings with LW/BW = 1.7–2.0 (1.5 in *Stalobrachys* gen. nov.), the pygofer with the posterior margin strongly sinuate (with elongate laterodorsal process projecting posteriorly in *Stalobrachys* gen. nov.), the gonostyli fused on about $\frac{1}{3}$ of their length (not or very shortly fused in *Stalobrachys* gen. nov.), and the laterodorsal part of the gonostyli with a strong spine or hook and with the lateral process bearing a spoon-shaped process (laterodorsal part elongate and laminate, without spine or hook, and bearing a spoon-shaped process apically in *Stalobrachys* gen. nov.)

The genus *Loisobrachys* Constant, 2008 is known from a single female and was placed close to *Hackerobrachys* (Constant 2008b). Hence, the males of the genus potentially share the condition of possessing gonostyli with a spoon-shaped process. However, *Olonia* can easily be separated from *Loisobrachys* by its slightly convex frons (strongly convex in *Loisobrachys*).

Historical review

1. Historical characters recognition review

Olonia was described by Stål (1862) within a key to the genera of Eurybrachidae without a spine under the eyes and with the clavus of the tegmina closed. From the key, the following characters could be extrapolated to define *Olonia*:

- (1) posterior tibiae with 3 lateral spines
- (2) frons transverse, with sides angulate
- (3) pro- and mesonotum broader than combined length
- (4) antennae short, placed under the eye

More recently, a key to the genera of Platybrachini was proposed by Fennah (1964), including *Olonia* for which the following distinctive combination of characters could be extrapolated:

- (1) tegmina with CuA vein forked close to the nodal line
- (2) tegmina with MP vein forked very close to the base
- (3) genae without knob-like process under the eye
- (4) frons more or less flat, without depressed areas near the base
- (5) antennae not surpassing the eyes
- (6) genital styles (= gonostyli) of male separated

Characters (2) and (6) are not correct: in *Olonia*, the MP is not forked very close to the base and Fennah's statement probably comes from confusion with the ScP and RP, which diverge very basally, and the gonostyli in the male genitalia are fused basally.

2. Historical species review

Stål (1862) erected the genus *Olonia* for three species from Australia, *Eurybrachys rubicunda* Walker, 1851, *E. apicalis* Walker, 1851 and *E. transversa* Walker, 1858. The year after, Stål (1863) added one species from Queensland, *O. viridiventris* Stål, 1863. Species were progressively added to the genus: *O. picea* by Kirkaldy (1906) (Queensland), *O. marginata* by Distant (1906) (Queensland), *O. alboapicata* and *O. nigroapicata* by Jacobi (1928) (Queensland and Western Australia, respectively), and *O. ornata* by Lallemand (1928) (Northern Territory). Jacobi (1928) synonymized *O. picea* Kirkaldy, 1906 with *O. transversa* (Walker, 1858), a view that I did not follow (Constant 2005b), restoring *O. picea* as a separate species and considering *O. transversa* as a *nomen dubium* because the available specimen labelled as type in BMNH does not match the original description by Walker (1858). One last species, *Cicada maura* Fabricius, 1775, was transferred to *Olonia* by Evans (1933) after being placed in the genus *Eurymela* Le Peletier & Serville, 1825 (Cicadellidae: Eurymelinae) for more than a century. Metcalf (1956) included 9 species in the genus in his catalogue of the Eurybrachidae. More recently, I transferred *O. viridiventris* Stål, 1863 to the genus *Hackerobrachys* Constant, 2006 and synonymized *O. nigroapicata* Jacobi, 1928 with *Fletcherobrachys stillata* (Bergroth, 1907) (Constant 2006b).

3. Historical classification review

The genus *Olonia* was placed by Schmidt (1908) in the tribe Platybrachini Schmidt, 1908 (equivalent to the present Platybrachinae, as the family Eurybrachidae was at that time treated as a subfamily of the Fulgoridae) based on the following set of characters:

- (1) clavus closed, with the claval veins (= Pcu and A1) fused before the apex of the clavus and Pcu+A1 reaching the apex of the clavus
- (2) ventral margin of eyes without a spine

Metcalf (1956) placed the genus in the tribe Platybrachini of the Platybrachinae, a view followed by Fennah (1964).

Description

COLOURATION. Head, pro- and mesonotum, and tegmina brown, from pale brown to nearly black, usually variegated with darker and paler areas. Tegmina often with yellowish marking in middle of clavus on vein A1; triangular white marking along costal margin on nodal line, rarely missing; smaller white marking at apicosutural angle, rarely missing; sometimes white marking along costal margin at midlength. Posterior wings brown, usually darker on apical half; sometimes with orange marking in middle; white triangular marking at apicocostal angle, rarely missing. Pro- and mesofemora with 2 pale rings, obsolete in dark species; pro- and mesotibiae with 3 pale rings, obsolete in dark species; metafemora red to brown, darker apically; metatibiae brown. Abdomen and ventral face of thorax bright red; genital segments brown to black; abdominal segment VII white in females.

HEAD. As broad as thorax; vertex 4–5 times as broad as long, concave, with all margins slightly carinate; anterior and posterior margins rounded, parallel; frons about 1.7–1.95 times as broad as long, slightly convex, slightly wrinkled to rugulose, with peridiscal carina slightly marked; upper margin of frons straight in normal view; clypeus slightly surpassing anterior trochanters, elongate, with median carina towards apex; labium reaching hind coxae, with apical segment longer than broad, acuminate, shorter and more slender than penultimate; no infra-ocular spines on genae; ocelli absent; antennae elongate, not surpassing eye, not visible from above; scape about as long as broad, pedicel subcylindrical, elongate, narrowing towards apex.

THORAX. About 1.45 times as broad as combined length of pro- and mesonotum; pronotum about half as long as mesonotum; pronotum with disc weakly wrinkled, carina parallel to anterior margin and with two slightly impressed points on disc; mesonotum with disc weakly wrinkled, median and peridiscal carinae slightly marked.

TEGMINA. Slightly convex in smaller species to nearly flat in the larger ones; curving down at nodal line, often with apex slightly curved upwards; elongate, about 2.1–2.5 times as long as broad; costal margin slightly sinuate; apical margin obliquely rounded; sutural margin slightly oblique after clavus. Venation: veins ScP+RA and RP separated close to base; first fork of MP on basal $\frac{1}{3}$; first fork of CuA slightly before apex of clavus; clavus closed; Pcu and A1 fused at $\frac{2}{3}$ of clavus length; Pcu+A1 reaching apical angle of clavus; numerous elongate cells along posterior half of costal margin and along apical margin.

POSTERIOR WINGS (Fig. 1A). Well developed, about as broad as tegmina, rounded apically; moderately narrow: LW/BW = 1.7–2.0; anal area moderately developed; sutural margin weakly trilobous; not reaching apex of tegmina at rest; dark brown to black, with apicosutural triangular white spot, rarely missing; sometimes with central, small to very large, orange marking. All main veins visible from base, forked after nodal line and sometimes forming closed cells; transverse veinlets delimiting elongate cells on apical $\frac{1}{3}$; veins A1 and A2 sometimes with 2–3 terminals. Arrangement of secondary veins and veinlets variable between specimens and sometimes between the two wings of the same specimen.

LEGS. Pro- and mesofemora and -tibiae dorsoventrally flattened, elongate and slender; metatibiae with 3 lateral and 9 apical spines; first metatarsomere ventrally with pad of microsetae at interno-apical angle and group of 6 spines (Fig. 1B). Metatibiotarsal formula: (3) 9/4/0.

MALE GENITALIA. Pygofer rather short, higher than long and sinuate in lateral view; posterior margin roundly projecting at dorsal $\frac{1}{3}$. Anal tube dorsoventrally flattened, elongate, with lateral margins usually curved lateroventrally; epiproct at basal $\frac{1}{3}$. Gonostyli fused basally, well sclerified, deeply divided in centroventral and laterodorsal parts, elongate and spinose or sword-shaped; laterodorsal part with strong spine or hook, sometimes bifid, and with large lateral process projecting laterad and bearing dorsoapical, articulate, spoon-shaped process. Aedeagus strongly reduced; dorsal portion of phallobase

point- or hook-shaped; ventral portion of phallobase as more or less reduced, sclerified lamina; phallus membranous, dorsoventrally flattened.

FEMALE TERMINALIA (Fig. 2) (based on *O. marginata*). Abdominal segment VI slightly broader apically and with posterior margin largely emarginate medially in ventral view, and with a small process projecting lateroventrally on each side of emargination (Fig. 2C); anal tube elongate and narrow, curved postero-ventrad, v-shaped in cross section beyond anus, lanceolate in dorsal view (Fig. 2A–B, D–E); gonopods unlobed, projecting dorsolaterad, longer than high, not surpassing anal tube (Fig. 2A–B, D–E); gonapophysis IX large, elongate, apically rounded and curved dorsad (Fig. 2A–B, D–E); gonocoxae VIII resembling reniform inflated pouch (Fig. 2A–E); gonapophysis VIII dorsoventrally flattened and fused together and with sternite VII in a large semicircular lobe slightly emarginate apically (Fig. 2A–E); anterior vagina small and membranous; posterior vagina strongly sclerified, dorsoventrally flattened, short and broad basally, constricted and subtriangular distally (Fig. 2D); bursa copulatrix attached posterodorsally, oval, much larger than posterior vagina (Fig. 2D–E); walls bearing weak, longitudinally reticulated ornamentation (Fig. 2D–E).

SEXUAL DIMORPHISM. Females slightly larger than males. Colour dimorphism also present, in different patterns according to species, sometimes combined:

- (1) white markings on anterior and posterior wings more developed in males
- (2) orange marking on posterior wing larger in males
- (3) apical margin of tegmina and posterior wings bordered with white in females, not in males

SIZE. ♂: 6.2–10.1 mm; ♀: 7.2–11.2 mm.

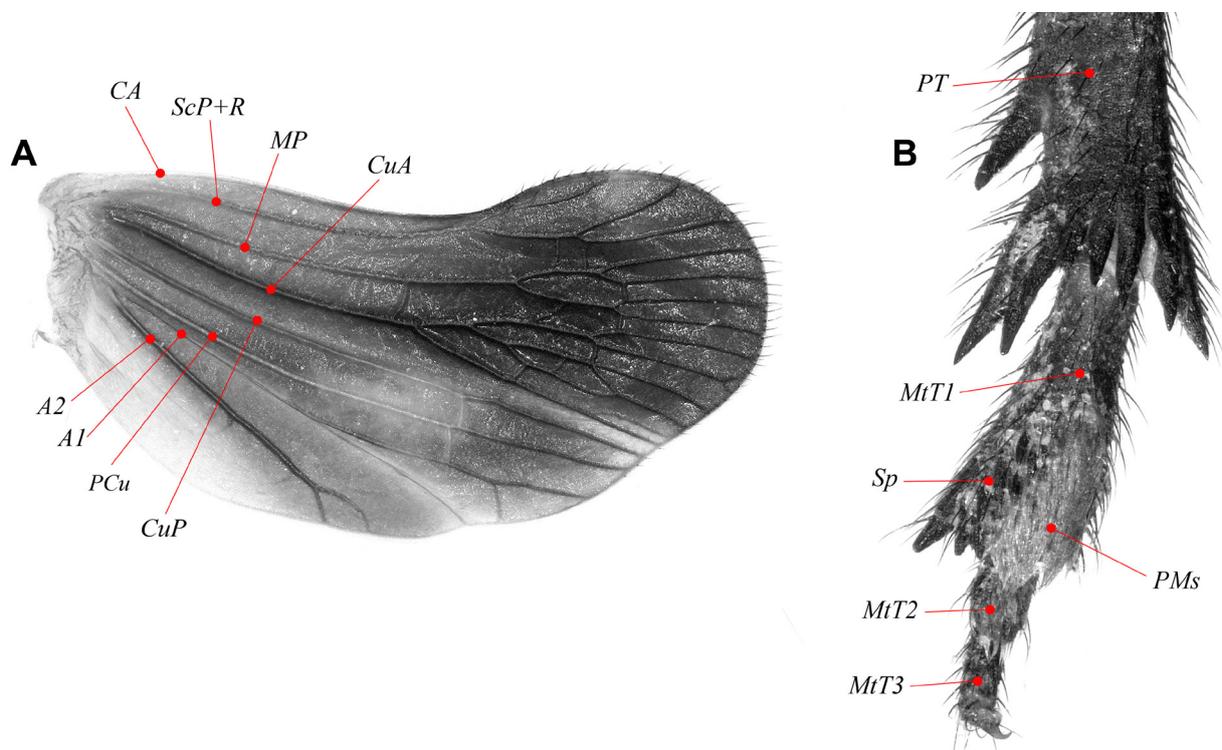


Fig. 1. *Olonia* Stål, 1862. **A.** Right posterior wing, venation. **B.** Right posterior leg, distal portion, ventral view. Abbreviations: *A1* = first anal vein; *A2* = second anal vein; *CA* = costa anterior; *CuA* = cubitus anterior; *CuP* = cubitus posterior; *MP* = media posterior; *MtT1* = first metatarsomere; *MtT2* = second metatarsomere; *MtT3* = third metatarsomere; *PCu* = postcubitus; *PMS* = pad of microsetae; *PT* = posterior tibia; *ScP + R* = subcosta posterior + radius; *Sp* = spines.

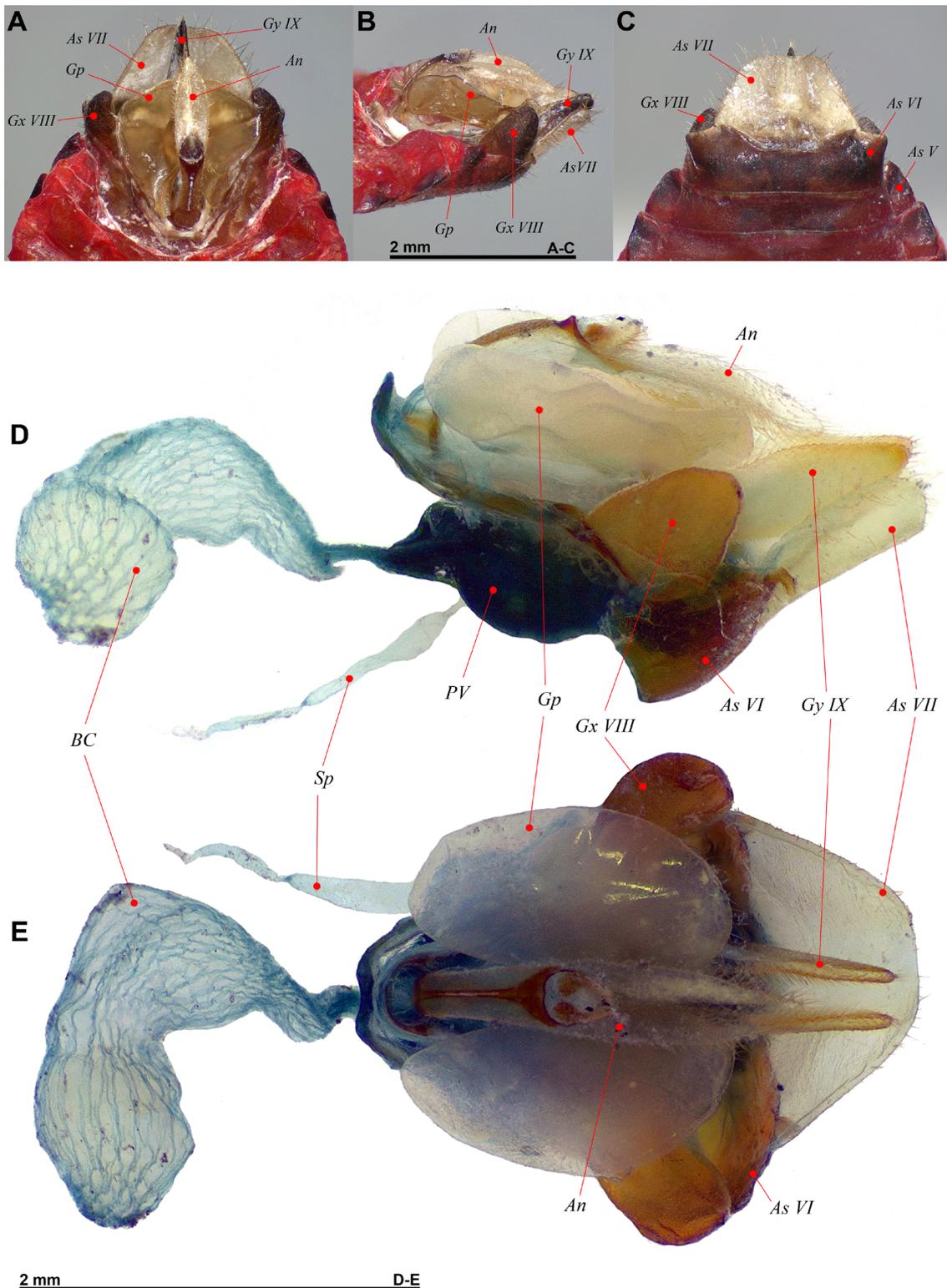


Fig. 2. *Olonia* Stål, 1862, female genitalia. **A.** External aspect, dorsal view. **B.** External aspect, left lateral view. **C.** External aspect, ventral view. **D.** Dissected, left lateral view. **E.** Dissected, dorsal view. Abbreviations: *An* = anal tube; *As V* = fifth abdominal sternite; *As VI* = sixth abdominal sternite; *As VII* = seventh abdominal sternite; *BC* = bursa copulatrix; *Gp* = gonoplac; *Gx VIII* = gonocoxa VIII; *Gy IX* = gonapophysis IX; *PV* = posterior vagina; *Sp* = spermatheca.

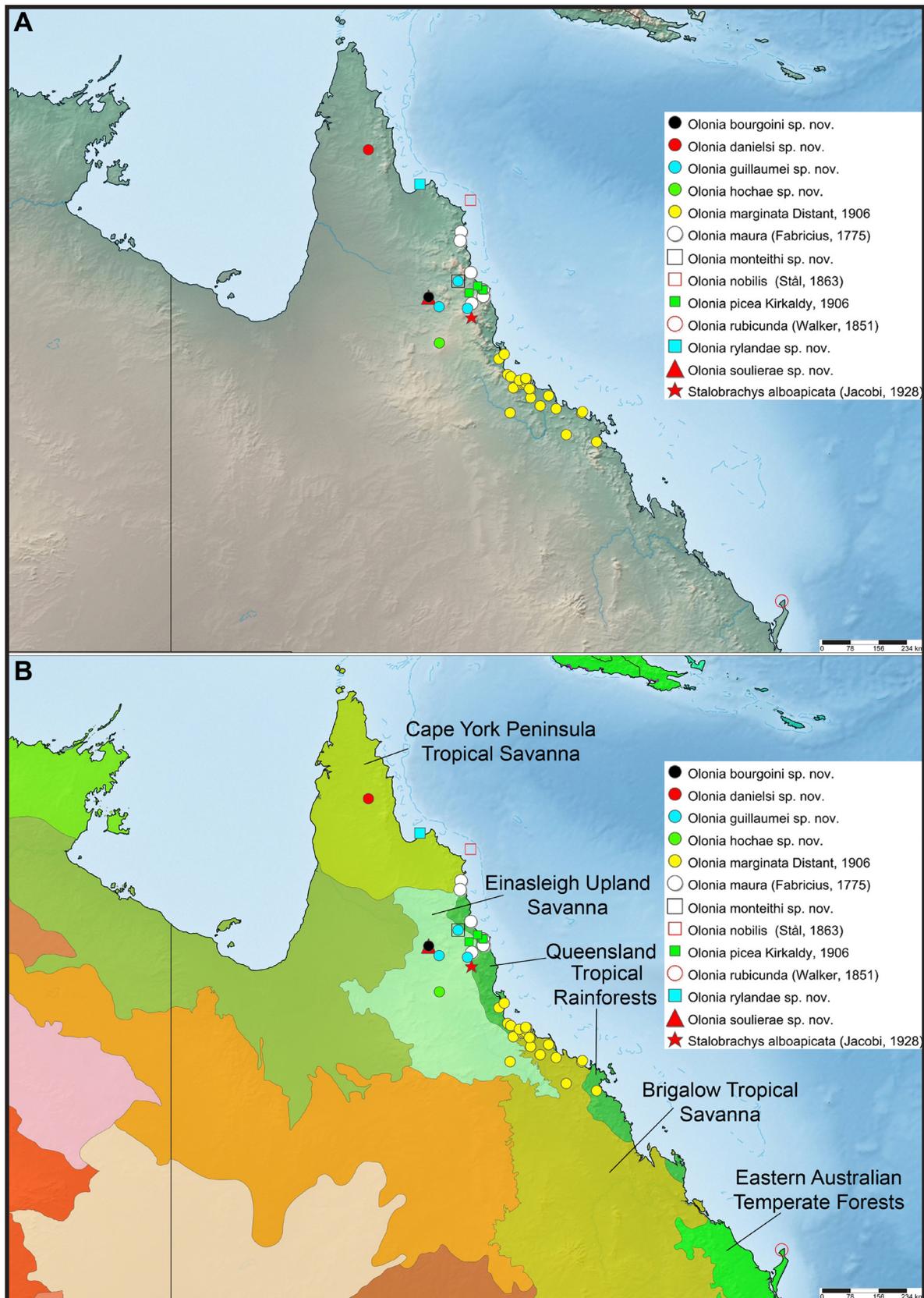


Fig. 3. *Olonia* spp. and *Stalobrachys alboapicata* gen. et comb. nov. (Jacobi, 1928), distribution maps. **A.** Topographic map. **B.** Bioregion map.

Distribution

Australia: eastern and northern Queensland, also on islands (Fig. 3).

Biology

According to the available data, species of *Olonia* seem to be present all year round and to be polyphagous, living on plants close to the ground. They were recorded from plants in the families Bursaceae, Convolvulaceae, Myrtaceae, Solanaceae, Urticaceae and Vitaceae. Known habitats range from open forests to beaches. All species seem to have a rather restricted distribution. Species of *Olonia* are present in all bioregions included in the distribution range of the genus (Fig. 3B).

Species included (12)

- O. bourgoini* sp. nov.
- O. danielsi* sp. nov.
- O. guillaumei* sp. nov.
- O. hochae* sp. nov.
- O. marginata* Distant, 1906
- O. maura* (Fabricius, 1775)
- O. monteithi* sp. nov.
- O. nobilis* (Stål, 1863) comb. nov.
- O. picea* Kirkaldy, 1906
- O. rubicunda* (Walker, 1851)
- O. rylandae* sp. nov.
- O. soulierae* sp. nov.

Remarks

Olonia apicalis (Walker, 1851) and *Olonia ornata* Lallemand, 1928 are here excluded from *Olonia* and transferred to *Maeniana* Metcalf, 1952 and *Platybrachys* Stål, 1859, respectively (see below).

Identification key to the species of *Olonia*

The following key uses the characters of the male genitalia; *O. rubicunda* (Walker, 1851) is not included, as it is only known from a single female. Females should not be identified unless a male from the same collecting event is available to support the identification.

1. Process of the laterodorsal part of gonostyli bifid (Figs 30D, 33D)2
 - Process of the laterodorsal part of gonostyli not bifid (Figs 5D, 10D, 19D)4
2. Process of the centroventral part of gonostyli with several teeth apically (Fig. 30C)
 -***O. maura*** (Fabricius, 1775)
 - Process of the centroventral part of gonostyli without teeth apically (Figs 33C, 42C)3
3. Processes of the laterodorsal part of gonostyli convergent apically, subequal in length and surpassing process of centroventral part in ventral view (Fig. 42A, C–D) ...***O. picea*** (Kirkaldy, 1906)
 - Processes of the laterodorsal part of gonostyli slightly diverging apically with dorsal one shorter than ventral one, and not reaching level of apex of process of centroventral part in ventral view (Fig. 33A, C–D)***O. monteithi*** sp. nov.
4. Processes of the laterodorsal part of gonostyli surpassing processes of centroventral part in ventral view (Figs 19C, 46C)5
 - Processes of the laterodorsal part of gonostyli shorter than processes of centroventral part in ventral view (Figs 5C, 10C, 37C)6

5. Processes of the laterodorsal part of gonostyli strongly sinuate with central portion straight (Fig. 46A, C) *O. rylandae* sp. nov.
– Processes of the laterodorsal part of gonostyli strongly and regularly curved ventrally (Fig. 19A, C) *O. marginata* Distant, 1906
6. Anal tube elongate and narrow, more than 3 times as long as broad, with sides subparallel (Figs 5B, 10B) 7
– Anal tube broader, less than 2.5 times as long as broad, with sides curved (Figs 8B, 37B) 9
7. Processes of the laterodorsal part of gonostyli falcate, broader at midlength and incurved (Fig. 5A, C–D) *O. bourgoini* sp. nov.
– Processes of the laterodorsal part of gonostyli regularly narrowing from base to apex and not incurved (Fig. 10A, C–D) 8
8. Processes of the laterodorsal part of gonostyli curved laterally (Fig. 10C–D); process of centroventral part of gonostyli with ventral margin convex in lateral view (Fig. 10A) *O. guillaumei* sp. nov.
– Processes of the laterodorsal part of gonostyli curved ventrally (Fig. 13C–D); process of centroventral part of gonostyli with ventral margin slightly concave in lateral view (Fig. 13A) *O. hochae* sp. nov.
9. Anal tube with apical margin notched and lateral margins subparallel beyond epiproct (Fig. 48B); process of centroventral part of gonostyli very elongate and narrow (Fig. 48A, C) *O. soulierae* sp. nov.
– Anal tube with apical margin not notched and lateral margins not subparallel beyond epiproct (Figs 8B, 37B); process of centroventral part of gonostyli not very elongate and narrow (Figs 8A, C, 37A, C) 10
10. Processes of the centroventral part of gonostyli incurved and slightly surpassing processes of lateroventral part in ventral view (Fig. 37C); anal tube regularly narrowing beyond epiproct (Fig. 37B) *O. nobilis* (Stål, 1863)
– Processes of the centroventral part of gonostyli strongly hooked dorsally at apex and strongly surpassing processes of lateroventral part in ventral view (Fig. 8C); anal tube broadening beyond epiproct to $\frac{2}{3}$ of its length (Fig. 8B) *O. danielsi* sp. nov.

Olonia bourgoini sp. nov.

[urn:lsid:zoobank.org:act:E8E1C13D-620D-499D-860C-8A6288526E9D](https://zoobank.org/urn:lsid:zoobank.org:act:E8E1C13D-620D-499D-860C-8A6288526E9D)

Figs 3–6

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings with conspicuous orange marking (Figs 4E, 6E)
- (2) pro- and mesofemora and tibiae largely yellowish (Figs 4A–B, 6A–B)
- (3) anal tube of male oblong with posterior margin emarginate (Fig. 5B)
- (4) centroventral part of gonostyli with long laminate process (Fig. 5A, C)
- (5) laterodorsal part of gonostyli with falcate process directed centroventrally (Fig. 5A, C–D)
- (6) rather large size: 9–10 mm

Etymology

This species is dedicated to Prof. Dr Thierry Bourgoin (MNHN), who collected the specimens of the type series.

Material examined

Holotype

AUSTRALIA • ♂; Queensland, Chillagoe Haunted Cave; 17°06' S, 144°25' E; "Muséum Paris, Australie (Queensland) Près de Chilagoe Hounted Cave-Cave 1", "S17°06' E144°25'", 13.III.1997, A. Soulier-Perkins & Th. Bourgoïn rec.", "QM-T244701"; QM.

Paratypes

AUSTRALIA • 1 ♂; same collection data as for holotype; MNHN • 1 ♀; "Australie, Chilagoe GPS 300, 11/12-III-1997, Th. Bourgoïn réc.", "Muséum Paris, piège lumineux, Th. Bourgoïn réc."; MNHN.

Note

On the labels of these specimens, "Chilagoe" and "Hounted" should be spelled "Chillagoe" and "Haunted", respectively.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 1): 9.15 mm; ♀ (n = 1): 9.7 mm; BV/LV = 4.2; BF/LF = 1.75; LP+LM/BT = 0.7; Ltg/BTg = 2.4; LW/BW = 1.77.

Male

HEAD (Fig. 4A–D). Vertex concave with anterior and posterior margins parallel, curved; variegated brown and yellow-brown with darker marking at lateral angles. Frons pale yellowish variegated with brown, with superolateral and dorsal margins broadly bordered with black and 2 transverse, aligned black markings above middle. Clypeus elongate, pale yellowish, with basal, black, horseshoe-shaped marking and oblique brown lines on sides. Genae pale yellowish, slightly darker around eyes. Labium dark brown, reaching metacoxae. Antennae dark brown, paler dorsally; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 4A–D). Pronotum brown variegated with yellowish and with blackish marking; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax brown, turning to pale yellowish ventrally. Mesonotum brown variegated with yellowish and black markings along median carina and along external side of peridiscal carinae; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae dark brown, paler dorsally.

TEGMINA (Fig. 4A, C). Brown variegated with yellowish; broad apical band and irregular markings along basal and costal margins black; pale yellow line on vein A1 at half length of clavus; large triangular white marking on costal margin on nodal line; white spot at apicosutural angle. Maximum breadth at nodal line; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 4E). Black-brown with darker marking at base of apical half along costal margin; elongate, transverse, with triangular white marking at apicocostal angle, extending on 6 cells; elongate orange marking between anal fold and vein CuP at $\frac{2}{3}$, not reaching margin. Margin of anal area sinuate; sutural margin with 2 clefts, cubital one slightly marked.

LEGS (Fig. 4A–D). Pro- and mesocoxae yellowish. Pro- and mesofemora yellowish with some brown markings and with black markings near apex. Pro- and mesotibiae yellowish with 3 ring-shaped black markings, larger one near apex. Pro- and mesotarsi brown with basal half of third tarsomere yellowish. Metacoxae and metafemora reddish yellow. Metatibiae yellowish with 3 lateral and 8 apical black-brown spines. Metatarsi yellowish with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

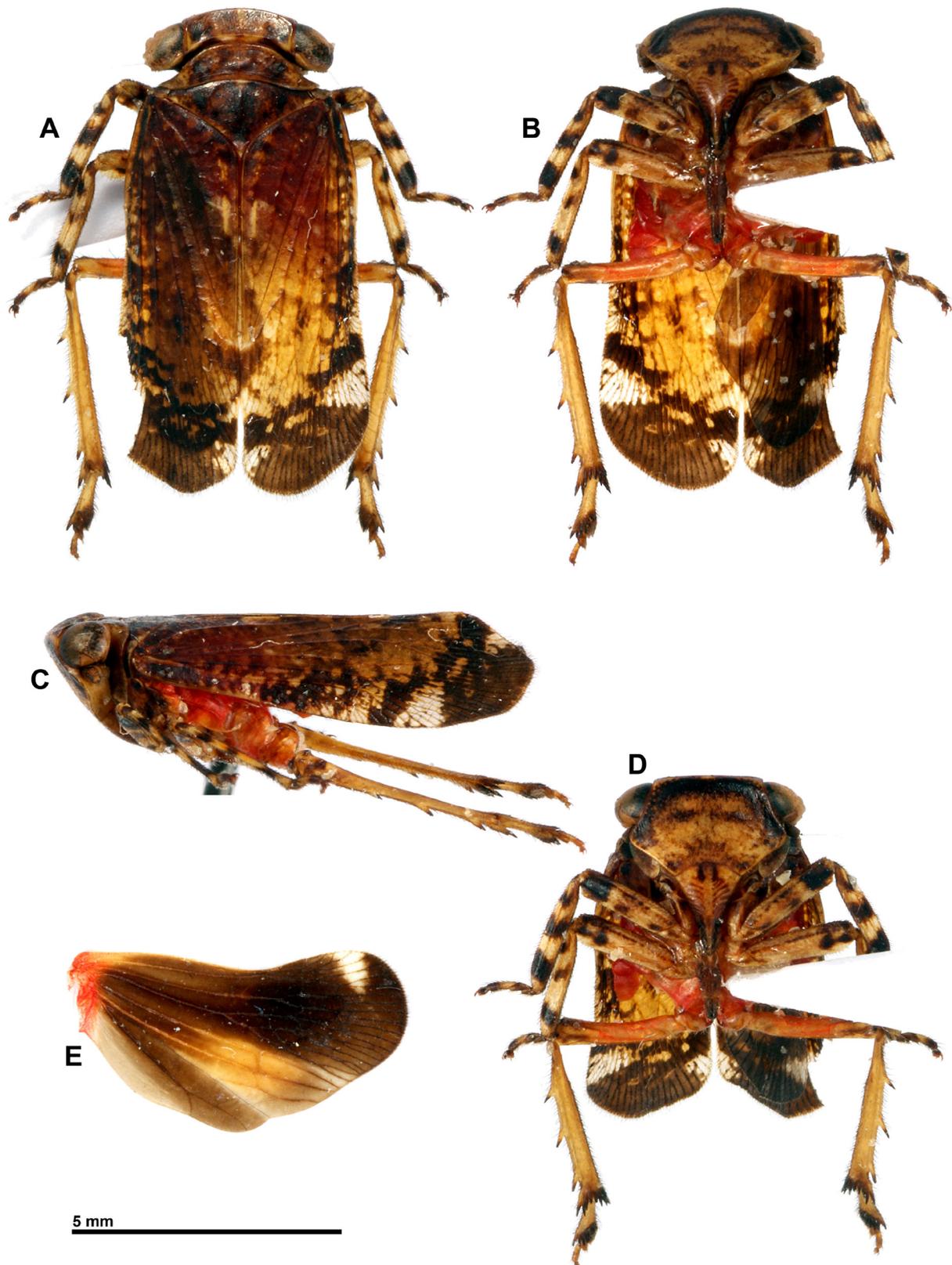


Fig. 4. *Olonia bourgoini* sp. nov., holotype, ♂. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

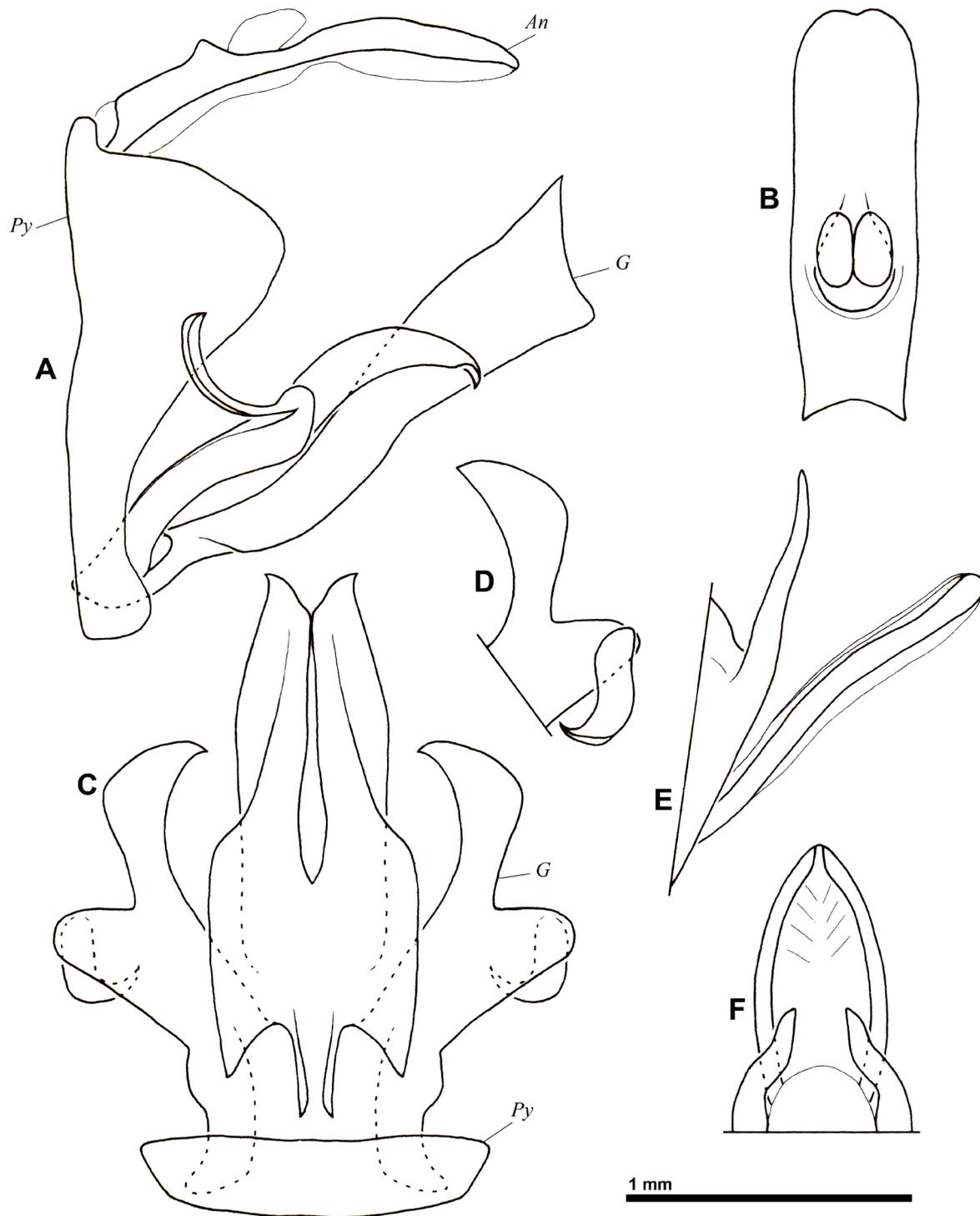


Fig. 5. *Olonia bourgoini* sp. nov., holotype, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.



Fig. 6. *Olonia bourgoini* sp. nov., paratype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

MALE GENITALIA (Fig. 5). Posterior margin of pygofer in lateral view strongly sinuate, strongly roundly projecting at dorsal $\frac{1}{3}$, narrow ventrally (Fig. 5A, C). Anal tube oblong, 3.15 times as long as broad, slightly curved ventrally in lateral view; lateral margins subparallel and slightly curved ventrally; apical margin emarginate in middle (Fig. 5A–B). Gonostyli fused to nearly half length of centroventral part and projecting posterodorsally (Fig. 5A, C). Centroventral part dorsoventrally flattened basally, progressively twisted and laminate towards apex, broader apically in lateral view, with apical margin concave, apicodorsal angle pointed and apicoventral angle angularly rounded (Fig. 5A, C). Laterodorsal part of gonostyli curved centroventrally, falcate and slightly concave ventrally, with lateral process rather broad and shorter than spoon-shaped process (Fig. 5A, C–D). Dorsal portion of phallobase with elongate process on each side, pointing dorsally and slightly curved internally at base (Fig. 5E). Ventral portion of phallobase along lateral margin of phallus, elongate, curved internally and nearly merging together apically, directed posterodorsally (Fig. 5E–F). Phallus dorsoventrally flattened, lanceolate in dorsal view (Fig. 5E–F).

Female

Similar to male but with frons entirely brown and with white markings of tegmina smaller (Fig. 6).

Distribution and biology

This species is currently recorded from a single location in tropical Queensland, near Chillagoe, in the Einasleigh Upland Savanna bioregion (Fig. 3). The specimens were collected in March.

Olonia danielsi sp. nov.

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Figs 3, 7–8

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings without orange marking (Fig. 7E)
- (2) pro- and mesofemora and tibiae largely dark brown (Fig. 7A–D)
- (3) anal tube of male narrow and parallel-sided on basal third, then subrhomboid (Fig. 8D)
- (4) centroventral part of gonostyli with long laminate process hooked apically (Fig. 8A, C)
- (5) laterodorsal part of gonostyli strongly curved before apex and pointing laterodorsally (Fig. 8A, C–D)
- (6) rather large size: 10 mm

Etymology

This species is dedicated to Greg Daniels who collected the holotype, together with his wife Alice.

Type material

Holotype

AUSTRALIA • ♂; Queensland, 3 km NW of Archer River crossing; 13°24' S, 142°55' E; “3km NW Archer Riv x-ing, Qld 13°24'S 142°55'E, 10 Apr 1989, G. and A. Daniels”, “QM Reg. #43783”; QM.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 1): 10.0 mm; BV/LV = 4.7; BF/LF = 1.95; LP+LM/BT = 0.72; Ltg/BTg = 2.3; LW/BW = 1.86.

Male

HEAD (Fig. 7A–D). Vertex concave with anterior and posterior margins parallel, curved; variegated brown and yellow-brown with darker marking at lateral angles. Frons brown variegated with yellowish,

with 2 irregular black markings on disc. Clypeus elongate, brown variegated with yellowish, and darker apically. Genae pale yellowish, with brown markings around eyes and under antennae. Labium dark brown, reaching metacoxae. Antennae dark brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 7A–D). Pronotum brown variegated with yellowish and with 2 small yellowish spots near lateral angles; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax brown. Mesonotum brown variegated with yellowish; broad, irregular, black marking along median carina; disc slightly transversely wrinkled; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae brown, paler dorsally.

TEGMINA (Fig. 7A, C). Brown variegated with yellowish and black; large, irregular, apical black marking; pale yellow, oblique, sinuate line on vein A1 at half length of clavus; large triangular white marking on costal margin on nodal line, slightly variegated with brown; white marking at apicosutural angle, slightly variegated with brown. Maximum breadth at nodal line; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 7E). Black-brown, paler near base along costal and anal margins and along anal fold; elongate, transverse, rather small white marking at apicocostal angle, extending on 3 cells. Margin of anal area sinuate; sutural margin with 2 clefts, cubital one slightly marked.

LEGS (Fig. 7A–D). Pro- and mesocoxae yellowish brown. Pro- and mesofemora dark brown with 2 obsolete yellowish rings. Pro- and mesotibiae dark brown with 2 slightly marked yellowish rings. Pro- and mesotarsi brown with basal half of third tarsomere paler. Metacoxae red. Metafemora reddish yellow with apex brown. Metatibiae brown with 3 lateral spines yellowish basally and 8 apical black-brown spines. Metatarsi brown with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 8). Posterior margin of pygofer in lateral view strongly sinuate, strongly roundly projecting at dorsal $\frac{1}{3}$, rather broad ventrally (Fig. 8A, C). Anal tube elongate, 2.05 times as long as broad, slightly curved ventrally in lateral view; lateral margins parallel on basal $\frac{1}{3}$, rhomboid on apical $\frac{2}{3}$ with posterior margin rounded (Fig. 8A–B). Gonostyli fused basally and coalescent on $\frac{4}{5}$ of length; diverging apically (Fig. 8C). Centroventral part elongate, laterally compressed and ending in a strong hook pointing dorsally (Fig. 8A, C). Laterodorsal part of gonostyli strongly sinuate in dorsal view with apical part strongly curved and directed dorsolaterally; lateral process in more dorsal position, slightly curved ventrally near apex and slightly directed anteriorly, longer than spoon-shaped process (Fig. 8A, C–D). Dorsal portion of phallobase high basally in lateral view, directed posteroventrally and with apical hook strongly curved dorsally (Fig. 8E). Ventral portion of phallobase along lateral margin of phallus, weakly sclerified, laterally flattened, elongate (Fig. 8E–F). Phallus dorsoventrally flattened, narrowing, bifid and slightly diverging apically in dorsal view (Fig. 8E–F).

Female

Unknown.

Distribution and biology

This species is currently recorded from a single location on Cape York Peninsula, Queensland (Fig. 3), in the Cape York Peninsula Tropical Savanna bioregion. The specimen was collected in April.

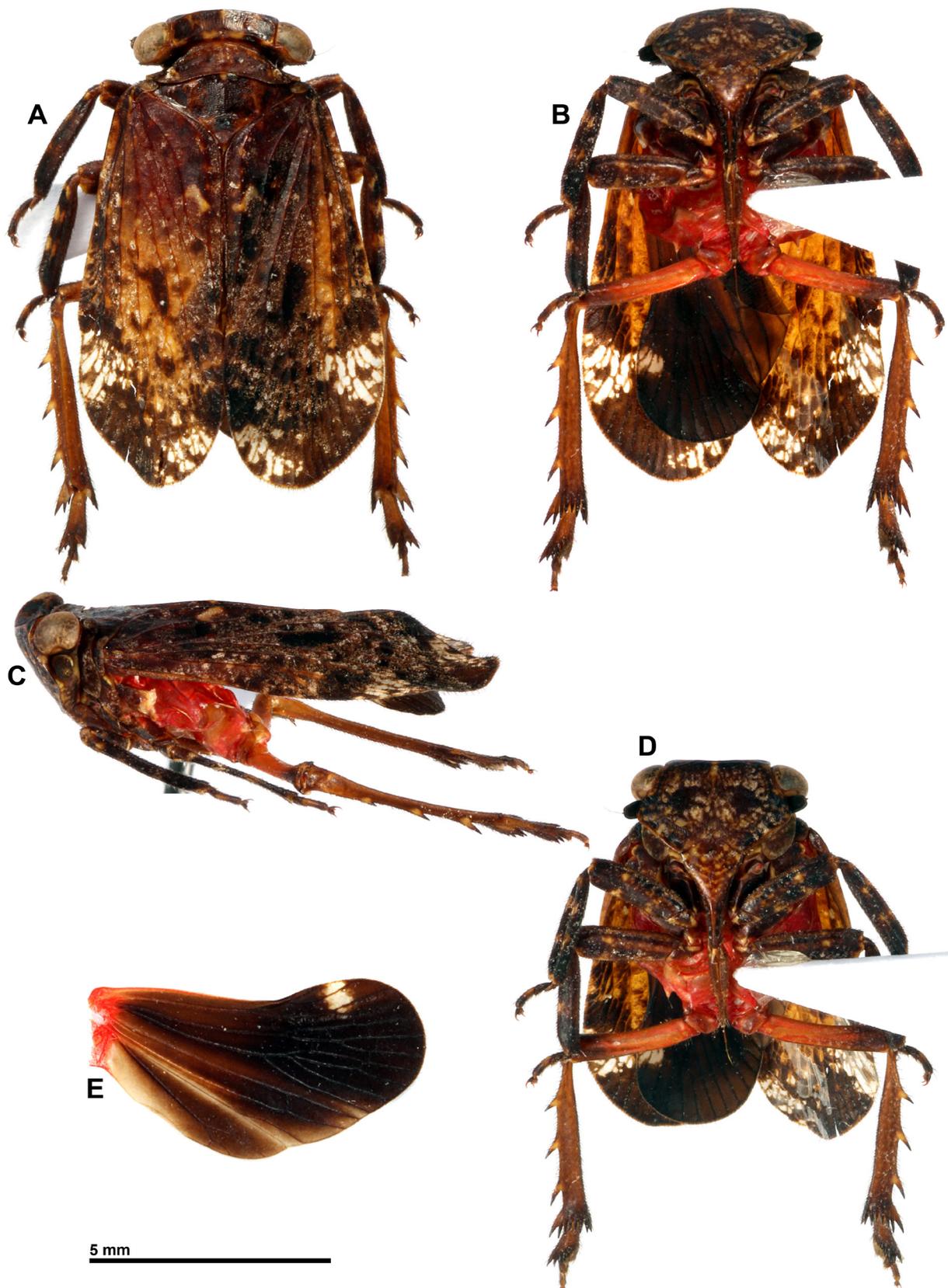


Fig. 7. *Olonia danielsi* sp. nov., holotype, ♂. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

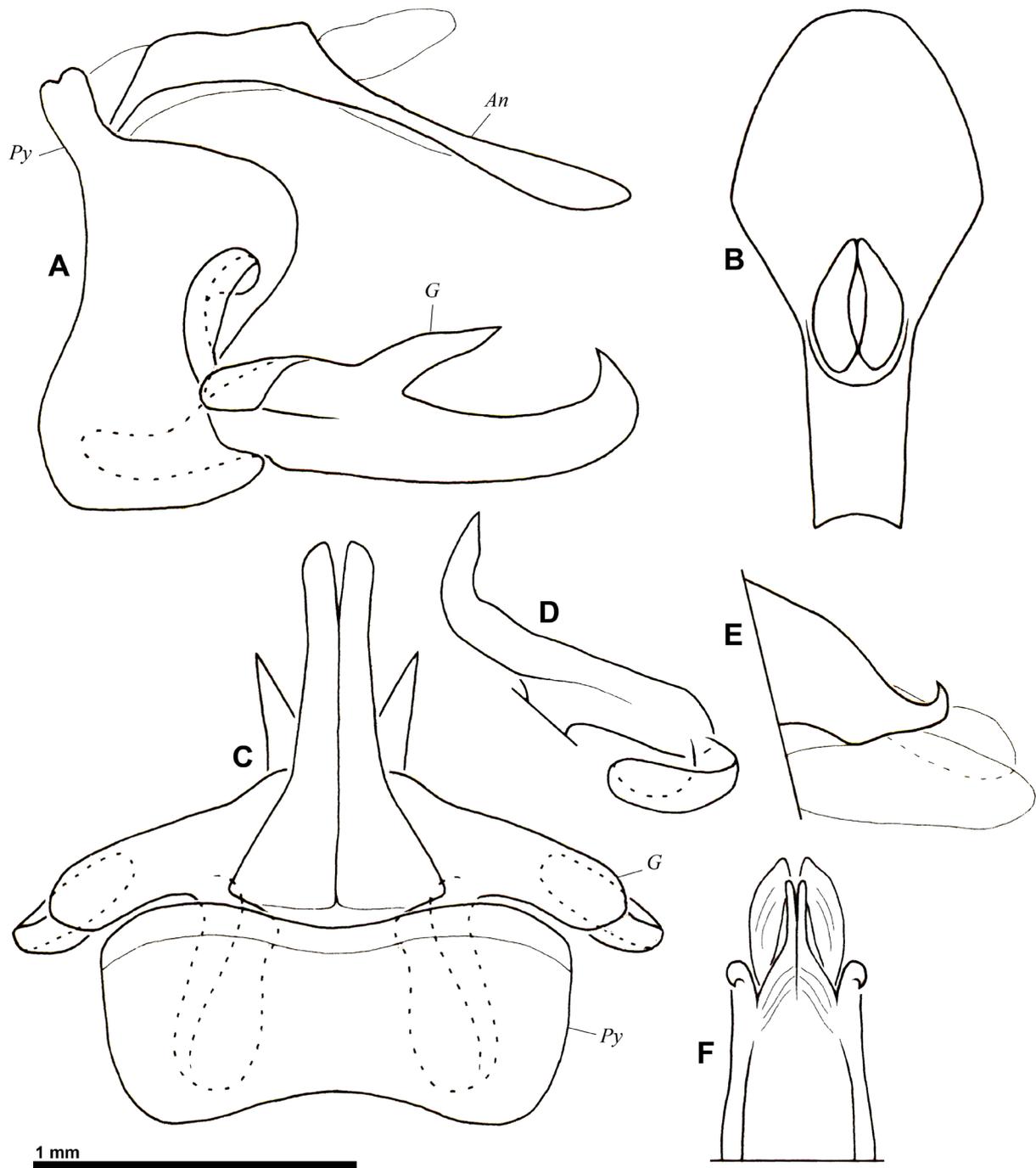


Fig. 8. *Olonia danielsi* sp. nov., holotype, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

Olonia guillaumei sp. nov.

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Figs 3, 9–11

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings with conspicuous orange marking (Figs 9E, 11E)
- (2) pro- and mesofemora and tibiae largely brown (Figs 9A–D, 11A–D)
- (3) anal tube of male oblong, with posterior margin rounded (Fig. 10B)
- (4) centroventral part of gonostyli with long laminate process (Fig. 10A, C)
- (5) laterodorsal part of gonostyli with hooked process directed lateroventrally (Fig. 10A, C–D)
- (6) rather large size: 9.5–10 mm

Etymology

This species is named after my son, Guillaume.

Type material

Holotype

AUSTRALIA • ♂; NE Queensland, Bakers Blue Mt, 17 km W of Mt Molloy; [16°42' S, 145°09' E]; "N.E. QLD, Bakers Blue Mt, 17 km W Mt Molloy, 800m, open For. 12.ix.1981, G. Monteith & D. Cook"; QM.

Paratypes

AUSTRALIA • 1 ♂; [17°20'26" S, 144°40'37" E]; "Almaden, Chillagoe Dist., N. Q. Jan. 1932, W.D. Campbell", "K64979"; AMS • 1 ♀; "Almaden, Chillagoe Dist., N. Q. March 1929, W.D. Campbell", "K58965"; AMS • 1 ♂; "Almaden, Chillagoe Dist., N. Q. June 1927, W.D. Campbell", "K56631."; RBINS • 1 ♂; [17°23' S, 145°23' E]; "Herberton, Q., 23 Aug 1970, F. McDonald", "MJF Collection, MJF003398"; ASCU.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 3): 9.9 mm (9.6–10.1); BV/LV = 4.15; BF/LF = 1.83; LP+LM/BT = 0.66; Ltg/BTg = 2.5; LW/BW = 1.80.

Male

HEAD (Fig. 9A–D). Vertex concave with anterior and posterior margins parallel, curved; yellowish variegated with brown and with black marking at lateral angles. Frons black-brown with mediobasal irregular marking yellowish variegated with brown. Clypeus elongate, brown with 2 short, black, longitudinal lines at base, with oblique yellowish lines on sides and with apex darker. Genae pale yellowish, slightly darker at place around eyes and with brown marking under antennae. Labium black-brown, reaching metacoxae. Antennae dark brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 9A–D). Pronotum brown variegated with yellowish, with blackish marking and with yellowish spot at lateral angles; slightly transversely wrinkled posteriorly; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax dark brown. Mesonotum black-brown with reddish markings along posterior margin, median yellowish spot along anterior margin and at apex of scutellum; slightly transversely wrinkled; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae brown.

TEGMINA (Fig. 9A, C). Brown variegated with yellowish and black; irregular transverse row of whitish spots at basal 1/3; large triangular white marking on costal margin on nodal line; white spot at apicosutural angle. Maximum breadth at nodal line; costal margin sinuate; apical margin obliquely rounded.

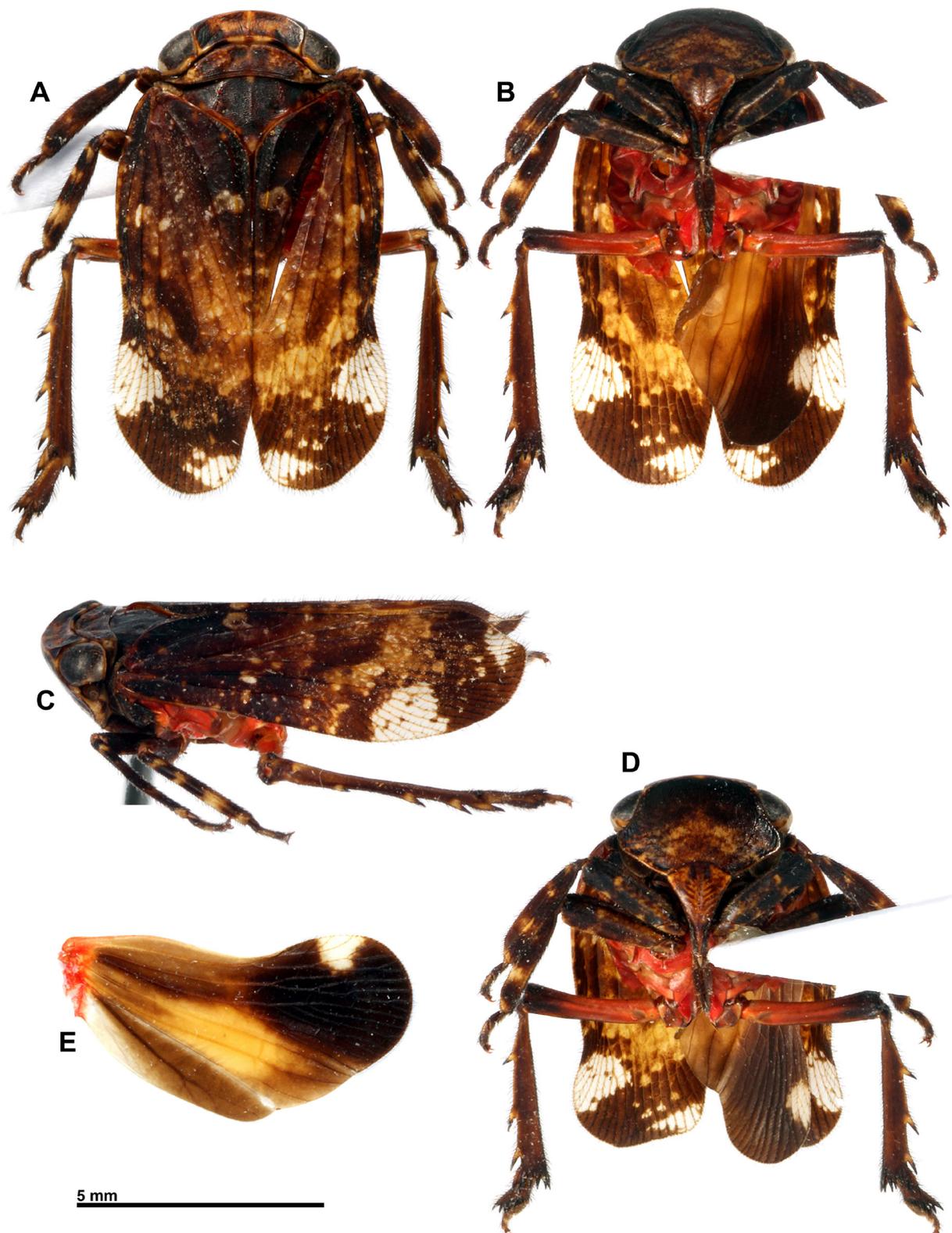


Fig. 9. *Olonia guillaumei* sp. nov., holotype, ♂. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Posterior wing.

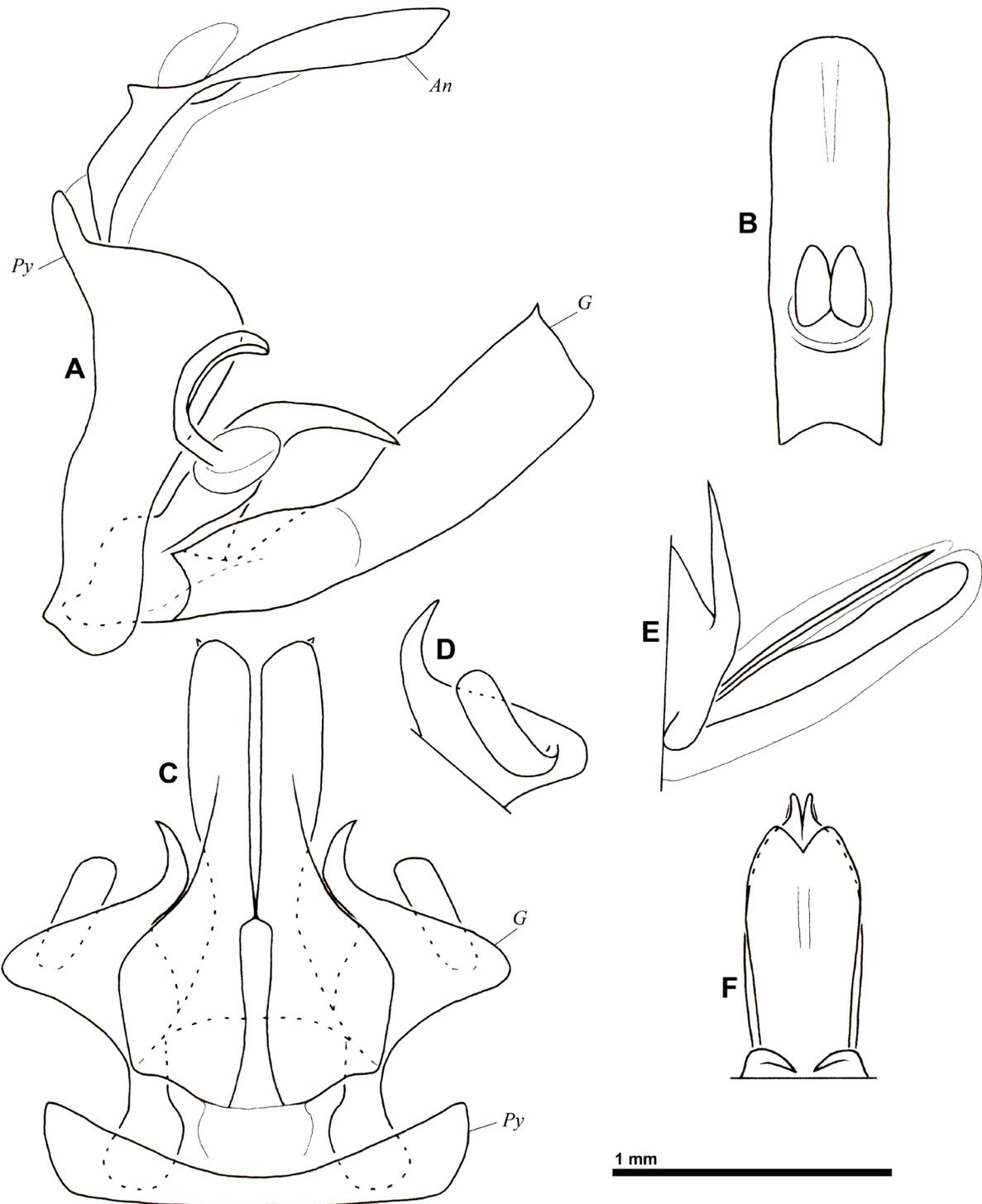


Fig. 10. *Olonia guillaumei* sp. nov., holotype, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

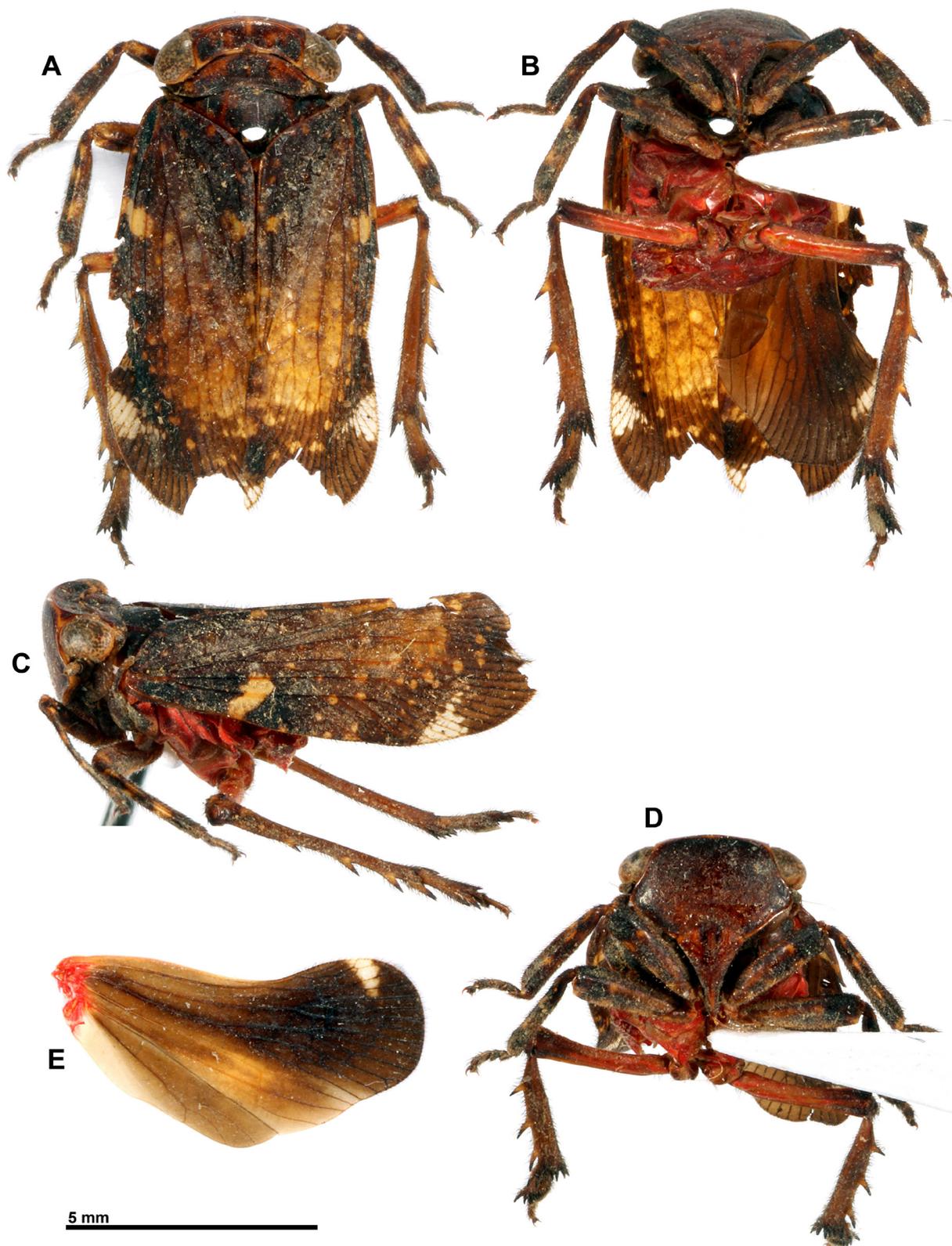


Fig. 11. *Olonia guillaumei* sp. nov., paratype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

POSTERIOR WINGS (Fig. 9E). Brown with apical $\frac{1}{3}$ black; transverse, broad, subtriangular white marking at apicocostal angle, extending on 4 cells; large, elongate orange marking between anal fold and vein CuA at basal $\frac{1}{3}$, not reaching margin. Margin of anal area slightly rounded; sutural margin with 2 clefts, cubital one slightly marked.

LEGS (Fig. 9A–D). Pro- and mesocoxae brown. Pro- and mesofemora black-brown with 2 obsolete yellowish rings. Pro- and mesotibiae dark brown with 3 obsolete ring-shaped yellowish markings, larger one near apex. Pro- and mesotarsi dark brown with basal half of third tarsomere paler. Metacoxae reddish brown; metafemora red with apex dark brown. Metatibiae brown, with 3 lateral spines yellowish basally and 8 apical, black-brown spines. Metatarsi dark brown with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 10). Posterior margin of pygofer in lateral view strongly sinuate, strongly roundly projecting at dorsal $\frac{1}{3}$, narrow ventrally (Fig. 10A, C). Anal tube oblong, 3.3 times as long as broad, slightly curved ventrally in lateral view; lateral margins subparallel and slightly curved ventrally on apical $\frac{2}{3}$; apical margin rounded (Fig. 10A–B). Gonostyli fused to nearly half length of centroventral part and projecting posterodorsally (Fig. 10A, C). Centroventral part dorsoventrally flattened basally, progressively twisted and laminate towards apex, broader apically in lateral view, with apical margin slightly sinuate, apicodorsal angle pointed and apicoventral angle angularly rounded (Fig. 10A, C). Laterodorsal part of gonostyli curved lateroventrally, hooked, with lateral process rather broad and about as long as spoon-shaped process (Fig. 10A, C–D). Dorsal portion of phallobase with elongate process on each side, pointing dorsally and slightly internally (Fig. 10E–F). Ventral portion of phallobase along lateral margin of phallus on basal half, then under phallus, narrowing and bifid apically, directed posterodorsally (Fig. 10E–F). Phallus dorsoventrally flattened, elongate, with apical margin strongly emarginate in dorsal view (Fig. 10E–F).

Female

Similar to male, but with frons brown turning to reddish-brown medioventrally; legs paler with yellowish rings more developed; posterior wings less contrasted, with apicocostal white marking narrower and orange marking less developed (Fig. 11).

Distribution and biology

This species is currently recorded from a small area west of Mareeba in North Queensland (Fig. 3), in the Einasleigh Upland Savanna bioregion. The specimens were collected in January, March, June, August and November; hence, the species may be present all year long.

Olonia hochae sp. nov.

[urn:lsid:zoobank.org:act:AAD09D97-D6FD-438D-BBF4-CE7326D8AD08](https://zoobank.org/urn:lsid:zoobank.org:act:AAD09D97-D6FD-438D-BBF4-CE7326D8AD08)

Figs 3, 12–14

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings without orange marking (Figs 12E, 14E)
- (2) pro- and mesofemora and -tibiae largely black-brown (Figs 12A–D, 14A–D)
- (3) anal tube of male oblong, with posterior margin narrowly rounded (Fig. 13B)
- (4) centroventral part of gonostyli with long laminate process concave on ventral margin (Fig. 13A, C)
- (5) laterodorsal part of gonostyli with spinose process curved ventrally (Fig. 13A, C–D)
- (6) rather large size: 9–10 mm

Etymology

This species is dedicated to Dr Hannelore Hoch, who collected the paratypes series together with Dr Manfred Asche. But ladies first!

Type material

Holotype

AUSTRALIA • ♂; Queensland, Undara National Park; 18°15' S, 144°41' E; “QLD: 18°15'Sx144°41'E, Undara NP, 17 ft Bore. 8 Dec 2002–8 Feb 2003, G. Monteith, vine scrub. fit intercept. 11252”, “QM-T244703”; QM.

Paratypes

AUSTRALIA • 4 ♂♂, 4 ♀♀; “Australia: Queensland, Undara Lava flow: betw. Mt. Garnet & Mt. Surprise, Yarramulla Sta.: surface around Pinwill’s Cave”, “18.i.1989, Au 24, M. Asche & H. Hoch”; ZMHB • 1 ♂, 1 ♀; same collection data as for preceding; RBINS.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 4): 9.7 mm (9.4–9.9); ♀ (n = 5): 10.4 mm (10.0–10.8); BV/LV = 3.9; BF/LF = 1.75; LP+LM/BT = 0.68; Ltg/BTg = 2.40–2.45; LW/BW = 1.78.

Male

HEAD (Fig. 12A–D). Vertex concave, with anterior and posterior margins parallel, curved; dark brown variegated with yellowish in middle. Frons black-brown, slightly tinged with reddish medioventrally. Clypeus elongate, black-brown with 2 short, black, longitudinal lines at base, with oblique reddish lines on sides and with apex darker. Genae brown with yellowish markings along anterior margin. Labium black, reaching metacoxae. Antennae black; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 12A–D). Pronotum dark brown with some yellowish markings; slightly transversely wrinkled posteriorly; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax dark brown. Mesonotum black-brown with reddish markings along posterior margin, median yellowish spot along anterior margin and at apex of scutellum; slightly wrinkled; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae brown.

TEGMINA (Fig. 12A, C). Dark brown with small yellowish or reddish spots; bigger, yellowish, slightly transverse spot on middle of clavus; large triangular white marking on costal margin on nodal line; white spot at apicosutural angle. Maximum breadth at nodal line; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 12E). Brown with apical $\frac{1}{3}$ black-brown; transverse, broad, subtriangular white marking at apicocostal angle, extending on 6 cells. Margin of anal area sinuate; sutural margin with 2 clefts, cubital one slightly marked.

LEGS (Fig. 12A–D). Pro- and mesocoxae dark brown. Pro- and mesofemora black-brown with reddish spots marking 2 obsolete rings. Pro- and mesotibiae black-brown with 3 obsolete ring-shaped reddish or yellowish markings, larger one near apex. Pro- and mesotarsi dark brown. Metacoxae reddish brown; metafemora red with apex dark brown. Metatibiae brown, with 3 lateral spines yellowish basally and 8 apical, black-brown spines. Metatarsi brown, with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 13). Posterior margin of pygofer in lateral view strongly sinuate, strongly roundly projecting at dorsal $\frac{1}{3}$, narrow ventrally (Fig. 13A, C). Anal tube oblong, 3.6 times as long as broad, slightly curved ventrally in lateral view; slightly constricted on basal $\frac{1}{3}$ and narrowing towards apex in dorsal view; lateral margins sinuate and slightly curved ventrally on apical $\frac{2}{3}$; apical margin narrowly rounded (Fig. 13A–B). Gonostyli fused to slightly further than basal third of centroventral part and projecting posterodorsally (Fig. 13A, C). Centroventral part dorsoventrally flattened basally,

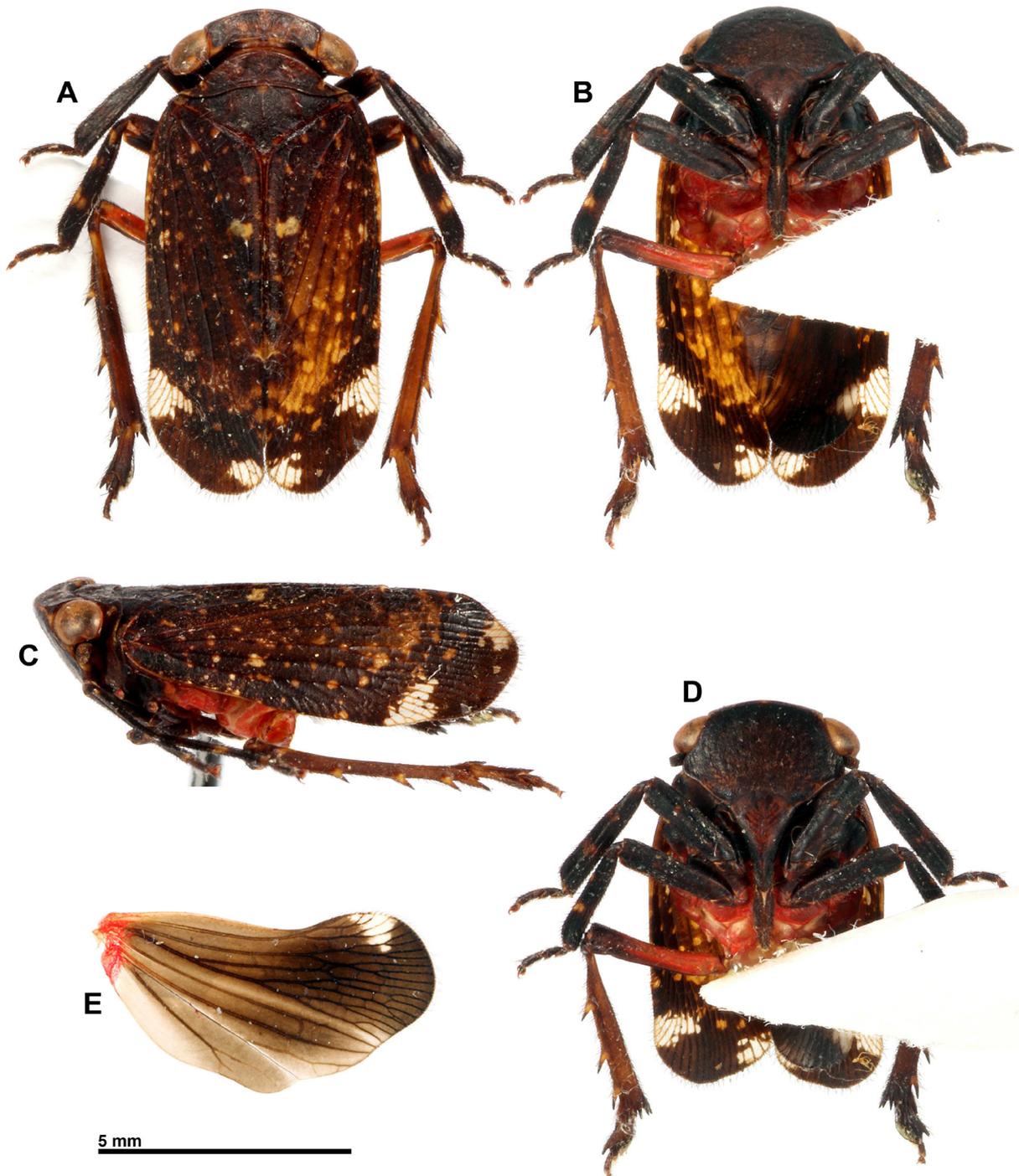


Fig. 12. *Olonia hochae* sp. nov., paratype, ♂. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Posterior wing.

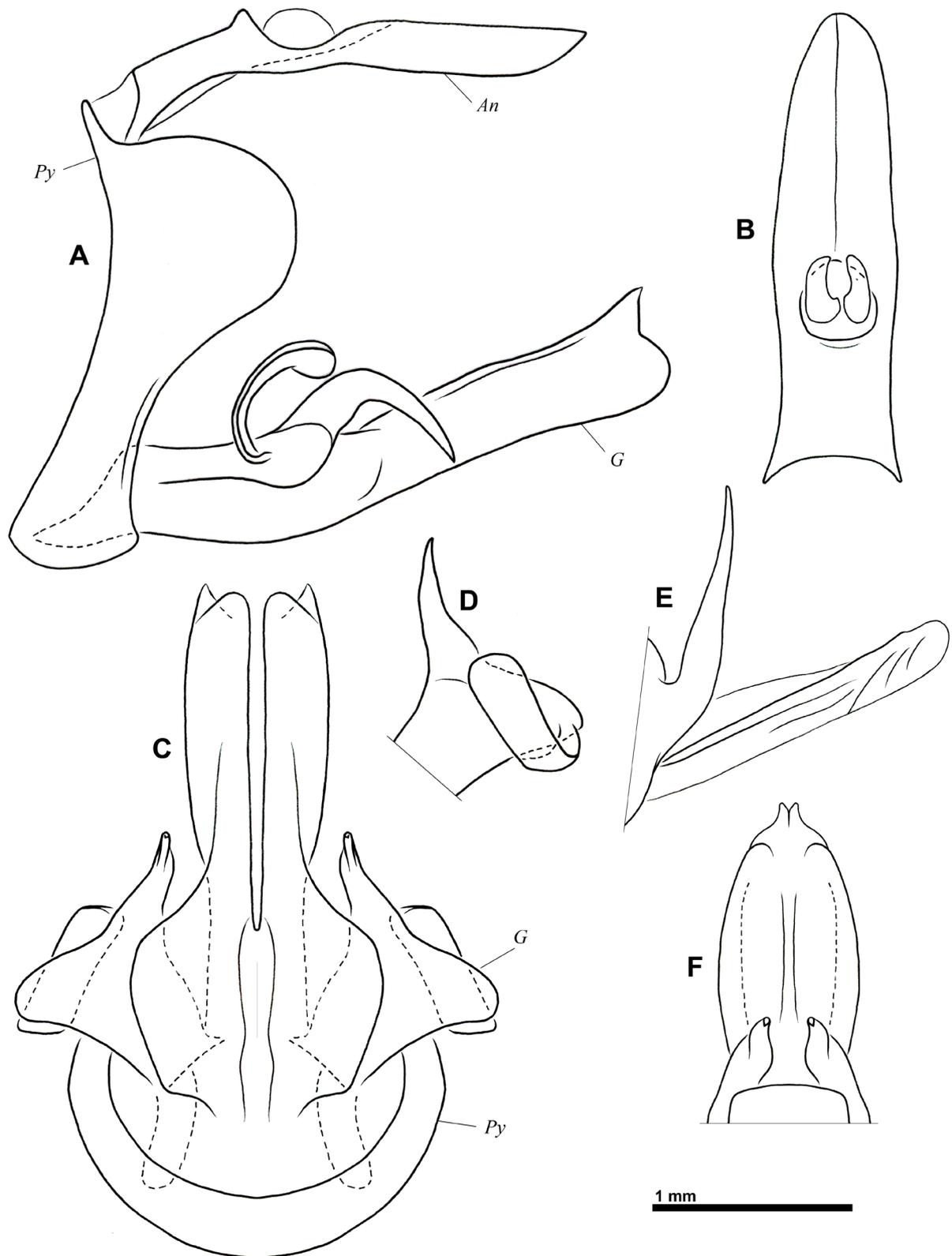


Fig. 13. *Olonia hochae* sp. nov., holotype, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

progressively twisted and laminate towards apex and with ventral margin concave in lateral view; slightly broader apically in lateral view with apical margin strongly sinuate, apicodorsal angle pointed and apicoventral angle rounded (Fig. 13A, C). Laterodorsal part of gonostyli with ventrally strongly curved, hooked process, with lateral process rather broad and about as long as spoon-shaped process (Fig. 13A, C–D). Dorsal portion of phallobase with strongly elongated process on each side, pointing dorsally and slightly internally (Fig. 13E–F). Ventral portion of phallobase along lateral margin of phallus on basal two thirds, then under phallus, narrowing and bifid apically, directed posterodorsally (Fig. 13E–F). Phallus dorsoventrally flattened, elongate, with apical margin emarginate in dorsal view (Fig. 13E–F).

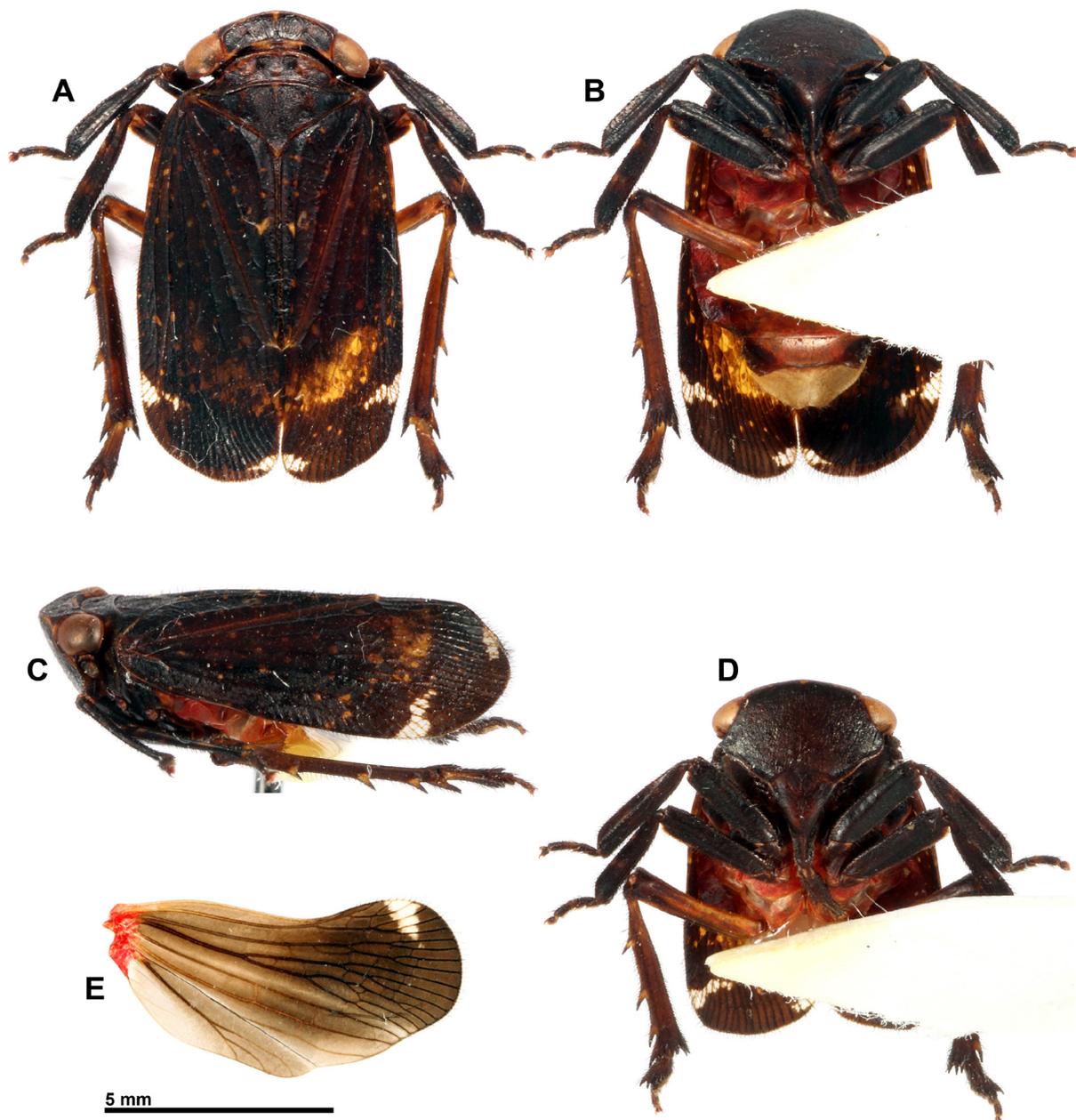


Fig. 14. *Olonia hochae* sp. nov., paratype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

Female

Similar to male, but darker; frons entirely black-brown; markings on pro- and mesofemora nearly absent; markings on pro- and mesotibiae reduced; white markings on tegmina smaller (Fig. 14).

Distribution and biology

This species is currently known from a series of ten specimens collected near Undara Lava Tubes in North Queensland (Fig. 3), in the Einasleigh Upland Savanna bioregion. All specimens were collected in January, on the same day; hence, the species is probably not scarce in its habitat.

Olonia marginata Distant, 1906 Figs 3, 15–25

Olonia marginata Distant, 1906: 206 (described).

Olonia marginata – Kirkaldy 1907: 105 (listed from Queensland). — Metcalf 1956: 64 (catalogued).

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings without orange marking (Figs 16A, 20A)
- (2) pro- and mesofemora and -tibiae largely black-brown (Figs 16A–D, 20A–D)
- (3) anal tube of male spatulate, constricted at basal $\frac{1}{3}$ (Fig. 19B)
- (4) centroventral part of gonostyli with narrow and strongly elongated process that is sinuate basally and hooked apically (Fig. 19A, C)
- (5) laterodorsal part of gonostyli with narrow and strongly elongated process that is strongly curved posteroventrally (Fig. 19A, C–D)
- (6) rather small size: 6–8 mm

Etymology

The species epithet *marginatus* (adjective, Latin) means ‘marginated’, ‘bordered’. It refers to the darkened costal and apical margins of the tegmina.

Material examined

Lectotype

AUSTRALIA • 1 ♀; Queensland; “Type”, “Type”, “*Olonia marginata*, type, Dist.”, “Queensland, F.P. Dodd, 1902 – 319”; here designated to provide a reference standard for this species; BMNH (Fig. 15).

Paralectotypes

AUSTRALIA • 2 ♀♀; “Queensland, F.P. Dodd, 1902 – 319”; BMNH.

Note

In the collections of BMNH, there are 3 additional specimens identified by Distant but which are not part of the type series: 1 ♀, “*Olonia marginata* Dist.”, “Townsville [19°25'26" S, 146°56'44" E] (Dodd)”, “B”, “Distant Coll. 1911-383”; 2 ♂♂, “Queensland, F.P. Dodd, Brit. Mus., 1907—54”.

These specimens are particularly interesting because (1) they are from the same collector as the type series; (2) they provide Townsville as the location for the type series, where the collector, Frederick Parkhurst Dodd (1861–1937), lived at the time before moving to Kuranda and (3) there are two males, which allows a characterization of the species based on male genitalia. These males are used here as a reference for the recognition of this species.

Additional material

AUSTRALIA • 3 ♂♂, 1 ♀; Townsville; Jan. 1945; B. Malkin leg.; USNM • 1 ♂, 1 ♀; same collection data as for preceding; RBINS • 1 ♀; Townsville; 2 Mar. 1956; I. Sutherland leg.; QM • 1 ♂; Townsville, Hervey Range; 19°22'42" S, 146°31'29" E; 22 Sep. 1981; D.C. Geijskes leg.; RMNH • 1 ♂; Townsville, Heatley; 19°17'35" S, 146°45'09" E; 4 Aug. 1976; D.C. and R. Geijskes leg.; RMNH • 1 ♂; Townsville; 7 Apr. 1977; Brown leg.; QPIM • 1 ♂; Pallarenda Point near Townsville; 19°11'20" S, 146°46'25" E, 26 Jan. 1965; E.C. Dahms leg.; open forest, by net; QM • 1 ♂; Townsville, Hyde Park Shopping Centre; 19°16'46" S, 146°47'49" E; 25 Feb. 1992; L.M. Brown leg.; on *Melaleuca quinquenervia*; ASCU • 1 ♂; Macrossan; 20°00'18" S, 146°26'44" E; 15 Apr. 1958; K.L. Hartley leg.; ANIC • 1 ♂; near Ingham; 18°39'00" S, 146°10'00" E; 13 Mar. 1961; R. Straatman leg.; on mangrove forest beach; ANIC • 2 ♂♂, 1 ♀; 10 mi. SSE of Collinsville [coordinates of Collinsville: 20°33'08" S, 147°50'38" E]; 14 Sep. 1950; E.F. Riek leg.; ANIC • 5 ♂♂, 1 ♀; 30 mi. S of Ayr; [coordinates of Ayr: 19°34'33" S, 147°24'18" E]; 9 Sep. 1950; E.F. Riek leg.; ANIC • 2 ♂♂, 1 ♀; same collection data as for preceding; RBINS • 1 ♂; 35 mi. SE of Ayr; 3 Oct. 1950; E.F. Riek leg.; ANIC • 1 ♀; 40 mi. SW of Ayr; 7 Oct. 1950; E.F. Riek leg.; ANIC • 1 ♀; 40 mi. S of Ayr; 10 Sep. 1950; E.F. Riek leg.; ANIC • 1 ♀; 50 mi. S of Ayr; 11 Sep. 1950; E.F. Riek leg.; ANIC • 1 ♂; 35 mi. SW of Ayr; 6 Oct. 1950; E.F. Riek leg.; ANIC • 1 ♂, 1 ♀; 60 mi. SW of Ayr; 8 Oct. 1950; E.F. Riek leg.; ANIC • 1 ♂, 1 ♀; same collection data as for preceding; RBINS • 1 ♀; 35 mi. NW of Bowen; [coordinates of Bowen: 20°00'33" S, 148°13'46" E]; 2 Oct. 1950; E.F. Riek leg.; ANIC • 2 ♂♂; 25 mi. NW of Bowen; 1 Oct. 1950; E.F. Riek leg.; ANIC • 1 ♂; 10 mi. S of Bowen; 26 Sep. 1950; E.F. Riek leg.; ANIC • 4 ♀♀; Townsville; alt. sea level; 13 Jan. 1962; E.S. Ross and D.Q. Cavagnaro leg.; CAS • 1 ♀; same collection data as for preceding; RBINS • 1 ♂, 1 ♀; Majors Creek; 19°37'36" S, 146°57'28" E; 27 Oct. 2006; G. Cocks leg.; on grapes; RBINS • 2 ♂♂, 2 ♀♀; Townsville; 12 Oct. 1983; M.E. Irwin and E.I. Schlinger leg.; INHS • 1 ♂, 1 ♀; same collection data as for preceding; RBINS • 1 ♀; 70 km NW of Townsville; 9 Nov. 1990; W.F. Chamberlain leg.; TAMU • 1 ♂, 1 ♀; Rollingsstone; 19°02'40" S, 146°23'01" E; 19 Apr. 1998; L.J. Cookson leg.; MVMA • 1 ♂; Surveyor Creek; 20°43'59" S, 148°36'00" E; 30 Jan. 1991; L.J. Cookson leg.; MVMA • 1 ♂, 1 ♀; Bowen, Queens Bay; 19°58'43" S, 148°14'50" E; 7 Jul. 1971; Z. Liepa leg.; ANIC • 1 ♂; 30 km S of Ingham; 10 Nov. 1990; G.M. Chamberlain leg.; TAMU • 1 ♂; Clare; 19°49'43" S, 147°11'46" E; 7 Apr. 1951; W.A.S. leg.; from sweeping in jute; QDPI • 1 ♂; Saltwater Creek near Toomulla; 19°05'56.37" S, 146°27'46.69" E; 20 Apr. 1998; L.J. Cookson leg.; MVMA • 1 ♂; 32 km S of Ayr; 6 Feb. 1964; J. Sedlacek leg.; BPBM • 1 ♂; Gumlu, 48 km SE of Ayr; 19°54'09" S, 147°35'30" E; 6 Feb. 1964; J. Sedlacek leg.; BPBM • 1 ♂; Herbert River; 18°31'59" S, 146°18'00" E; 1919; J.F. Illingworth leg.; on (sugar)cane; BPBM • 1 ♂, 2 ♀♀; Townsville, Strand Beach, Northern End; [coordinates of Strand Beach: 19°14'23" S, 146°48'24" E]; 15 Mar. 2016; V. Ryland leg.; on *Ipomoea pes-caprae*; RBINS • 2 ♀♀; same collection data as preceding; 19 Feb. 2016; QM • 1 ♀; same collection data as preceding; 25 Feb. 2016; QM • 1 ♂; Townsville, port wall; [coordinates of Townsville port: 19°15'17" S, 146°50'11" E]; 15 Mar. 2016; V. Ryland leg.; on *Ipomoea pes-caprae*; RBINS • 1 ♂; Townsville, small boat harbour wall; [coordinates of Townsville small boat harbour: 19°15'12" S, 146°49'24" E]; 19 Mar. 2016; V. Ryland leg.; on *Ipomoea pes-caprae*; RBINS • 1 ♂; same collection data as for preceding; QM • 1 ♂; Townsville, Bushland Beach; 19°11'19" S, 146°40'40" E; 4 Apr. 2016; V. Ryland leg.; on *Ipomoea pes-caprae*; RBINS • 1 ♂, 1 ♀; same collection data as for preceding; QM • 2 ♂♂; Magnetic Island, Nelly Bay Beach; [coordinates of Nelly Bay: 19°09'53" S, 146°51'01" E]; 11 Feb. 2016; V. Ryland leg.; on *Ipomoea pes-caprae*; RBINS • 1 ♂, 1 ♀; same collection data as for preceding; QM • 1 ♀; Magnetic Island, Geoffrey Bay Beach; [coordinates of Geoffrey Bay: 19°09'17" S, 146°51'54" E]; 10 Feb. 2016; V. Ryland leg.; on *Ipomoea pes-caprae*; RBINS • 1 ♂; same collection data as for preceding; QM • 1 ♂, 1 ♀; same collection data as preceding; 24 Feb. 2016; RBINS • 2 ♂♂, 1 ♀; same collection data as for preceding; QM • 1 ♂, 1 ♀; Magnetic Island, Picnic Bay Beach; [coordinates of Picnic Bay: 19°10'54" S, 146°50'27" E]; 5 Feb. 2016; V. Ryland leg.; on *Ipomoea pes-caprae*; RBINS • 1 ♀; same collection data as for preceding; QM • 1 ♀; same collection data as preceding; 12 Feb. 2016; RBINS • 1 ♀; same collection data as preceding; 18 Feb. 2016; QM • 1 ♀; same collection data

as preceding; 27 Feb. 2016; RBINS • 1 ♂, 2 ♀♀; same collection data as for preceding; QM • 1 ♂; same collection data as preceding; 10 Jan. 2016; on *Eucalyptus* sapling; RBINS • 1 ♂; Magnetic Island; 19°08'18" S, 146°50'04" E; 30 Mar. 1929; QM • 2 ♂♂, 1 ♀; Magnetic Island; A.M. Lea leg.; SAM • 1 ♀; same collection data as preceding; G. F. Hill leg.; SAM • 1 ♂, 3 ♀♀; Magnetic Island; 8 Nov. 2006; J. Constant leg.; on *Canarium australianum* sapling; RBINS • 1 ♀; Magnetic Island; 1914; Dr Scharff leg.; BMNH • 1 ♂; Alligator Creek; 19°23'56"S 146°55'36"E; 1 Apr. 1934; H. Hacker leg.; USNM • 1 ♀; same collection data as preceding; 25 Apr. 1934; USNM.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 9): 7.1 mm (6.2–7.7); ♀ (n = 3): 7.7 mm (7.5–8.0); BV/LV = 4.5; BF/LF = 1.68; LP+LM/BT = 0.68; Ltg/BTg = 2.35; LW/BW = 1.86.

Male

HEAD (Figs 16A–D, 17A–D, 18A–D). Vertex concave, with anterior and posterior margins parallel, curved; brown with darker marking at lateral angles. Frons uniformly black-brown. Clypeus elongate, entirely black-brown. Genae black-brown with yellowish markings along anterior margin. Labium black, reaching metacoxae. Antennae black-brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Figs 16A–D, 17A–D, 18A–D). Pronotum brown with small yellowish spot on each side; slightly wrinkled; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax dark brown. Mesonotum brown variegated with blackish; yellowish spot on middle of anterior margin and at apex of scutellum; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae dark brown.

TEGMINA (Figs 16A, C, 17A, C, 18A, C). Brown slightly variegated with yellowish and black; often pale yellowish marking on vein A1 at midlength of clavus; marked with black along costal margin, more broadly so on posterior half, and along posterior margin (Figs 16A, C, 17A, C); sometimes a broad, whitish, transverse band at proximal $\frac{1}{3}$, bordered with black markings along costal margin (Fig. 18A, C). Darker, median, irregular marking at apical $\frac{2}{3}$; triangular white marking on costal margin on nodal line, sometimes reduced; no white spot at apicosutural angle. Costal and sutural margins subparallel; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Figs 16E, 17E, 18E). Dark brown, paler on anal area and with large blackish area reaching apical margin; elongate, transverse, subtriangular white marking at apicocostal angle, extending on 3–4 cells. Margin of anal area slightly sinuate; sutural margin with 1 cleft, cubital one not marked.

LEGS (Figs 16A–D, 17A–D, 18A–D). Pro- and mesocoxae black-brown. Pro- and mesofemora black-brown with reddish spots marking obsolete rings. Pro- and mesotibiae black-brown with reddish markings on 3 obsolete rings. Pro- and mesotarsi black-brown with basal half of third tarsomere paler. Metacoxae reddish; metafemora reddish with apex brown. Metatibiae brown, with 3 lateral spines paler basally and 8 apical black-brown spines. Metatarsi brown, with a ventral row of 6 black spines on first tarsomere.

ABDOMEN (Fig. 18F). Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 19). Posterior margin of pygofer in lateral view strongly sinuate, strongly roundly projecting at dorsal $\frac{1}{3}$, rather broad ventrally (Fig. 19A, C). Anal tube spatulate, 2.2 times as long as broad, laterally constricted at level of epiproct, slightly curved ventrally in lateral view; lateral margins slightly curved ventrally on apical $\frac{2}{3}$; apical margin rounded (Fig. 19 A–B). Gonostyli fused on basal

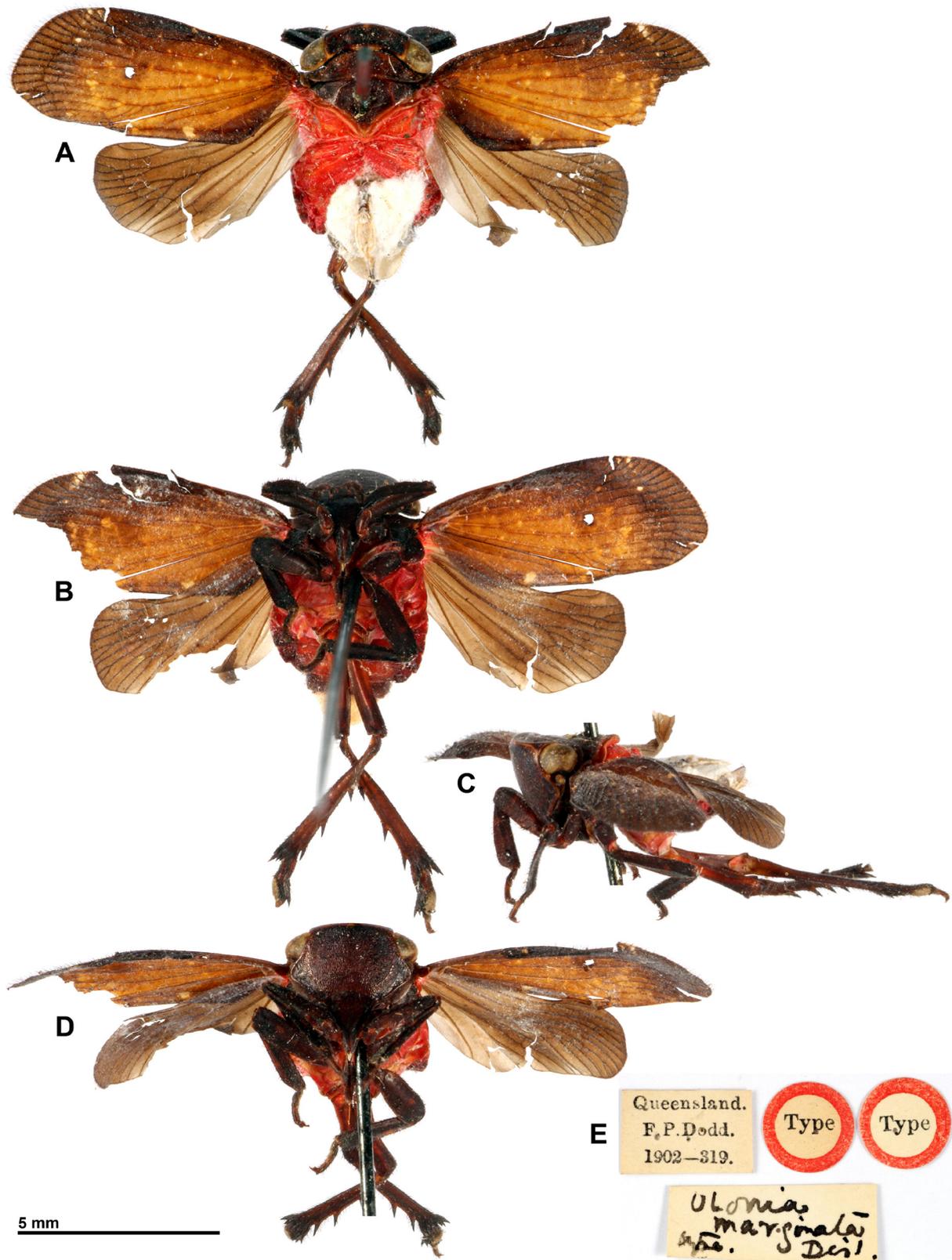


Fig. 15. *Olonia marginata* Distant, 1906, lectotype, ♀. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Labels.

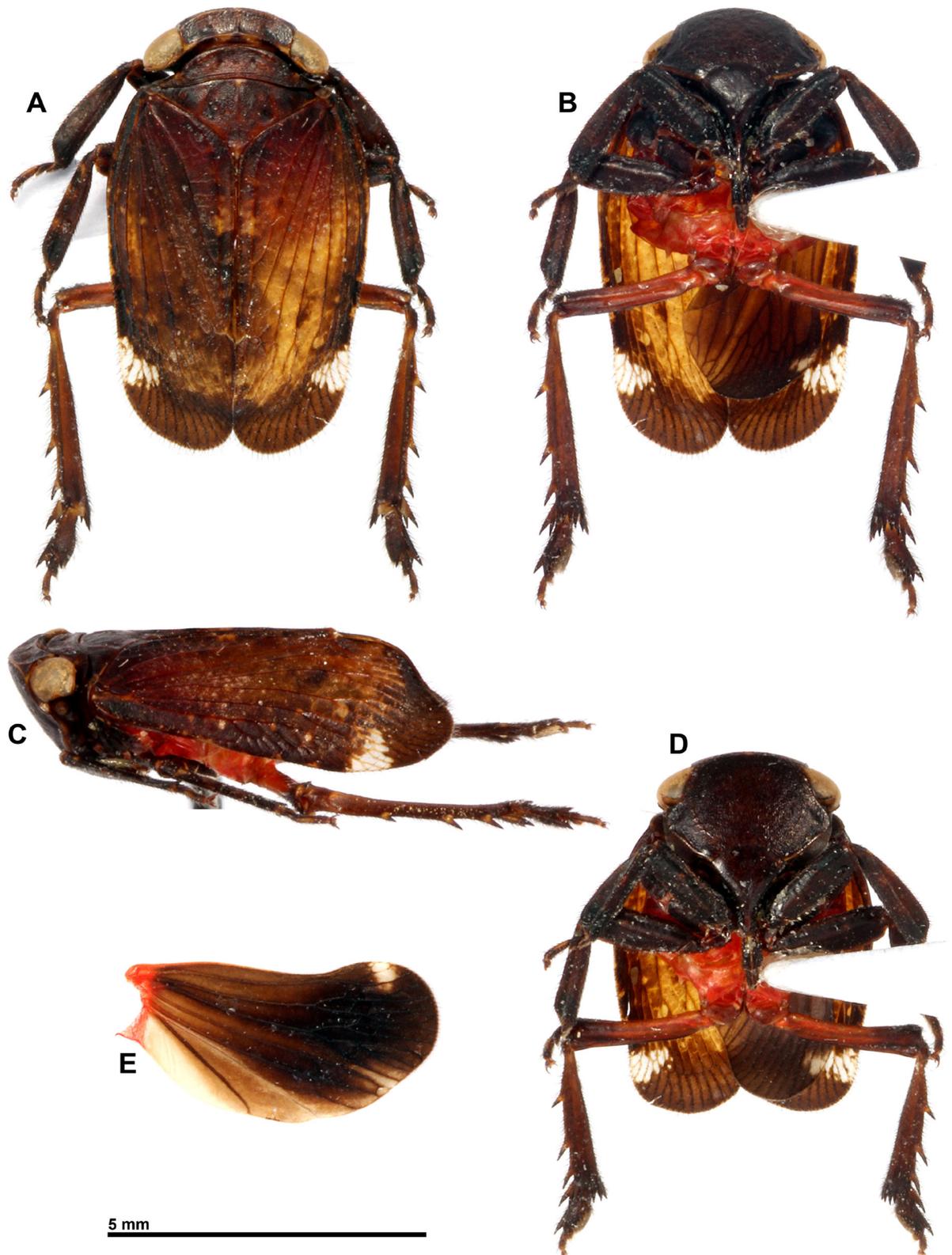


Fig. 16. *Olonia marginata* Distant, 1906, ♂ from mainland. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

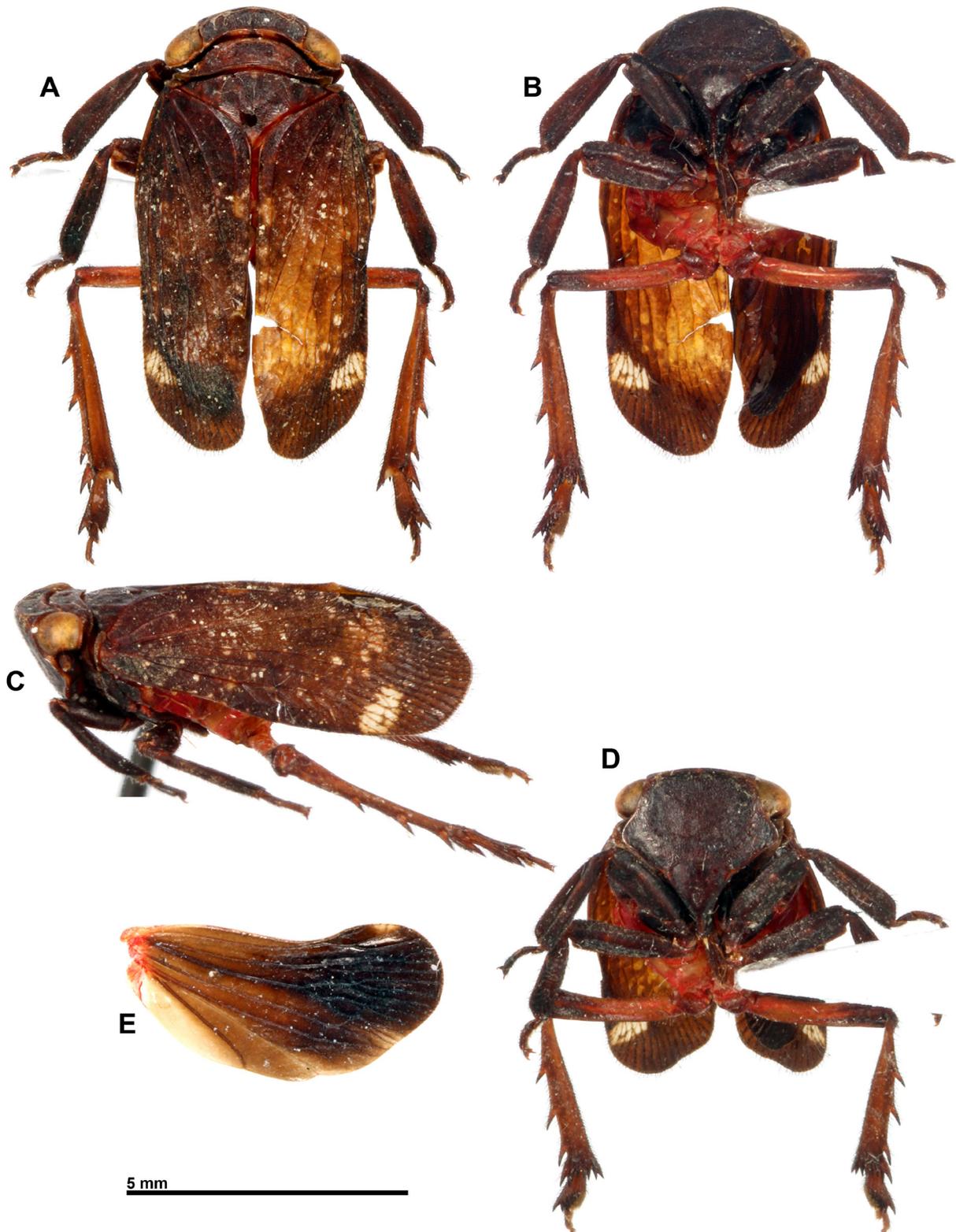


Fig. 17. *Olonia marginata* Distant, 1906, dark form, ♂ from Magnetic Island. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

third of length of centroventral part and projecting posteriorly (Fig. 19A, C). Centroventral part broad and dorsoventrally flattened on basal third, then strongly narrowing into a long spinose process strongly sinuate basally in lateral view and ending in narrow hook curved ventrally (Fig. 19A, C). Laterodorsal part of gonostyli strongly elongate and curved posteroventrally, surpassing level of centroventral part

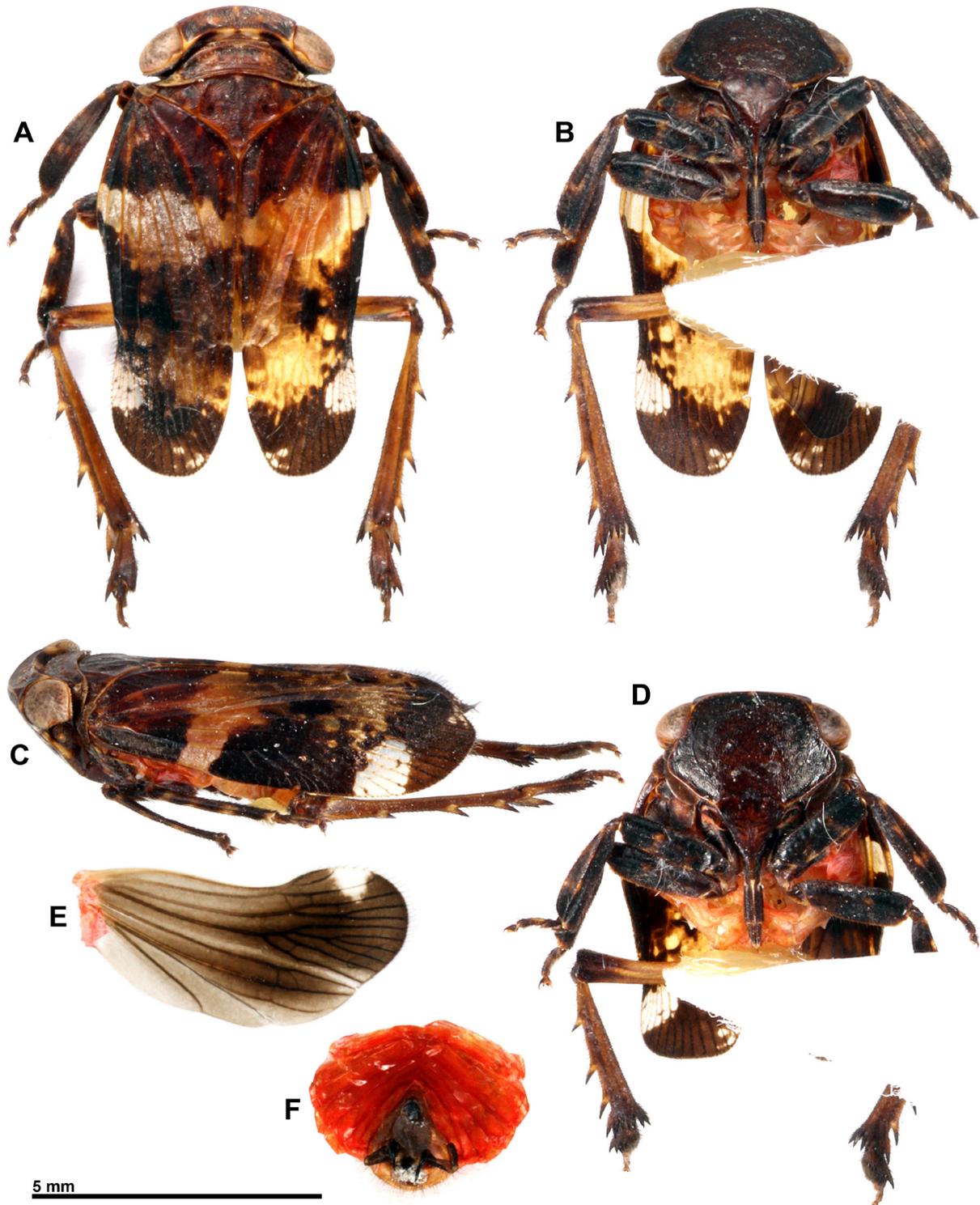


Fig. 18. *Olonia marginata* Distant, 1906, form with pale bands, ♂ from Magnetic Island. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing. **F.** Abdomen, dorsal view.

ventrally; lateral process elongate, projecting laterally and longer than spoon-shaped process (Fig. 19A, C–D). Dorsal portion of phallobase with hooked process on each side, progressively narrowing from base to apex, directed posterocentrally and with apex pointing dorsally (Fig. 19E–F). Ventral portion of phallobase trilobed in dorsal view, with median lobe surpassing phallus (Fig. 19E–F). Phallus dorsoventrally flattened, rather broad, with apical margin emarginate in middle (Fig. 19E–F).

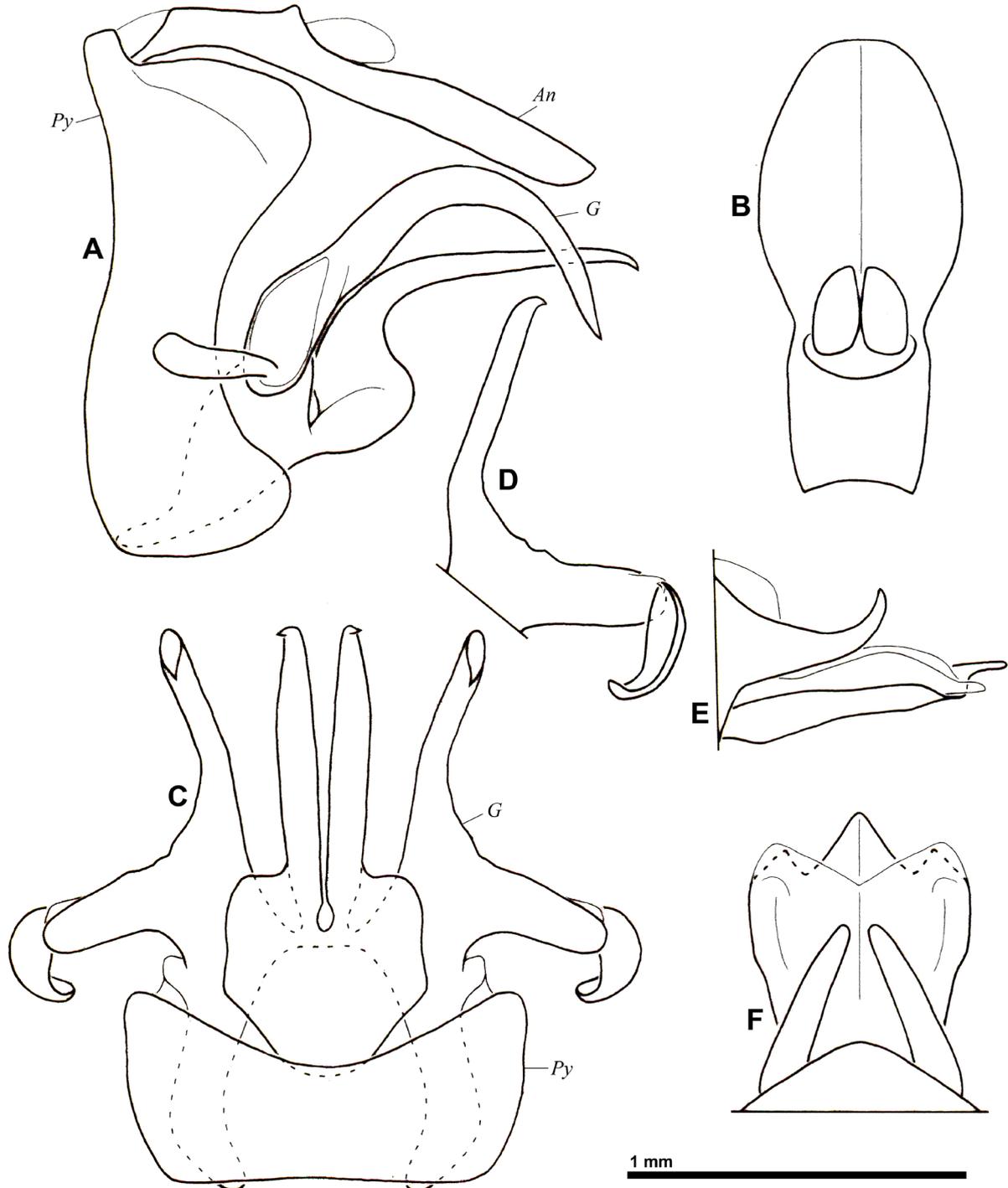


Fig. 19. *Olonia marginata* Distant, 1906, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

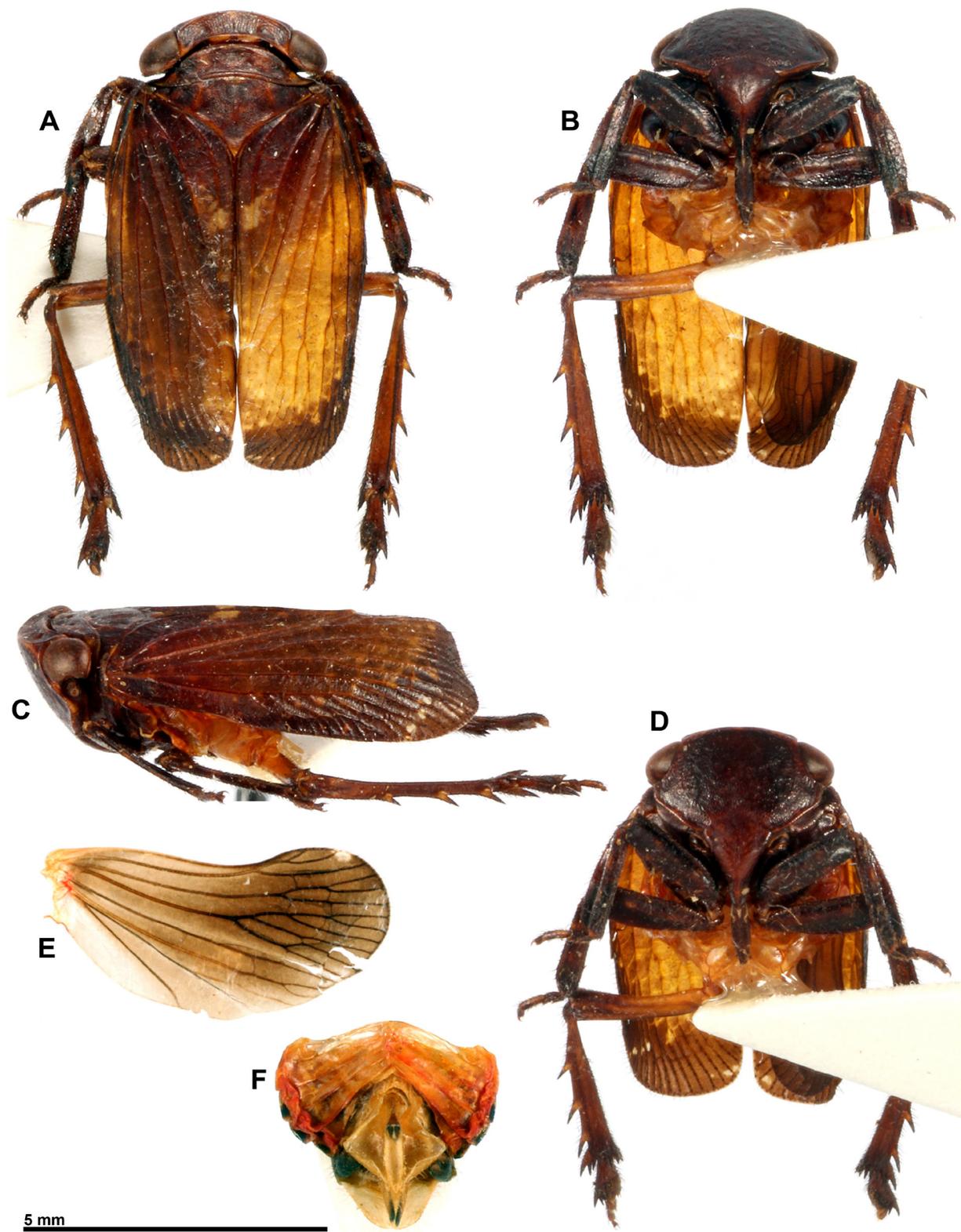


Fig. 20. *Olonia marginata* Distant, 1906, ♀ from mainland. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing. **F.** Abdomen, dorsal view.

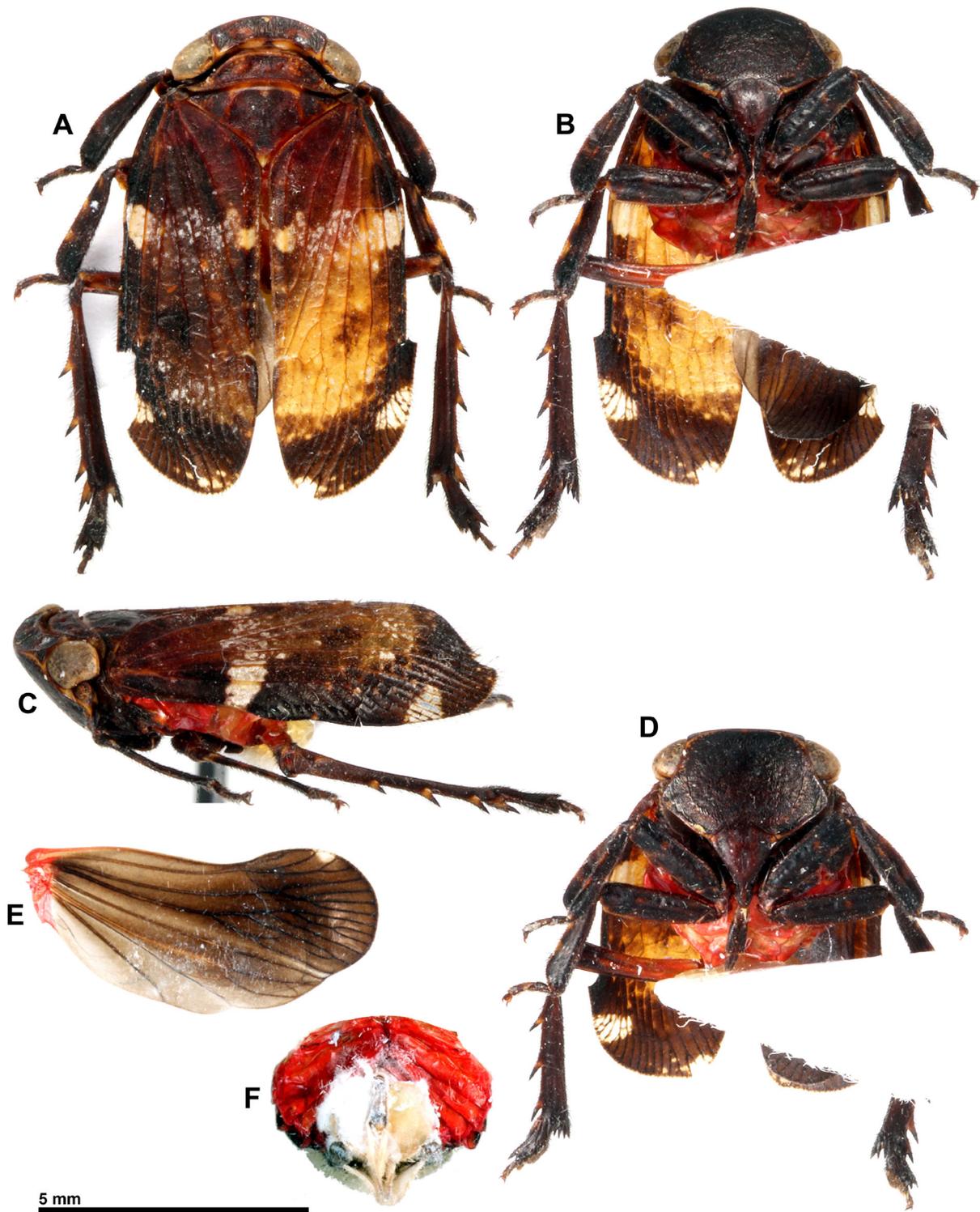


Fig. 21. *Olonia marginata* Distant, 1906, ♀ from Magnetic Island. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing. **F.** Abdomen, dorsal view.

Female

Similar to male, but with white spot at apicosutural angle of posterior wing reduced, extending on 1 cell (Figs 15, 20). Sometimes with a reduced white transverse band at basal $\frac{1}{3}$ of tegmina visible in a marking along costal and sutural margins (Fig. 21).

Nymph

HEAD (Fig. 22). Brown, as broad as pronotum, frons flattened, with lateral rounded row of yellowish tubercles; group of yellowish tubercles dorsally on each side of joint between frons and vertex; clypeus reaching mesocoxae.

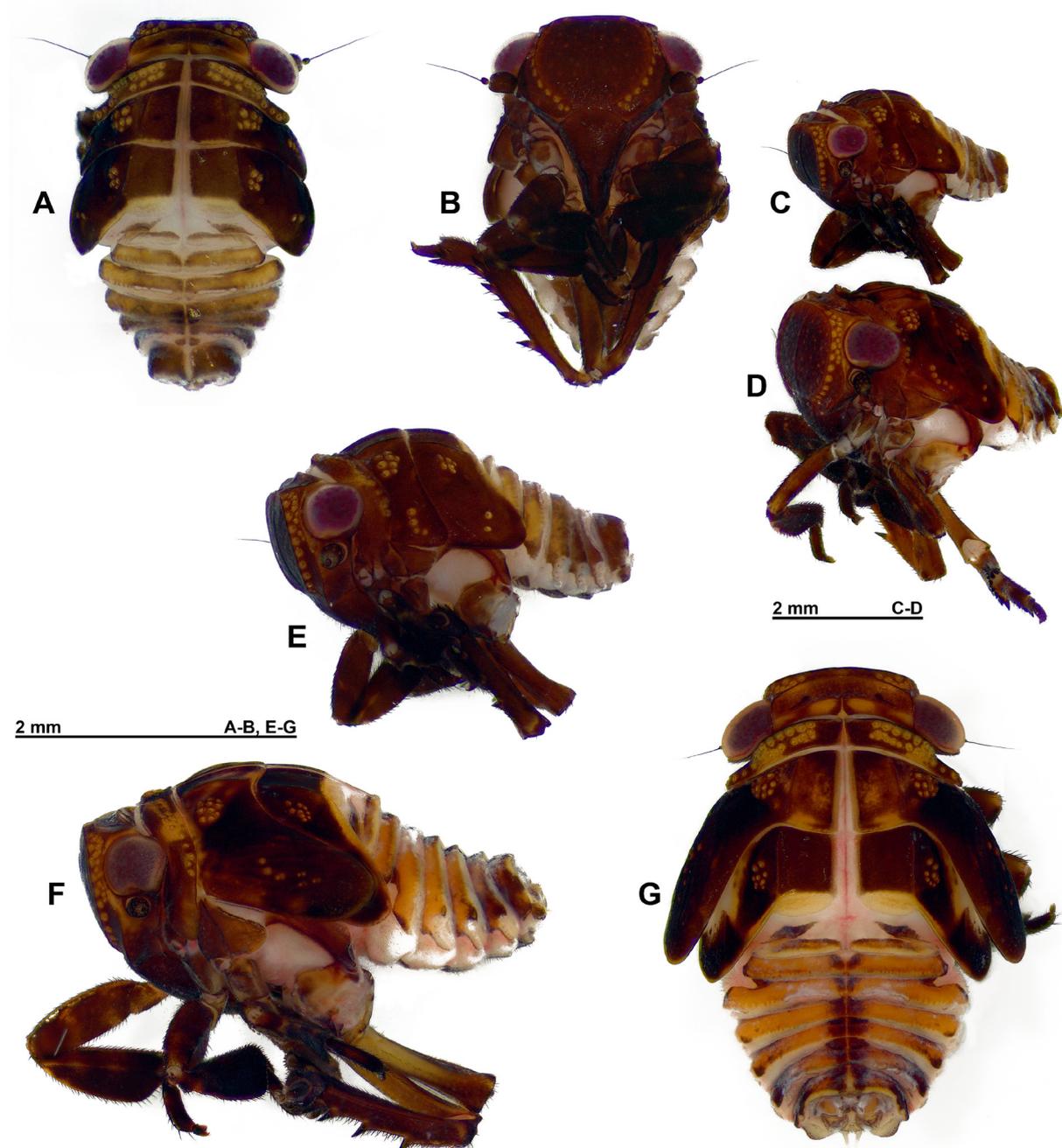


Fig. 22. *Olonia marginata* Distant, 1906, nymphs from Townsville, Bushland Beach, 4 Apr. 2016. A–E. Fourth instar nymphs. A. Dorsal view. B. Ventral view. C–E. Lateral view. F–G. Fifth instar nymph. F. Lateral view. G. Dorsal view.

THORAX (Fig. 22). Brown; pronotum short, with two dense rows of yellowish tubercles on each side; meso- and metanotum with a circular group of yellowish tubercles at base of wing buds and some tubercles near apex of wing buds.

LEGS (Fig. 22). Brown slightly variegated with yellowish; large in comparison to body size. Anterior and median femora and tibiae broad and dorsoventrally flattened; posterior tibiae with three strong lateral spines.

ABDOMEN (Fig. 22). Rather short, less than half as long as head and thorax combined, and narrower than thorax; pale yellowish brown with dorsal median line and apex darker. Live specimens with abdomen



Fig. 23. *Olonia marginata* Distant, 1906, in nature, Townsville. A–C. Bushland Beach, 4 Apr. 2016. A. Habitat. B–C. Nymph on *Ipomoea pes-caprae* (L.) R.Br. D. Egg case on *Ipomoea pes-caprae*, Rowes Bay Beach, 25 Feb. 2016. E–F. Townsville Port Wall, 15 Mar. 2016. E. Habitat. F. Female on *Ipomoea pes-caprae*. G. Female parasitized by larva of Epipyropidae (Lepidoptera), Rowes Bay Beach, 26 Feb. 2016. H. Habitat, Small Boat Harbour, 19 Feb. 2016. I. Habitat, Strand Beach, 15 Mar. 2016. J. Habitat, Pallarenda Beach, 25 Feb. 2016. Photographs by V. Ryland.

covered in white wax, except three apical segments with brown wax; each segment bearing a waxy plate on each side; two movable, very elongate, waxy filaments at apex of abdomen (Figs 23B–C, 25B–C, K, N).

Distribution and biology

This species is recorded from an area around Townsville in North Queensland (Fig. 3), in the Einasleigh Upland Savanna, Queensland Tropical Rainforests and Brigalow Tropical Savanna bioregions. Adult specimens were collected in January, February, March, April, August, September and October, while



Fig. 24. *Olonia marginata* Distant, 1906, in nature, Magnetic Island, 8 Nov. 2006, on *Canarium australianum* F.Muell. saplings. **A–B.** Male on 2 m tall sapling. **C.** Two m tall sapling of *C. australianum*. **D–E.** Female on small *C. australianum* sapling. **F.** Terminal leaves of small *C. australianum* sapling. **G.** Leaves of *C. australianum*. **H.** Fruits of *C. australianum*. **I.** The author and his 5 month old daughter Emilie observing specimens of *O. marginata*. Photographs by the author except I by L. Despontin.



Fig. 25. *Olonia marginata* Distant, 1906, in nature, Magnetic Island. **A–G.** Picnic Bay Beach. **A.** Habitat, 6 Feb. 2016. **B–C.** Nymph on *Ipomoea pes-caprae* (L.) R.Br, 6 Feb. 2016. **D.** Adult on *Ipomoea pes-caprae*, 6 Feb. 2016. **E.** Pair on *Corymbia tessellaris* K.D.Hill & L.A.S.Johnson regrowth, 31 Jan. 2016. **F.** Male on *Eucalyptus* sp. sapling, 10 Jan. 2016. **G.** Female on *Pipturus argenteus* Wedd., 29 Jun. 2014. **H–K.** Nelly Bay, 11 Feb. 2016. **H.** Habitat. **I.** Adults on *Ipomoea pes-caprae*. **J.** Male. **K.** Nymph on *Ipomoea pes-caprae*. **L–O.** Geoffrey Bay. **L.** Habitat, 10 Feb. 2016. **M.** Male on *Ipomoea pes-caprae*, 10 Feb. 2016. **N.** Nymph on *Ipomoea pes-caprae*, 10 Feb. 2016. **O.** Female on *Ipomoea pes-caprae*, 24 Feb. 2016. Photographs by V. Ryland.

nymphs were observed in Feb. and Apr. (Figs 23B–C, 25B–C, K), and an egg mass in Apr. (Fig. 23D), leading to the conclusion that the species is probably present most of the year. A caterpillar of an unidentified species of Epipyropidae (Lepidoptera) was observed on the back of a female specimen (Fig. 23G).

Olonia marginata is polyphagous and has been collected or observed on *Melaleuca quinquenervia* (Cav.) S.T.Blake (Myrtaceae), *Eucalyptus* sp. (Myrtaceae) (Fig. 25F), *Canarium australianum* F.Muell. (Bursaceae) (Fig. 24), *Vitis vinifera* L. (Vitaceae), *Pipturus argenteus* Wedd. (Urticaceae) (Fig. 25G), *Corymbia tessellaris* K.D.Hill & L.A.S.Johnson (Myrtaceae) (Fig. 25E) and in numbers on *Ipomoea pes-caprae* (L.) R.Br. (Convolvulaceae) (Figs 23B–D, F, 25B–D, I–K, M–O), on stems and young branches with a thin skin.

It is interesting to note that the 2016 sampled locations in Magnetic Island and Townsville (Figs 23A, E, H–J, 25A, H, L) were all more or less destroyed by the Tropical Cyclone Yasi in Feb. 2011, which covered these habitats in smashing waves and left them buried under sand and beach debris (pers. com. V. Ryland, Feb. 2016). The species has, however, re-colonised the areas in less than five years, displaying a good degree of adaptation to habitat disturbance.

Olonia maura (Fabricius, 1775)

Figs 3, 27–31

Cicada maura Fabricius, 1775: 686 (described).

Cicada maura – Goeze 1778: 158 (described, catalogued). — Fabricius 1781: 326 (described, catalogued); 1787: 272 (described); 1794: 40 (described). — Gmelin 1789: 2107 (described, catalogued). — Donovan 1805 (described, illustrated). — Turton 1806: 595 (described).

Flata maura – Fabricius 1803: 51 (described, catalogued, transferred to *Flata*). — Schaum 1850: 73 (listed).

Cicada matura [sic] – Donovan 1820: 120 (described).

Cercopis maura – Donovan 1820: 121 (described, transferred to *Cercopis*).

Lystra maura – Germar 1830: 55 (described, catalogued, transferred to *Lystra*).

Eurymela maura – Signoret 1850: 507 (described (based on a misidentified specimen according to Distant 1908: 105), transferred to *Eurymela*). — Walker 1852: 1149 (catalogued). — Kirkaldy 1906: 356 (listed).

Eurybrachys maura – Signoret 1858: xxxii (transferred to *Eurybrachys*).

Eurymeloides maura – Distant 1908: 105 (transferred to *Eurymeloides*).

Olonia maura – Evans 1933: 89 (transferred to *Olonia*). — Metcalf 1956: 64 (catalogued).

Note

Signoret (1850) based the transfer of this species to *Eurymela* (Cicadellidae) on a misidentified specimen (Fig. 26). This specimen indeed belongs to the tribe Eurymelini, identified as a member of the genus *Platyeurymela* Evans, 1933 or *Pauroeurymela* Evans, 1933 by M.J. Fletcher (pers. com., Apr. 2016) based on the examination of photographs of the specimen in NHMW.

Diagnosis

The species can be recognized by the following combination of characters:

- (1) hind wings without orange marking (Figs 29E, 31E)
- (2) pro- and mesofemora and -tibiae largely dark brown (Figs 29A–D, 31A–D)
- (3) anal tube of male spatulate, constricted at half length (Fig. 30B)

- (4) centroventral part of gonostyli with strong elongate process tapering distally and with 4–5 apical teeth (Fig. 30A, C)
- (5) laterodorsal part of gonostyli with strong furcate process, elongate process strongly curved posterolaterally (Fig. 30A, C–D)
- (6) rather small size: 7–8 mm

Females of *O. maura* are nearly impossible to separate from females of the sympatric *O. picea* and should be identified only if collected together with males.

Etymology

The species epithet *maurus* (adjective, Latin) means ‘Moor’, the inhabitants of North Africa, and by extension, ‘dark skinned’. It refers to the dark colour of the species.

Material examined

Lectotype (here designated)

AUSTRALIA • ♂; “Australia”, “63 47”, “BMNH(E) #668802”; here designated to provide a reference standard for the species; BMNH (Fig. 27).

Paralectotype

AUSTRALIA • ♀; “63 47”, “BMNH(E) #668803”; BMNH (Fig. 28).

Note

The lectotype and paralectotype are placed together in the Banks collection under two labels: “*Cicada maura*, Fab. Entom. p. 686.24”, “Type”. The specimens were collected by Joseph Banks and his team at the mouth of the Endeavour River (coordinates: 15°27'32" S, 145°13'28" E), when Captain James Cook’s ship *Endeavour* was beached and repaired, 17 Jun.–4 Aug. 1770 (Banks 1771).

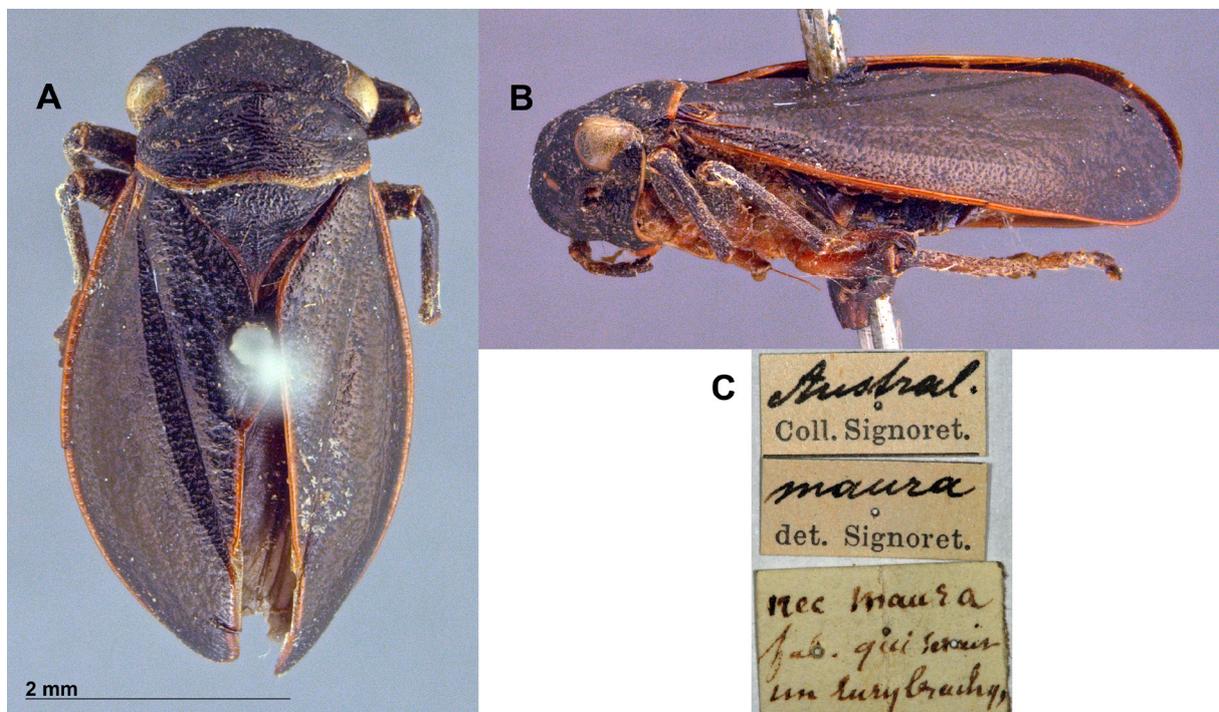


Fig. 26. Eurymelini sp. erroneously identified as *Olonia maura* (Fabricius, 1775) by Signoret (1850). **A.** Habitus, dorsal view. **B.** Habitus, lateral view. **C.** Labels. Photographs by H. Bruckner (NHMW).

Additional material

AUSTRALIA • ♂, paralectotype of *O. picea*; Cairns; 16°55'19" S, 145°46'32" E; Aug. 1904; BPBM • 1 ♂; Cairns; SAM • 1 ♂; Cairns; J.F. Illingworth leg.; on potato; BPBM • 1 ♂; Cairns; J.F. Illingworth leg.; on grass; ASCT • 1 ♀; same collection data as for preceding; SAM • 1 ♂; Cairns; J.F. Illingworth leg.; on ti tree; BPBM • 1 ♂; Gordonvale; 17°04'34" S, 145°47'40" E; 1 Mar. 1922; E. Jarvis leg.; on *Eucalyptus*; BMNH • 1 ♂; same locality as preceding; 28 Jun. 1923; W.C. Dormer leg.; QM • 1 ♂; Port Douglas; 16°29'01" S, 145°27'55" E; 9 Jul. 1971; Z. Liepa leg.; ANIC • 1 ♂; Annan River, 3 km SW of Black Mt; 15°41' S, 145°12' E; 27 Sep. 1980; T. Weir leg.; ANIC • 1 ♂; Cooktown; 31 Apr. 1972; RBINS • 1 ♂; Atherton Tablelands; [coordinates of Atherton: 17°15'37" S, 145°29'23" E]; J.G. Brooks leg.; AM • 1 ♂; Meringa; 17°04'42" S, 145°46'30" E; 27 Dec. 1946; J. Rosser leg.; QM • 1 ♂, 1 ♀; Meringa; Sep. 1919; F. Muir leg.; BPBM.

Note

One male labelled “Sol. Is., Guadalcanar I. [sic], 1-1921, J.A. Kusche coll., Collection of W.M. Giffard” from the BPBM has also been examined. This record from the Solomon Islands, very far off the Australian coast, is regarded as highly doubtful and a probable case of mislabeling.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 3): 7.2 mm (6.9–7.5); BV/LV = 3.6; BF/LF = 1.82; LP+LM/BT = 0.62; Ltg/BTg = 2.4; LW/BW = 1.97.

Male

HEAD (Figs 27A–D, 29A–D). Vertex slightly concave, with anterior and posterior margins parallel, curved; medium to dark brown with sides slightly darker. Frons uniformly coloured, medium to dark brown. Clypeus elongate, entirely brown. Genae brown to black with yellowish markings along anterior margin. Labium brown to black, reaching metacoxae. Antennae black-brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Figs 27A–D, 29A–D). Pronotum uniformly brown, sometimes variegated with yellowish; slightly wrinkled; 2 small impressed points on disc slightly marked; sometimes very obsolete median carina. Lateral fields of prothorax coloured as pronotum. Mesonotum medium to dark brown, somewhat darker than pronotum, sometimes variegated with yellowish; yellowish minute spot at apex of scutellum; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae medium to dark brown.

TEGMINA (Figs 27A, C, 29A, C). Medium to dark brown; often pale yellowish marking on vein A1 at midlength of clavus; marked with dark brown or black along costal margin, more broadly so on posterior half, and along posterior margin. Often darker, median, irregular marking at apical $\frac{2}{3}$; triangular white marking on costal margin on nodal line, sometimes reduced; sometimes, a number of minute white spots at apicosutural angle. Costal and sutural margins subparallel; costal margin nearly straight; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 29E). Brown, paler on anal area and progressively darker on apical half; small, subtriangular white marking at apicocostal angle, extending on 2–4 cells. Margin of anal area slightly sinuate; sutural margin with 1 slight cleft, cubital one nearly not marked.

LEGS (Figs 27A–D, 29A–D). Pro- and mesocoxae black-brown. Pro- and mesofemora black-brown variegated with brown. Pro- and mesotibiae medium to dark brown, sometimes with 3 paler obsolete rings. Pro- and mesotarsi black-brown with basal half of third tarsomere paler. Metacoxae reddish brown; metafemora reddish with apex brown. Metatibiae brown, with 3 lateral spines paler basally and 9 apical black-brown spines. Metatarsi brown with a ventral row of 6 black spines on first tarsomere.

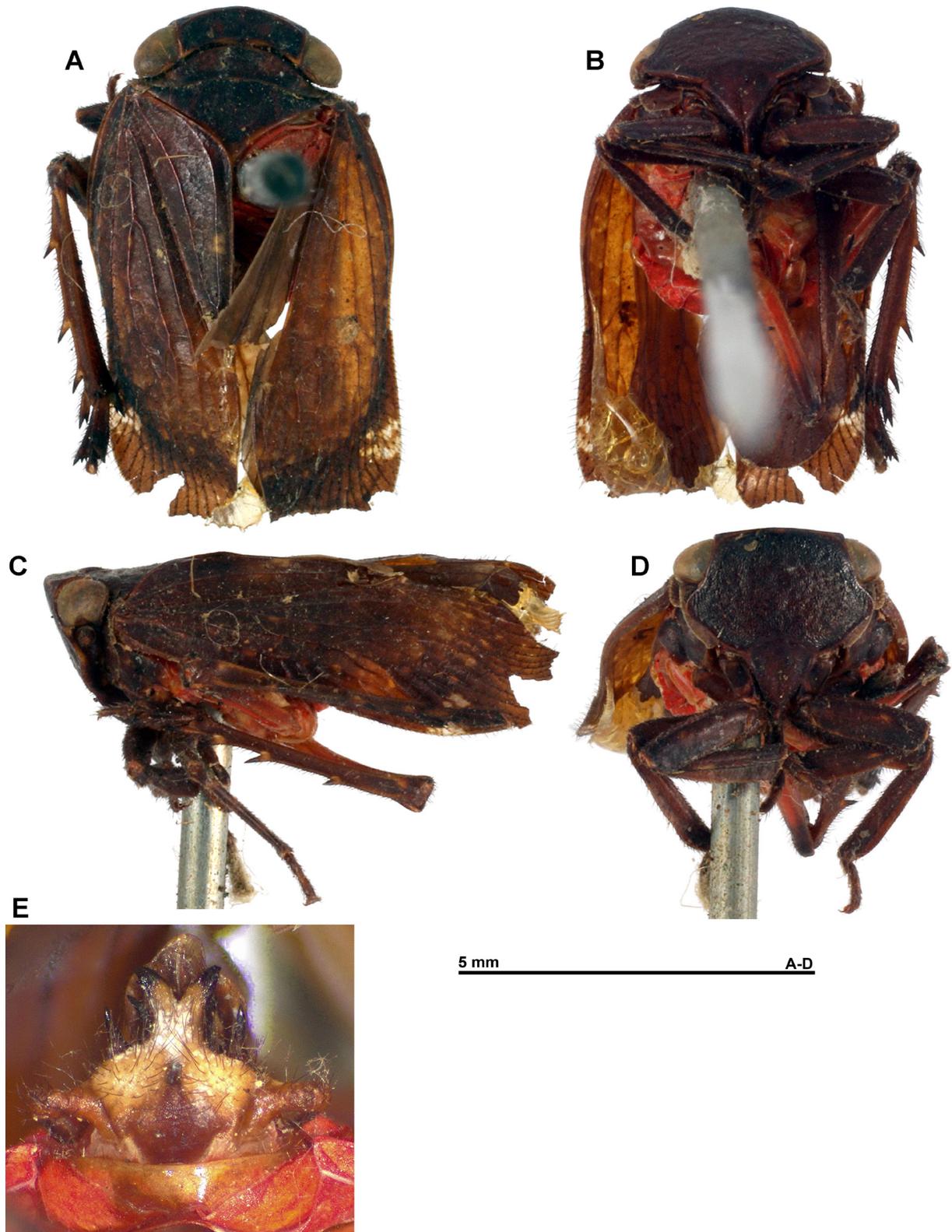


Fig. 27. *Olonia maura* (Fabricius, 1775), lectotype, ♂. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Genitalia, posteroventral view (not to scale).

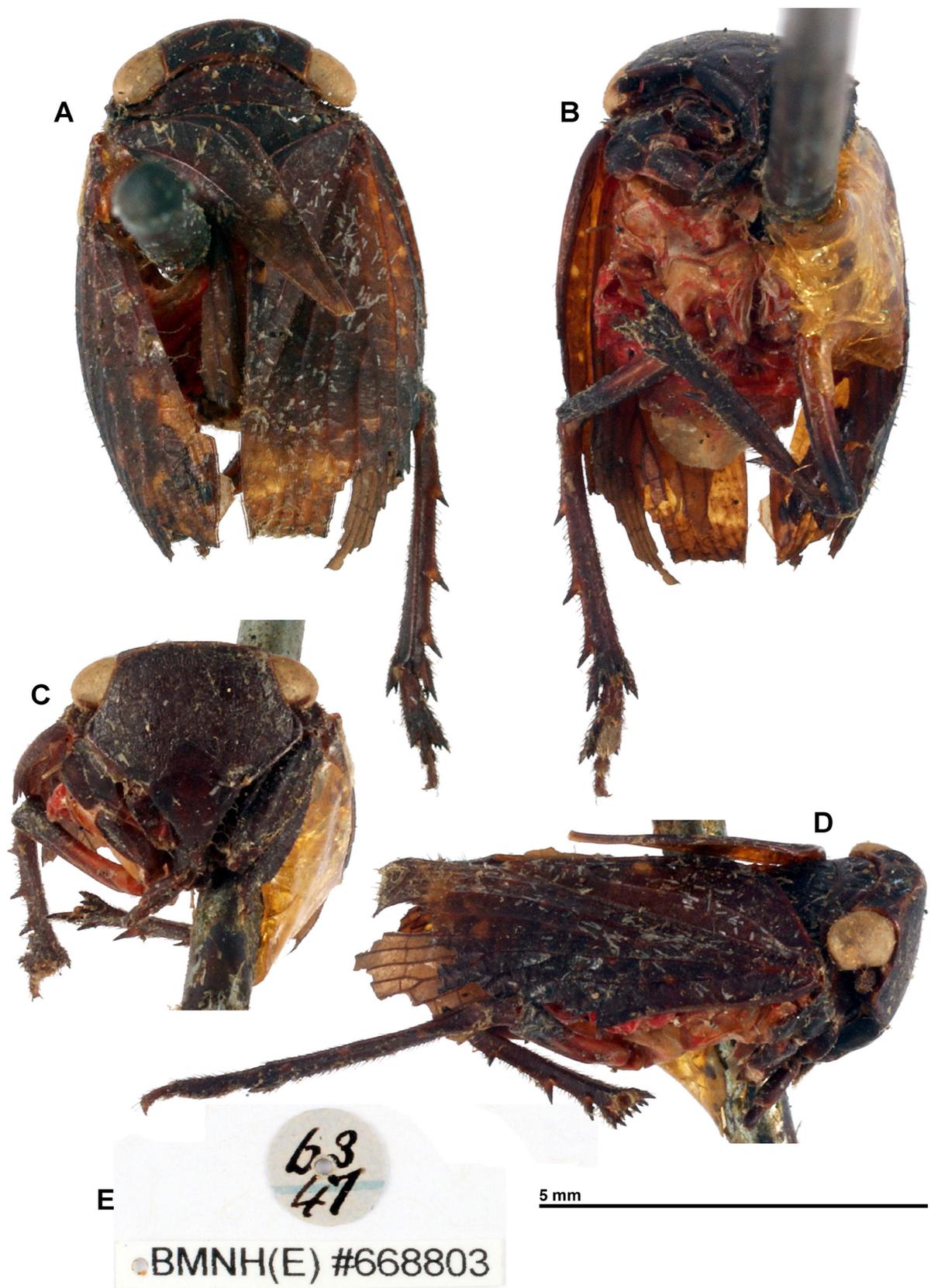


Fig. 28. *Olonia maura* (Fabricius, 1775), paralectotype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, normal view of frons. **D.** Habitus, lateral view. **E.** Labels.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 30). Posterior margin of pygofer in lateral view sinuate, strongly roundly projecting at dorsal $\frac{1}{4}$, moderately broad ventrally (Fig. 30A, C). Anal tube with lateral margins parallel before epiproct, constricted at level of epiproct, then obovate in apical half, 1.95 times as long as broad,

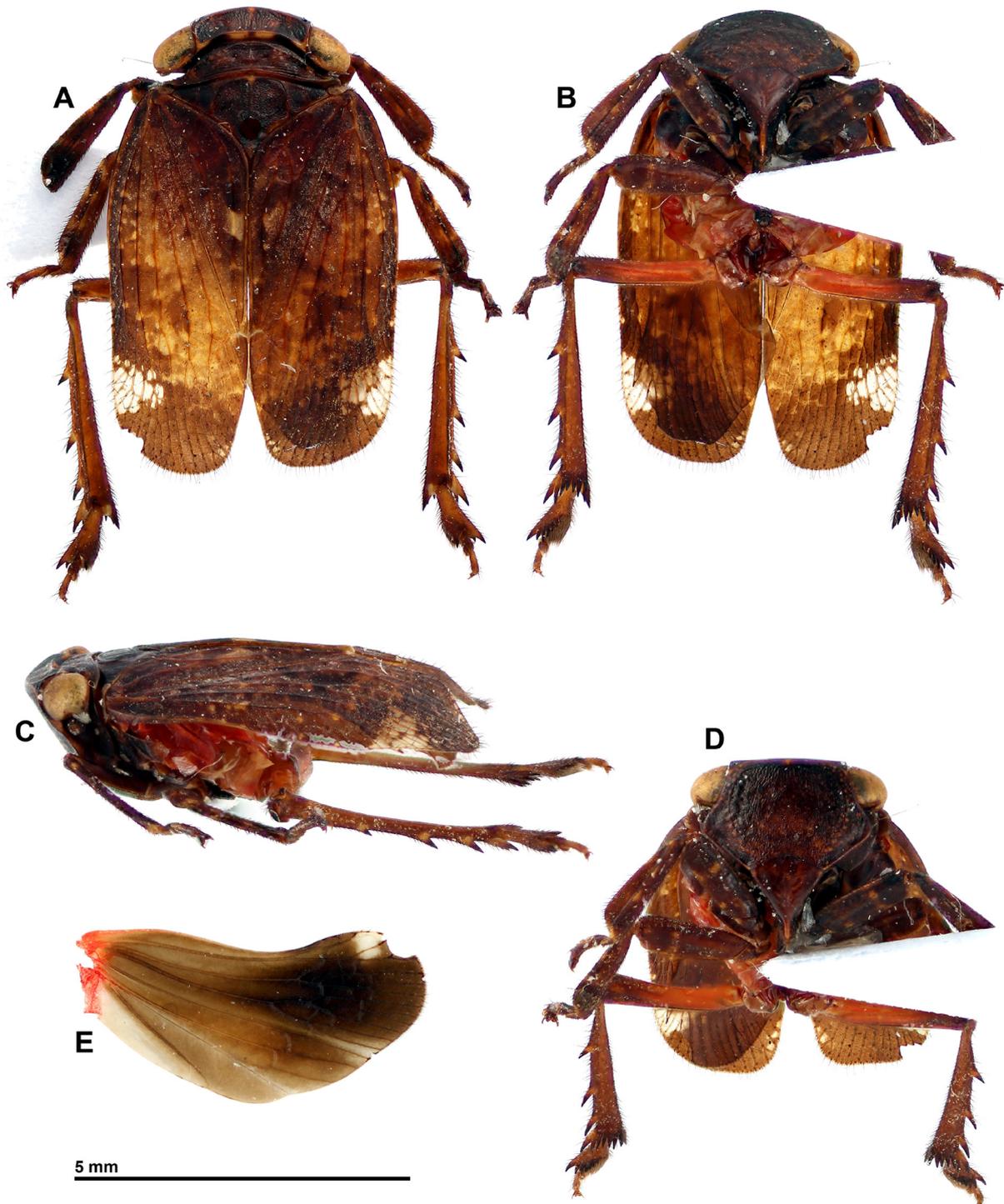


Fig. 29. *Olonia maura* (Fabricius, 1775), ♂. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Posterior wing.

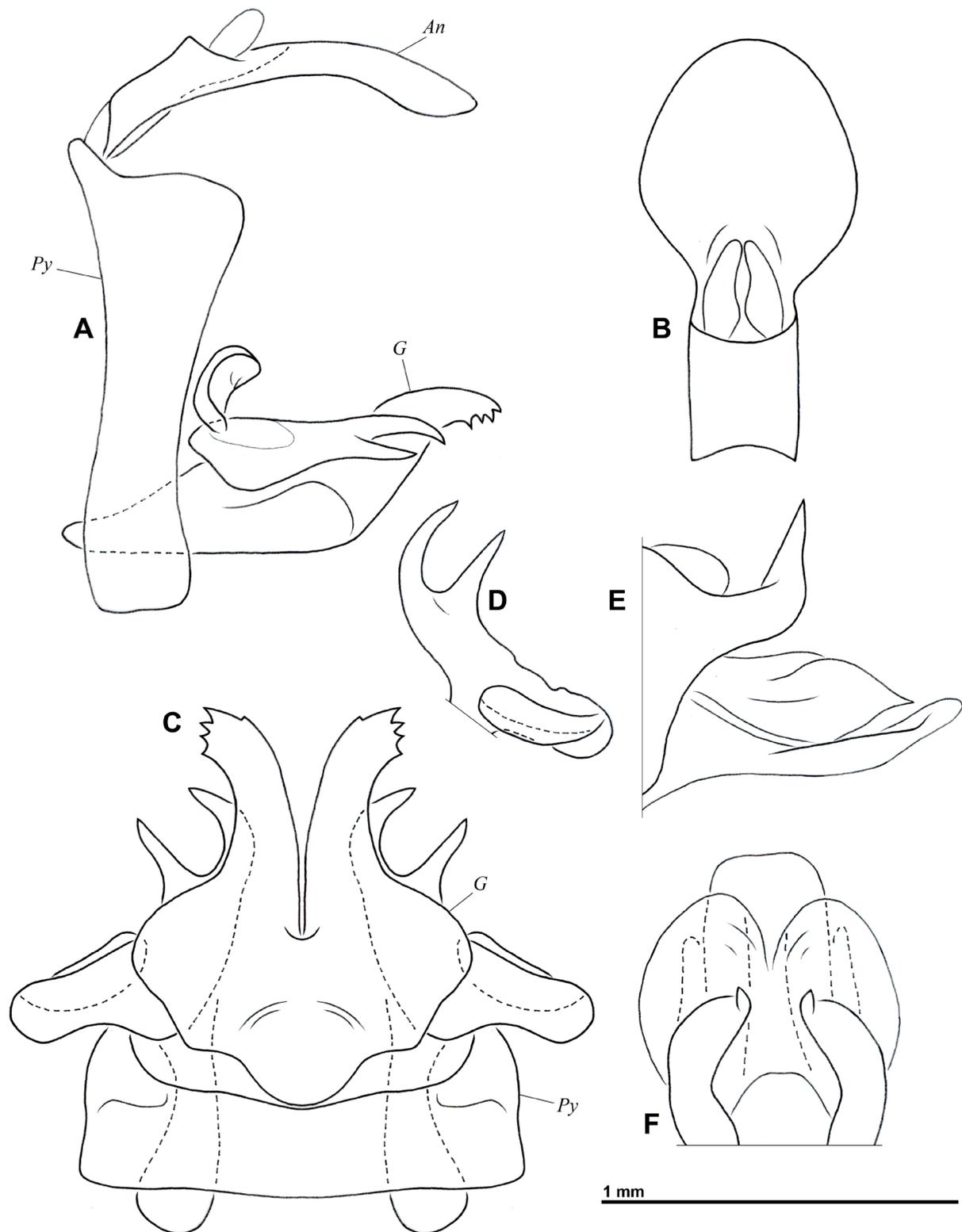


Fig. 30. *Olonia maura* (Fabricius, 1775), ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

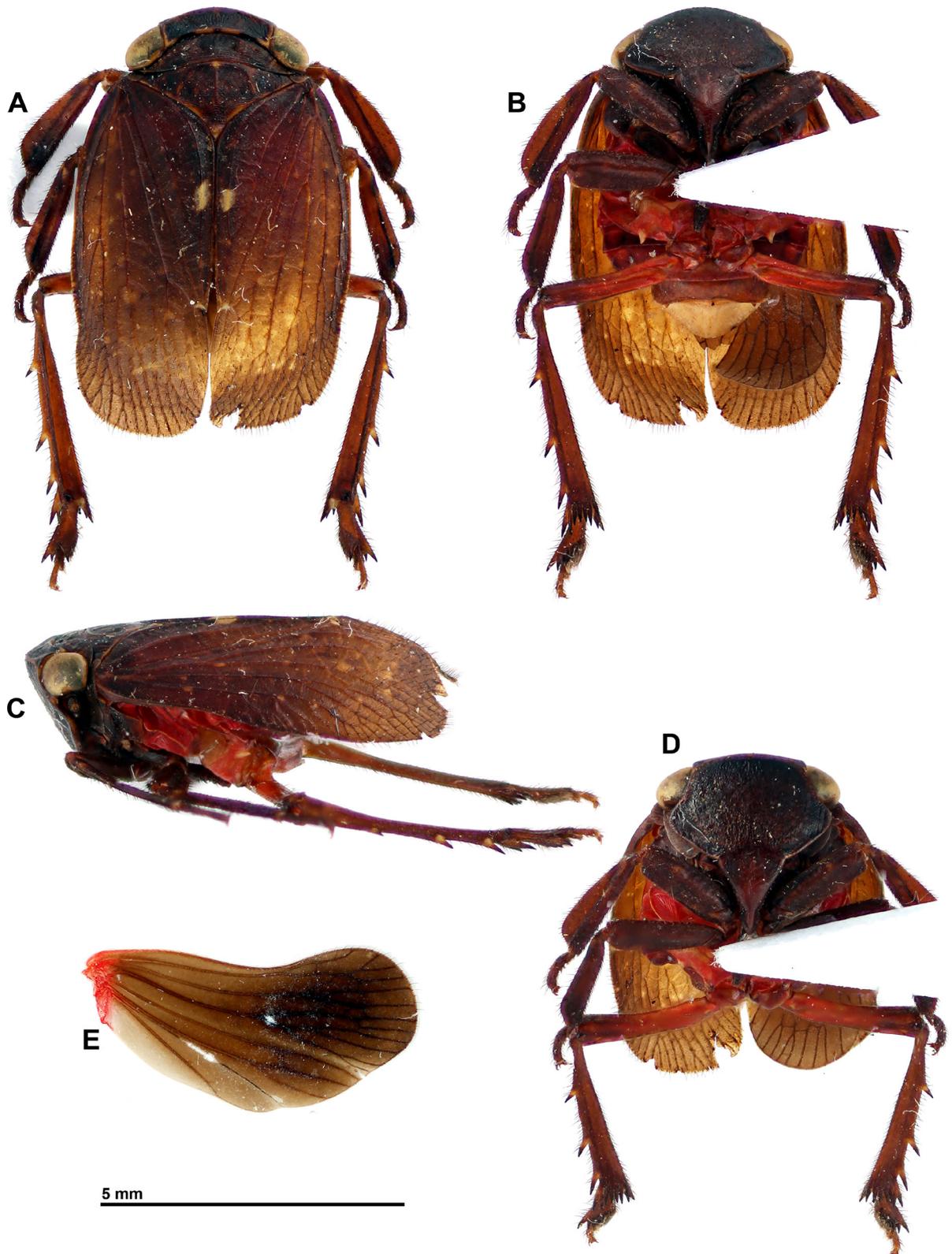


Fig. 31. *Olonia maura* (Fabricius, 1775), ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

slightly curved ventrally in lateral view; apical margin rounded (Fig. 30A–B). Gonostyli fused on basal $\frac{2}{5}$ of length of centroventral part and projecting posterodorsally (Fig. 30A, C). Centroventral part broad and dorsoventrally flattened on basal $\frac{2}{5}$, then with 2 strong divergent processes projecting posterodorsally, with apex curved ventrally and bearing 4–5 strong teeth (Fig. 30A, C). Laterodorsal part of gonostyli elongate and directed posterolaterally, dorsoventrally flattened, shorter than centroventral part and strongly bifid, apically with the two processes subparallel; lateral process elongate, projecting anterolaterally and about as long as spoon-shaped process (Fig. 30A, C–D). Dorsal portion of phallobase with hooked process on each side, progressively narrowing from base to apex, directed posterocentrally, slightly converging posteriorly, curved in dorsal view and with apex pointing dorsally (Fig. 30E–F). Ventral portion of phallobase unilobed in dorsal view, with median lobe surpassing phallus (Fig. 30E–F). Phallus dorsoventrally flattened, rather broad, with apical margin deeply emarginate in middle (Fig. 30E–F).

Female

Similar to male, but without white spot on costal margin of tegmina at nodal line and with white spot at apicosutural angle of posterior wing strongly reduced, visible as a small, slightly paler marking (Fig. 31).

Distribution and biology

This species is currently recorded from an area around Cairns in North Queensland (Fig. 3), in the Cape York Peninsula Tropical Savanna and Queensland Tropical Rainforests bioregions. Specimens were collected in March, April, June, July and August. One specimen was collected on *Solanum tubiferum* L. (Solanaceae) and one on *Eucalyptus* sp.

Olonia monteithi sp. nov.

[urn:lsid:zoobank.org:act:B4773609-83FD-4B11-9982-D4F96C291299](https://zoobank.org/act:B4773609-83FD-4B11-9982-D4F96C291299)

Figs 3, 32–34

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings without orange marking (Figs 32E, 34E) and with apical margin bordered with white in females (Fig. 34E)
- (2) pro- and mesofemora and -tibiae largely black-brown (Figs 32A–D, 34A–D)
- (3) anal tube of male obovate after epiproct and with posterior margin truncate (Fig. 33B)
- (4) centroventral part of gonostyli with strong pointed processes curved laterally (Fig. 33A, C)
- (5) laterodorsal part of gonostyli bifid, with dorsal process shorter than ventral one and both directed posteriorly (Fig. 33A, C–D)
- (6) medium size: 8.3–9.7 mm

Etymology

This species is dedicated to Dr Geoff Monteith, who collected the type specimens.

Type material

Holotype

AUSTRALIA • ♂; NE Queensland, Bakers Blue Mt, 17 km W of Mt Molloy; [16°42' S, 145°09' E]; “N.E. QLD, Bakers Blue Mt, 17 km W Mt Molloy, 800m, open For. 12.ix.1981, G. Monteith & D. Cook”, “QM-T24410”; QM.

Paratype

AUSTRALIA • 1 ♀; same collection data as for holotype; QM.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 1): 8.3 mm; ♀ (n = 1): 9.7 mm; BV/LV = 4.26; BF/LF = 1.67; LP+LM/BT = 0.64; Ltg/BTg = 2.47; LW/BW = 1.84.

Male

HEAD (Fig. 32A–D). Vertex concave, with anterior and posterior margins parallel, curved; black-brown slightly variegated with brown. Frons black-brown slightly wrinkled and with paler markings along dorsal margin. Clypeus elongate, black-brown, tinged with reddish in middle and with small reddish point at basolateral angles. Genae brown-black with yellowish markings around eyes and along anterior margin. Labium black-brown, reaching metacoxae. Antennae black-brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 32A–C). Pronotum black-brown slightly variegated with yellowish along anterior margin and with small yellowish spot at lateral angles; slightly transversely wrinkled posteriorly; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax black-brown. Mesonotum black-brown, slightly wrinkled, with apex of scutellum slightly reddish; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae black-brown.

TEGMINA (Fig. 32A, C). Black-brown with median area paler; small reddish spots on basal half and clavus; large white transverse marking along costal margin at half length, with additional smaller white spots in a row; large triangular white marking on costal margin on nodal line, nearly extending to white spots at apicosutural angle. Maximum breadth at basal $\frac{1}{3}$; costal margin sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 32E). Brown with apical half turning to black and anal area paler; transverse, broad, subtriangular white marking at apicocostal angle, extending on 7–8 cells. Margin of anal area slightly sinuate; sutural margin with 2 clefts, cubital one slightly marked.

LEGS (Fig. 32A–D). Pro- and mesocoxae black-brown. Pro- and mesofemora black-brown with 2 obsolete yellowish rings. Pro- and mesotibiae black-brown with 3 obsolete yellowish rings, larger one near apex. Pro- and mesotarsi dark brown with basal half of third tarsomere paler. Metacoxae reddish brown; metafemora red with apex black-brown. Metatibiae brown, with 3 lateral spines slightly yellowish basally and 8 apical, black-brown spines. Metatarsi dark brown, with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 33). Posterior margin of pygofer in lateral view sinuate, strongly roundly projecting at dorsal $\frac{1}{3}$, moderately broad ventrally (Fig. 33A, C). Anal tube with lateral margins subparallel before epiproct, constricted at level of epiproct, then obovate in apical half, 2.33 times as long as broad, slightly curved ventrally in lateral view; apical margin roundly truncate (Fig. 33A–B). Gonostyli fused on basal $\frac{2}{5}$ of length of centroventral part and projecting posterodorsally (Fig. 33A, C). Centroventral part broad and dorsoventrally flattened on basal $\frac{2}{5}$, then with 2 strong divergent processes curved laterally and pointed apically (Fig. 33A, C). Laterodorsal part of gonostyli elongate and directed posterodorsally, laterally flattened, not surpassing centroventral part and strongly bifid apically, with the two pointed processes forming an open C and projecting posteriorly, dorsal process shorter than ventral one; lateral process elongate, projecting posterolaterally and slightly longer than spoon-shaped process (Fig. 33A, C–D). Dorsal portion of phallobase with hooked process on each side, progressively narrowing from base to apex, directed dorsally, slightly converging apically (Fig. 33E–F). Ventral portion of phallobase unilobed in dorsal view, broadly rounded apically and with median lobe surpassing phallus (Fig. 33E–F). Phallus dorsoventrally flattened, rather broad, with apical margin deeply emarginate in middle (Fig. 33E–F).

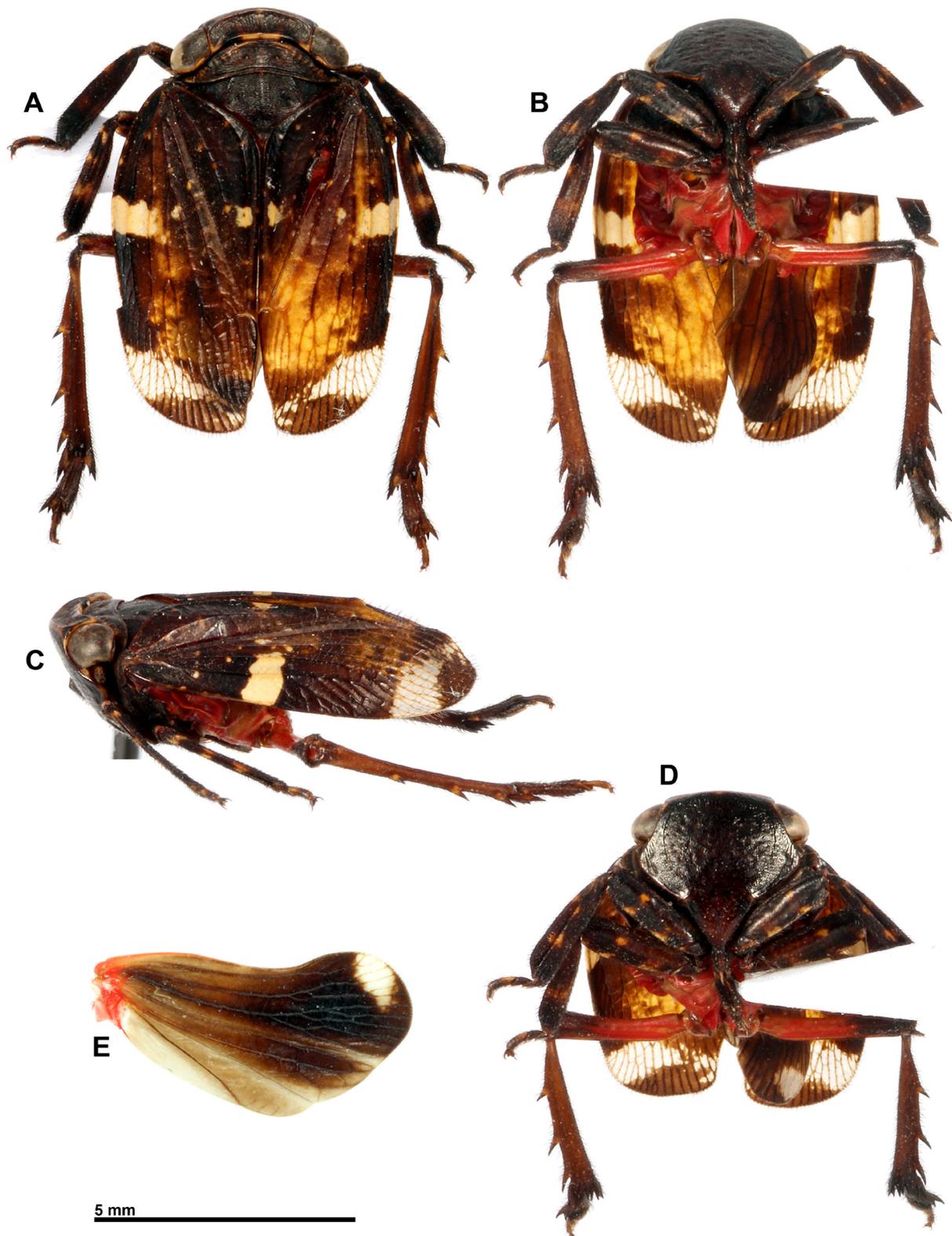


Fig. 32. *Olonia monteithi* sp. nov., holotype, ♂. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

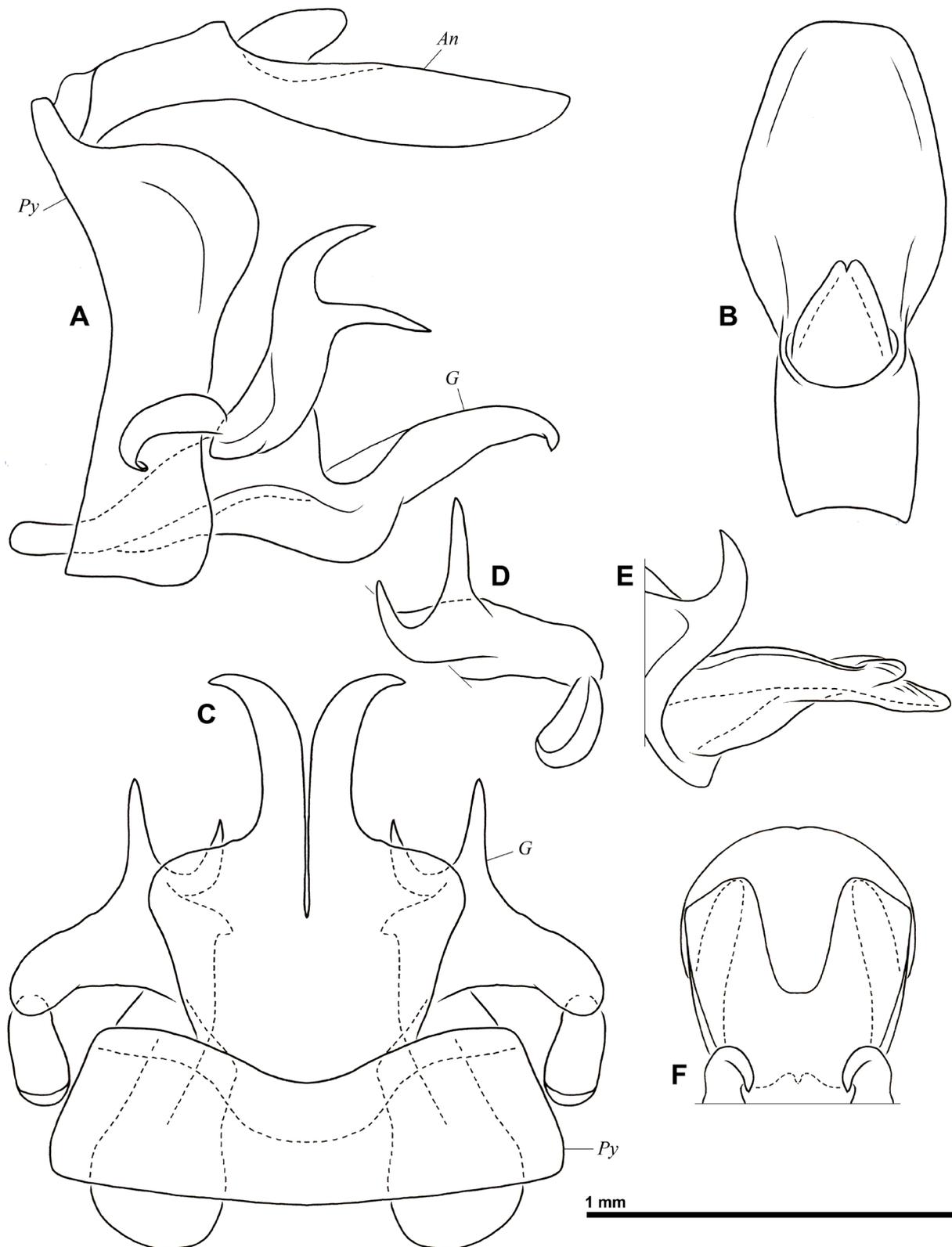


Fig. 33. *Olonia monteithi* sp. nov., holotype, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

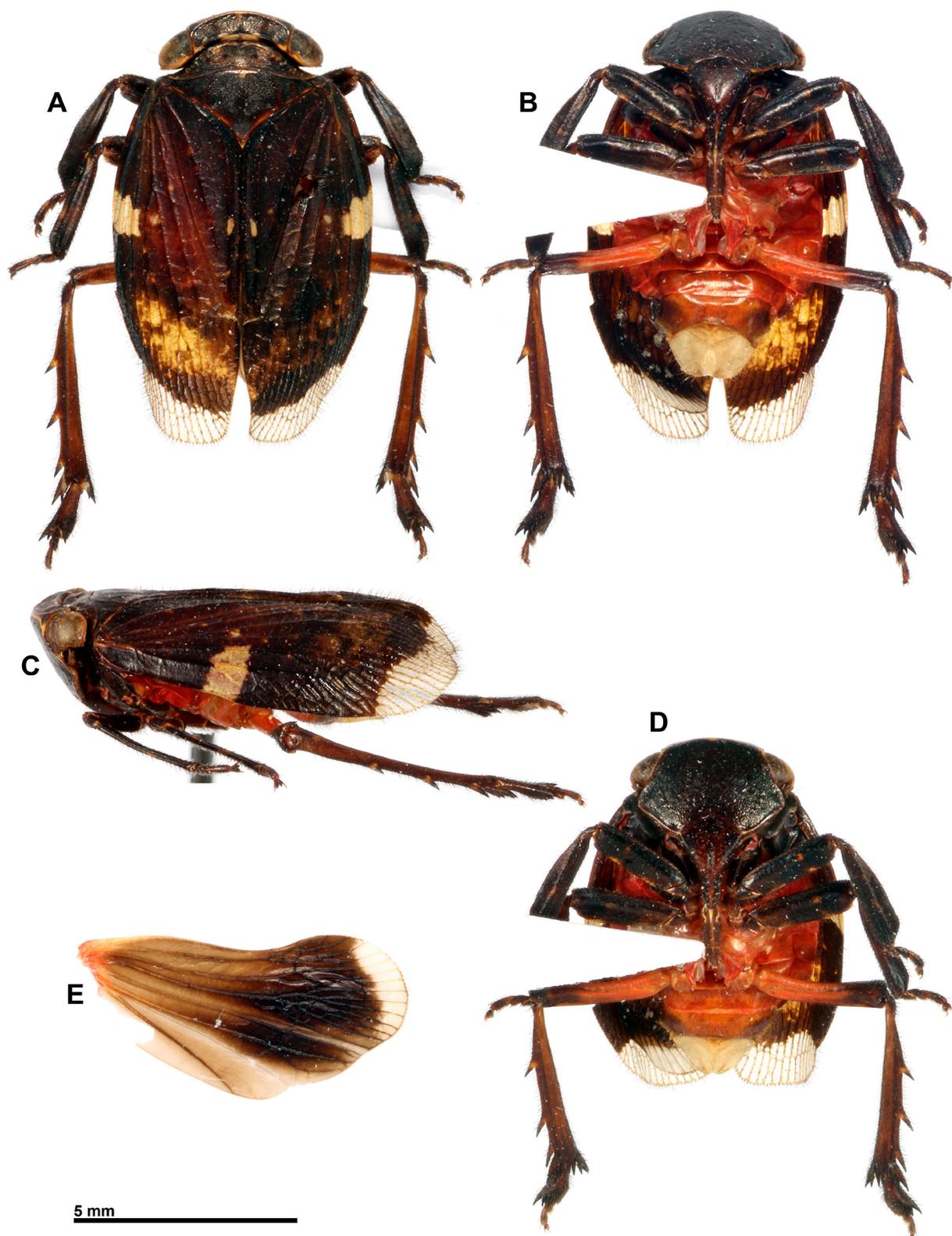


Fig. 34. *Olonia monteithi* sp. nov., paratype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

Female

Similar to male, but with markings on anterior and median legs less visible; tegmina with apical margin largely bordered with white; posterior wings with apical margin largely bordered with white, white area extending on 11 cells (Fig. 34).

Distribution and biology

This species is currently recorded from Blue Bakers Mountain, an isolated mountain area in North Queensland (Fig. 3), in the Einasleigh Upland Savanna bioregion. The specimens were collected in November in open forest at 800 m a.s.l. In an unpublished report available on the Internet, Ingram (1998) mentions that the location is a refuge with many endemic species of invertebrates (molluscs and isopods).

Olonia nobilis (Stål, 1863) comb. nov.
Figs 3, 35–39

Lyncilia nobilis Stål, 1863: 248 (described).

Lyncilia nobilis – Kirkaldy 1906: 448 (mentioned as unknown to the author in a work on Australian Auchenorrhyncha). — Metcalf 1956: 68 (catalogued).

Platybrachys nobilis – Fennah 1964: 160 (transferred to *Platybrachys* Stål, 1959; *nomen implicitum*).

Note

Fennah (1964) synonymized *Lyncilia* Stål, 1863 with *Platybrachys* Stål, 1859 but failed to formally propose the new combination *Platybrachys nobilis* (Stål, 1863) for the single species contained in that genus. The latter combination is here treated as a *nomen implicitum*. Examination of the holotype and recent additional material of *Lyncilia nobilis* revealed that the species actually belongs to the genus *Olonia*.

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings with large conspicuous orange marking (Figs 36E, 38E) and with narrow apical white border in females (Fig. 38E)
- (2) pro- and mesofemora and -tibiae largely black-brown (Figs 36A–D, 38A–D)
- (3) anal tube of male acuminate apically and narrowing basally to level of epiproct (Fig. 37B)
- (4) centroventral part of gonostyli with long and narrow process curved ventrointernally (Fig. 37A, C)
- (5) laterodorsal part of gonostyli with long and narrow process curved ventrally (Fig. 37A, C–D)
- (6) rather large size: 9–11 mm

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Lizard Island; [14°40'08" S, 145°27'34" E]; “Lizard Inseln”, “Type”, “*Lyncilia nobilis* Stål, ♀, Edm. Schmidt determ. 1907”, “Mus. Zool. Polonicum Warszawa 12/45”; ZMPA (Fig. 35).

Other material

AUSTRALIA • 1 ♂; NE Queensland, Lizard Island; 5 Jul. 1987; J. Grimshaw leg.; QDPI • 2 ♀♀; collection data as for preceding; on track to airstrip; 8 Jul. 1987; QDPI • 1 ♀; collection data as for preceding; 3 Jul. 1987; hand collected near research station; QDPI • 1 ♀; same collection data as for preceding; RBINS • 1 ♂, 1 ♀; Lizard Island; 15 Nov. 1974; M.S. and B.J. Moulds leg.; AMS.

Material examined from photographs

AUSTRALIA • 2 ♀♀; Lizard Island, Casuarina Beach; 1 Mar. 2016; A. Hoggett leg.; on *Ipomoea pes-caprae*.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 2): 9.0 mm; ♀ (n = 5): 10.5 mm (10.2–10.8); BV/LV = 4.4; BF/LF = 1.76; LP+LM/BT = 0.68; Ltg/BTg = 2.2–2.4; LW/BW = 1.72.

Male

HEAD (Fig. 36A–D). Vertex concave, with anterior and posterior margins parallel, curved; brown variegated with yellowish. Frons reddish black-brown, slightly rugulose. Clypeus elongate, dark reddish brown. Genae black-brown with yellowish markings around eyes and along anterior margin. Labium black-brown, reaching metacoxae. Antennae dark brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 36A–D). Pronotum brown variegated with yellowish and blackish; slightly transversely wrinkled posteriorly; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax dark brown. Mesonotum dark brown variegated with reddish and blackish, with median yellowish spot along anterior margin and at apex of scutellum; slightly transversely wrinkled; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae brown.

TEGMINA (Fig. 36A, C). Brown with costal margin along posterior half and apical margin after nodal line, bordered with black; small reddish to yellowish spots; conspicuous transverse, white marking along costal margin at half length; small white marking on vein A1 at half length of clavus; triangular white marking on costal margin on nodal line; white spot at apicosutural angle. Maximum breadth at basal third; costal margin sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 36E). Bright orange with apical $\frac{1}{3}$ black; basal $\frac{1}{4}$ between anal fold and vein MP, anal area and border along sutural margin brown; transverse, broad, white marking at apicocostal angle, extending on 2 cells. Margin of anal area straight; sutural margin with 2 clefts, cubital one slightly marked.

LEGS (Fig. 36A–D). Pro- and mesocoxae black-brown. Pro- and mesofemora black-brown. Pro- and mesotibiae black-brown with 3 obsolete ring-shaped reddish to yellowish markings, larger one near apex, more developed on mesotibiae. Pro- and mesotarsi dark brown, with basal half of third tarsomere paler. Metacoxae reddish brown; metafemora reddish with apex dark brown. Metatibiae brown, with 3 lateral spines yellowish basally and 8 apical, black-brown spines. Metatarsi dark brown, with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 37). Posterior margin of pygofer in lateral view strongly sinuate, strongly roundly projecting at dorsal $\frac{1}{2}$, narrow ventrally (Fig. 37A, C). Anal tube subovate, 2.1 times as long as broad, laterally emarginate at level of epiproct, curved ventrally in lateral view; lateral margins slightly curved ventrally on apical $\frac{2}{3}$; narrowing and acuminate apically (Fig. 37A–B). Gonostyli fused basally and projecting posteroventrally (Fig. 37A, C). Centroventral part elongate and narrow, diverging at base, then curved centroventrally, crossing near apex (Fig. 37A, C). Laterodorsal part of gonostyli elongate and narrow, progressively slightly curved posteroventrally, ventrally surpassing level of centroventral part; lateral process elongate, projecting lateroventrally and slightly concave dorsally; slightly longer than spoon-shaped process (Fig. 37A, C–D). Dorsal portion of phallobase with short and strong hooked process on each side, abruptly curved dorsally (Fig. 37E–F). Ventral portion of phallobase forming



Fig. 35. *Olonia nobilis* (Stål, 1863) comb. nov., holotype, ♀. A. Habitus, dorsal view. B. Habitus, lateral view. C. Habitus, normal view of frons. D. Habitus, ventral view. E. Labels.

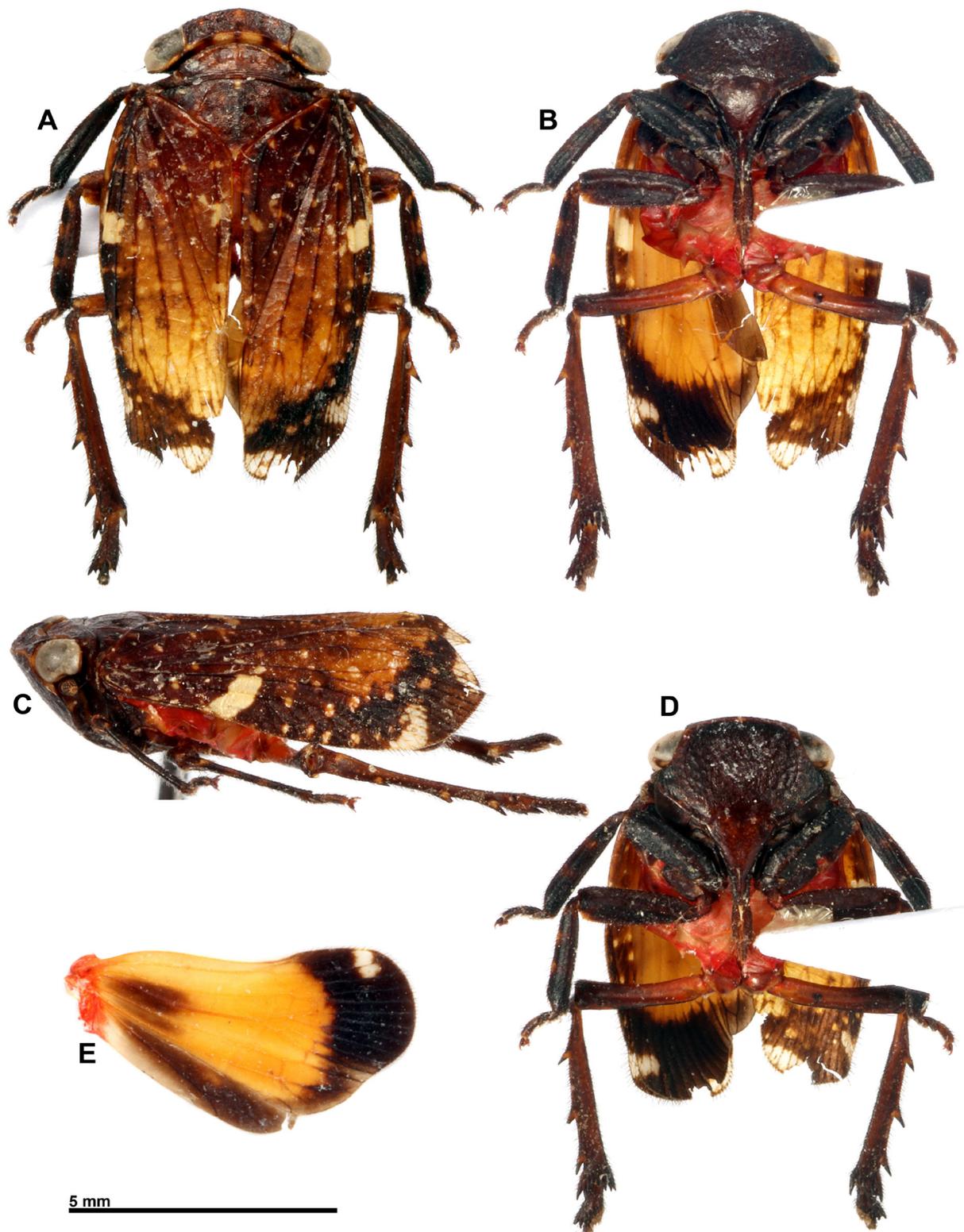


Fig. 36. *Olonia nobilis* (Stål, 1863) comb. nov., ♂. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

elongate process along each side of phallus, not merging apically (Fig. 37E–F). Phallus dorsoventrally flattened, obovate (Fig. 37E–F).

Female

Similar to male, but tegmina without white spot along costal margin on nodal line and at apicosutural angle, replaced by narrow white border along apical margin; posterior wings brown with apical ¼ black,

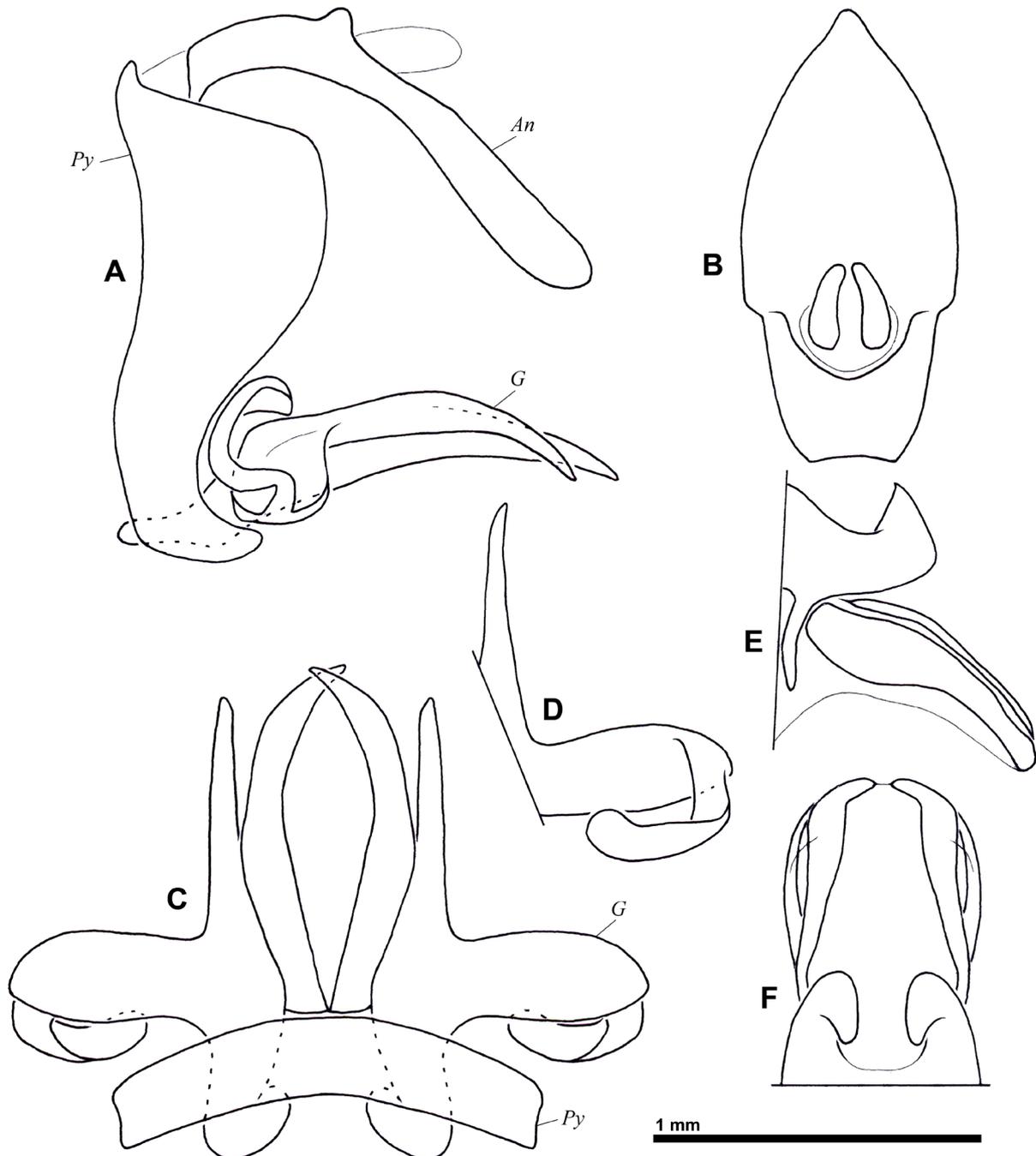


Fig. 37. *Olonia nobilis* (Stål, 1863) comb. nov., ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

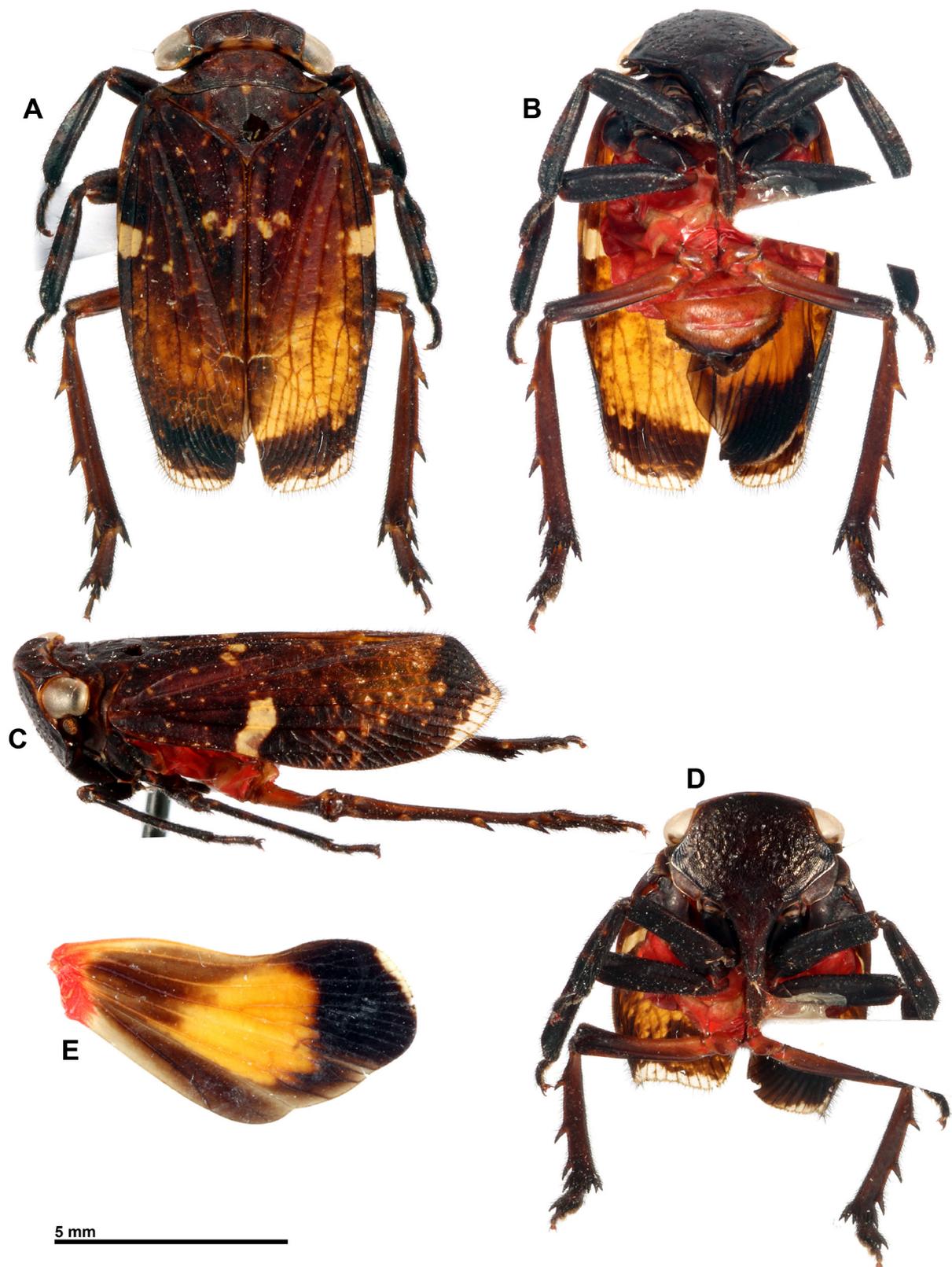


Fig. 38. *Olonia nobilis* (Stål, 1863) comb. nov., ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

large central bright orange marking extending from anal fold to vein ScP+R, and apical margin narrowly bordered with white along 5–6 cells (Fig. 38).

Distribution and biology

This species has only been recorded from Lizard Island, a small 10 km² island where it seems to be restricted, off the eastern coast of Cape York Peninsula (Fig. 3) in the Cape York Peninsula Tropical Savanna bioregion. The specimens were found in March, July and November, at low altitude according to the available data (the island culminates at 359 m a.s.l.). Several specimens were observed on stems of *Ipomoea pes-caprae* but not on those of *I. macrantha* Roem. & Schult. (A. Hoggett pers. com.) (Fig. 39).

Olonia picea Kirkaldy, 1906

Figs 3, 40–43

Olonia picea Kirkaldy, 1906: 445 (described; compared with *O. rubicunda*, *O. transversa* and *O. apicalis*).

Olonia picea – Kershaw & Muir 1922: 208 (note on male genitalia). — Muir 1923: 231, pl. 5, fig. 12 (male genitalia described and illustrated). — Jacobi 1928 (synonymized with *O. transversa* (erroneous)). — Metcalf 1956: 66 (catalogued, as *O. transversa* (erroneous)). — Constant 2005b: 66 (removed from synonymy with *O. transversa*).

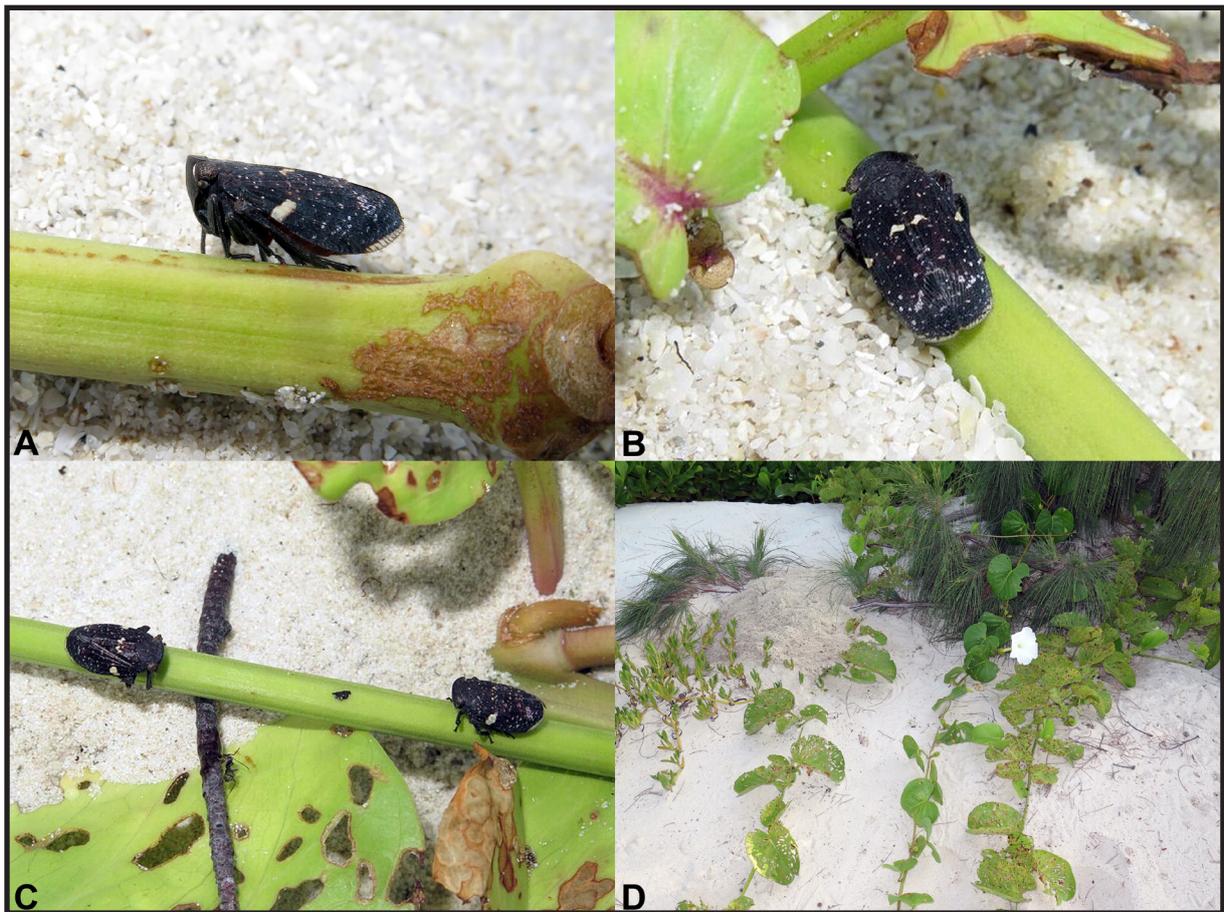


Fig. 39. *Olonia nobilis* (Stål, 1863) comb. nov. in nature, Lizard Island, Casuarina Beach, 1 Mar. 2016. A–B. Female on *Ipomoea pes-caprae*. C. Two females on *Ipomoea pes-caprae*. D. Habitat. Photographs by A. Hoggett.

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings without orange marking (Figs 41E, 43E)
- (2) pro- and mesofemora and tibiae largely black-brown (Figs 41A–D, 43A–D)
- (3) anal tube of male obovate, narrowing at basal $\frac{1}{3}$ (Fig. 42B)
- (4) centroventral part of gonostyli with strong, elongate process curved laterally and pointed apically (Fig. 42A, C)
- (5) laterodorsal part of gonostyli strongly bifid, with dorsal and ventral process forming a C together (Fig. 42A, C–D)
- (6) rather small size: 6.5–7.5 mm

Females of *O. picea* are nearly impossible to separate from females of the sympatric *O. maura* and should be identified only if collected together with males.

Etymology

The species epithet *piceus* (adjective, Latin) means ‘piceous’, ‘brown’. It refers to the general colour of the species.

Material examined

Lectotype (here designated)

AUSTRALIA • 1 ♂; Queensland, Cairns; [16°55'19" S, 145°46'32" E]; “Cairns, Q., Austr. 8-1904”, “Allotype”, “*Olonia picea* Kirk. ♂ 308”; here designated to provide a reference standard for the species; BPBM (Fig. 40).

Paralectotypes

AUSTRALIA • 1 ♀; “Cairns, Q., Austr. 7, 1904”, “Holotype”, “*Olonia picea* Kirk. ♀ 308”; BPBM • 1 ♀; “Cairns, Q., Austr. 7, 1904”, “Paratype” (BPBM); 2 ♂♂, “Cairns, Q., Austr. 8-1904”, “♂”, “Paratype”; BPBM.

Note

One of the two male paralectotypes is actually a male of *O. maura* (Fabricius, 1775).

Additional material

AUSTRALIA • **Queensland** – 1 ♂; Edge Hill, NW of Cairns; 16°54'06" S, 145°44'35" E; 30 Jul. 1969; James E. Tobler leg.; CAS • 1 ♂; same collection data as for preceding; 20 Aug. 1969; RBINS • 1 ♀; same collection data as for preceding; CAS • 1 ♂; Cairns; 12 Aug. 1969; CAS • 1 ♂; Cairns; 26 Feb.–1 Mar. 1985; W. Middlekauff leg.; CAS • 1 ♂; Cairns, Pine Creek; 18 Jan. 1962; E.B. Britton and G. Wilson leg.; BMNH • 1 ♂; 19 km NE of Mareeba; [coordinates of Mareeba: 16°59'42" S, 145°25'23" E]; 7 Jan.–12 Feb. 1985; Storey and Halfpapp leg.; interception trap; site n° 26; QPIM • 1 ♂; Kuranda; 16°49'11" S, 145°38'12" E; 28 Apr. 1957; W.W. Wirth leg.; USNM.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 6): 7.2 mm (6.8–7.4); ♀ (n = 1): 7.2 mm; Ltg/BTg (♂) = 2.3.

Male

HEAD (Fig. 41A–D). Vertex slightly concave, with anterior and posterior margins parallel, curved; medium to dark brown; obsolete median carina shortly marked along posterior margin. Frons uniformly coloured, medium to dark brown, sometimes darker at lateral angles. Clypeus elongate, entirely medium to dark brown. Genae brown to black with yellowish markings along anterior margin. Labium brown

to black, reaching metacoxae. Antennae black-brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

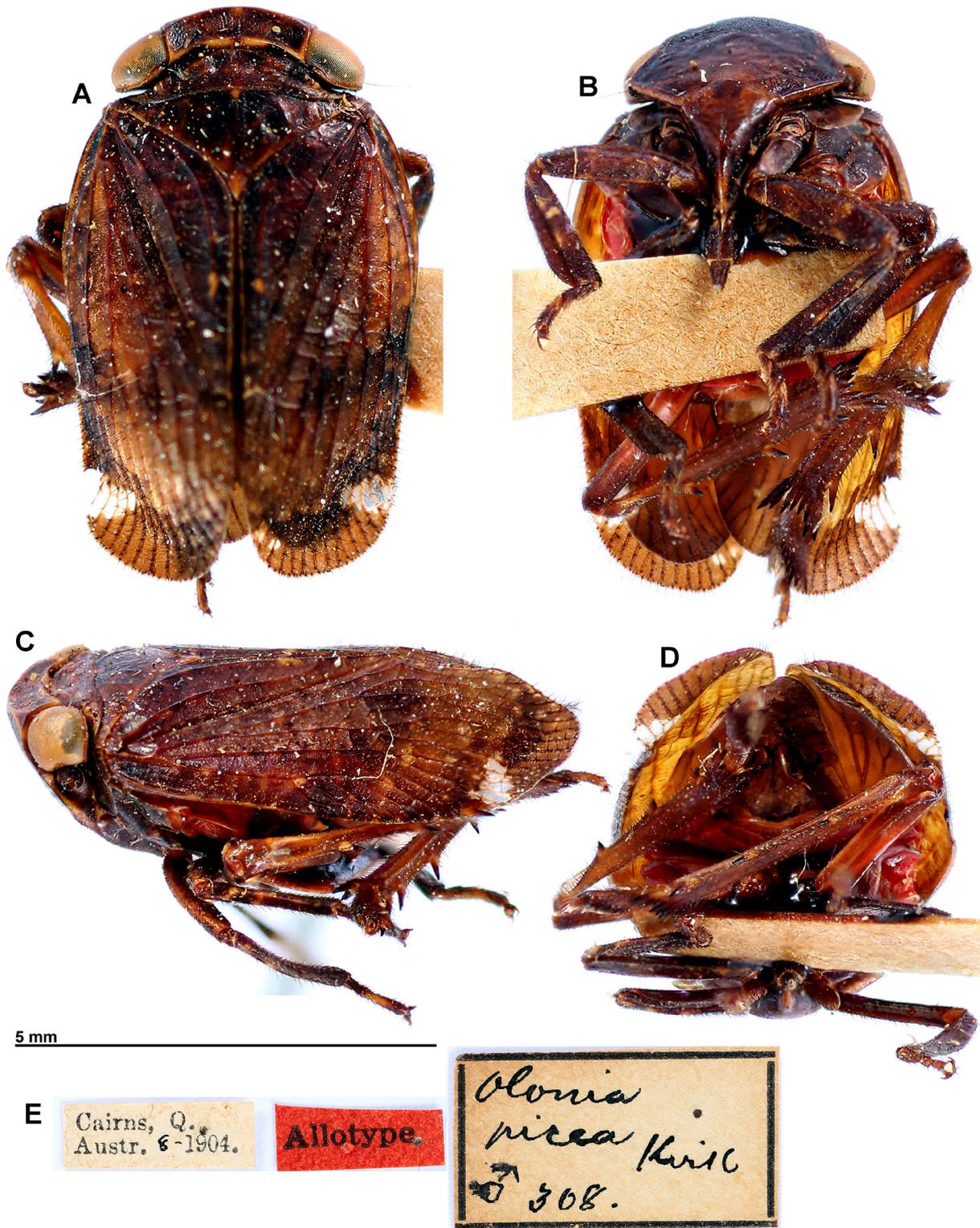


Fig. 40. *Olonia picea* Kirkaldy, 1906, lectotype, ♂. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Labels.

THORAX (Fig. 41A–D). Pronotum uniformly medium to dark brown; slightly wrinkled; 2 small impressed points on disc slightly marked. Lateral fields of prothorax coloured as pronotum. Mesonotum medium to dark brown variegated with darker zones on middle and sides; minute yellowish spot at apex of scutellum; median and peridiscal carinae weakly marked; median carina ending before scutellum; slight impression before scutellum. Red ventrally. Tegulae medium to dark brown.

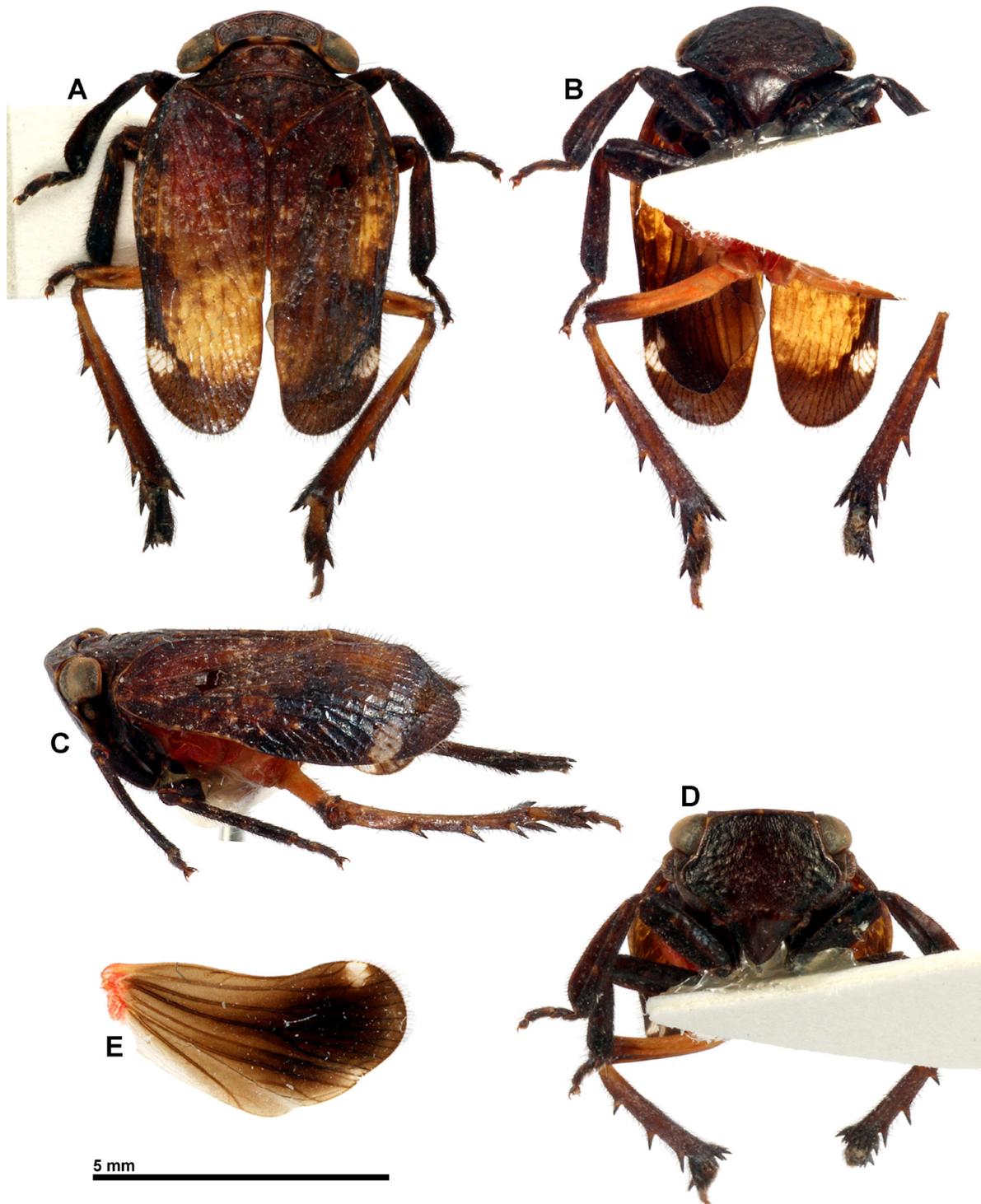


Fig. 41. *Olonia picea* Kirkaldy, 1906, ♂. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

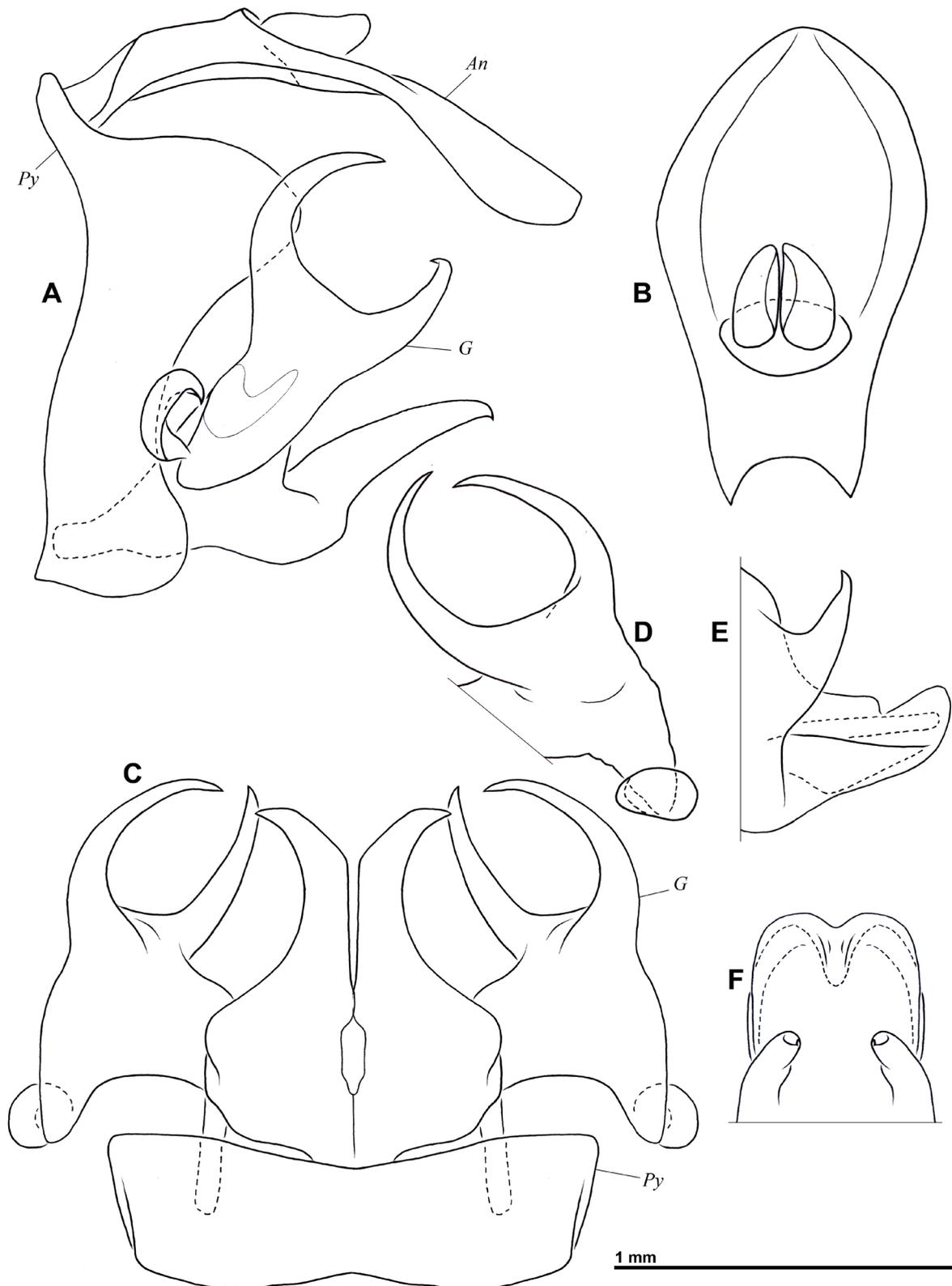


Fig. 42. *Olonia picea* Kirkaldy, 1906, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

TEGMINA (Fig. 41A, C). Medium to dark brown; often pale yellowish marking on vein A1 at midlength of clavus; marked with dark brown or black along costal margin, more broadly so on posterior half, and along posterior margin. Often darker, median, irregular marking at apical $\frac{2}{3}$; triangular white marking on costal margin on nodal line, sometimes reduced; sometimes a number of minute white spots at

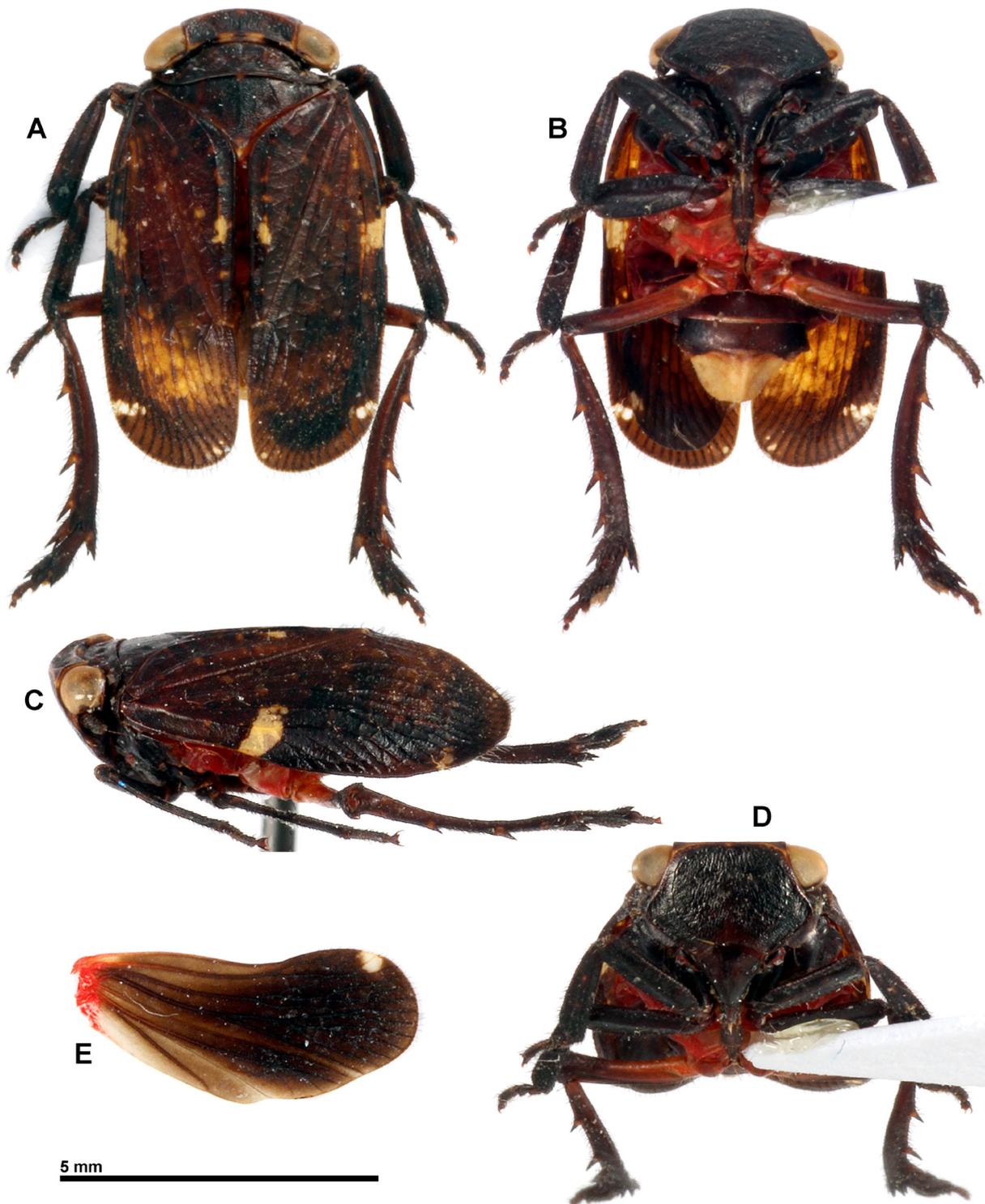


Fig. 43. *Olonia picea* Kirkaldy, 1906, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

apicosutural angle. Costal and sutural margins subparallel; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 41E). Brown, paler on anal area and progressively darker on apical half; small, somewhat rounded white marking at apicocostal angle, extending on 2–3 cells. Margin of anal area slightly sinuate; sutural margin with 1 slight cleft, cubital one nearly not marked.

LEGS (Fig. 41A–D). Pro- and mesocoxae black-brown. Pro- and mesofemora black-brown variegated with brown. Pro- and mesotibiae medium to dark brown, sometimes with 3 paler obsolete rings. Pro- and mesotarsi black-brown, with basal half of third tarsomere paler. Metacoxae reddish; metafemora reddish with apex brown. Metatibiae brown, with 3 lateral spines paler basally and 9 apical black-brown spines. Metatarsi brown, with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 42). Posterior margin of pygofer in lateral view strongly sinuate, strongly roundly projecting at dorsal $\frac{1}{3}$, moderately broad ventrally (Fig. 42A, C). Anal tube obovate, 1.9 times as long as broad, narrowing at level of epiproct, slightly curved ventrally in lateral view; lateral margins slightly curved ventrally on apical half; apical margin roundly pointed (Fig. 42A–B). Gonostyli fused on basal half of length of centroventral part and projecting posteriorly (Fig. 42A, C). Centroventral part broad and dorsoventrally flattened on basal half, then narrowing into an elongate, curved, spinose process directed laterally and ending in ventrally curved point (Fig. 42A, C). Laterodorsal part of gonostyli strong and curved dorsocentrally, slightly surpassing level of centroventral part ventrally, bifid, with a dorsal and a ventral process elongate and pointing apically, and forming a C together; lateral process elongate, projecting posterolaterally and longer than spoon-shaped process (Fig. 42A, C–D). Dorsal portion of phallobase with hooked process on each side, progressively narrowing from base to apex, directed posterodorsally and with apex pointing centrally (Fig. 42E–F). Ventral portion of phallobase subquadrate, with apical margin emarginate in dorsal view and with median lobe not surpassing phallus (Fig. 42E–F). Phallus dorsoventrally flattened, rather broad, with apical margin emarginate in middle (Fig. 42E–F).

Female

Similar to male but with white spot on costal margin of tegmina at nodal line reduced, and sometimes with white, transverse marking on costal margin of tegmina slightly before half length and extending to vein RP (Fig. 43).

Distribution and biology

This species has been recorded from an area around Cairns in North Queensland (Fig. 3), in the Einasleigh Upland Savanna and Queensland Tropical Rainforests bioregions. Specimens were collected in January, February, March, April, July, August and September. No host plant known.

***Olonia rubicunda* (Walker, 1851)**

Figs 3, 44

Eurybrachys rubicunda Walker, 1851: 391 (described).

Eurybrachys rubicunda – Kirkaldy 1906: 445 (listed as belonging to *Olonia*).

Olonia rubicunda – Stål 1862: 488 (transferred to *Olonia*). — Distant 1906: 206 (type species of *Olonia*).
— Metcalf 1956: 65 (catalogued).

non *Olonia rubicunda* – Jacobi 1928: 4 ((re)described from Kimberley District (erroneous, based on misidentified specimens)). — Lallemand 1935: 675 (mentioned from Northern Territory (erroneous, based on misidentified specimens)).

Diagnosis

The species can be recognized by the following combination of characters (♀):

- (1) hind wings without orange marking (Fig. 44B)
- (2) pro- and mesofemora and tibiae largely black-brown (Fig. 44C, E)
- (3) tegmina brown, darker along costal and apical margins (Fig. 44B)
- (4) rather small size: 8.5 mm

This species is currently known only from a single female and the diagnosis will need to be augmented with characters of the male genitalia when male specimens become available.

Etymology

The species epithet *rubicundus* (adjective, Latin) means ‘bright red’. It refers to the colour of the abdomen in this species.

Material examined

Holotype

AUSTRALIA • ♀; Queensland, Sandy Cape; [24°43'46" S, 153°12'30" E]; “N.H., Sandy Cape; / on the reverse / 46 73”, “Type”, “22. *Eurybrachys* [sic] *rubicunda*”, “Re-pinned on stainless”; left anterior and posterior, and right median legs glued on labels attached to the pin of the specimen, left tegmen missing; BMNH (Fig. 44).

Note

“N.H.” on the first label stands for New Holland, a former name for Australia.

Description

MEASUREMENTS AND RATIOS. LT: ♀ (n = 1): 8.5 mm (extrapolated); BV/LV = 4.25; BF/LF = 1.76; LP + LM/BT not measurable; LTg/BTg = 2.5; LW/BW not measurable.

Female

HEAD (Fig. 44A–E). Vertex concave, with anterior and posterior margins parallel, curved; brown with darker marking at lateral angles. Frons uniformly reddish brown. Clypeus elongate, entirely reddish brown. Genae yellowish with elongate brown marking under eye. Labium black-brown, reaching metacoxae. Antennae black-brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 44A–E). Pronotum brown and slightly wrinkled; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax brown. Mesonotum brown variegated with blackish; peridiscal carinae weakly marked. Red ventrally. Tegulae brown. (Mesonotum damaged in the examined specimen.)

TEGMINA (Fig. 44B). Brown with small yellowish spots; pale yellowish marking on vein A1 at $\frac{2}{3}$ of clavus; marked with black along costal margin, more broadly so on posterior half, and along posterior margin. Triangular white marking on costal margin on nodal line; yellowish markings at apicosutural angle. (Remaining right tegmen of examined specimen damaged.)

POSTERIOR WINGS (Fig. 44B). Brown, paler on anal area and with large darker area reaching apical margin; elongate, transverse, subtriangular white marking at apicocostal angle, extending on 3 cells. Margin of anal area slightly sinuate; sutural margin with 1 cleft, cubital one not marked.

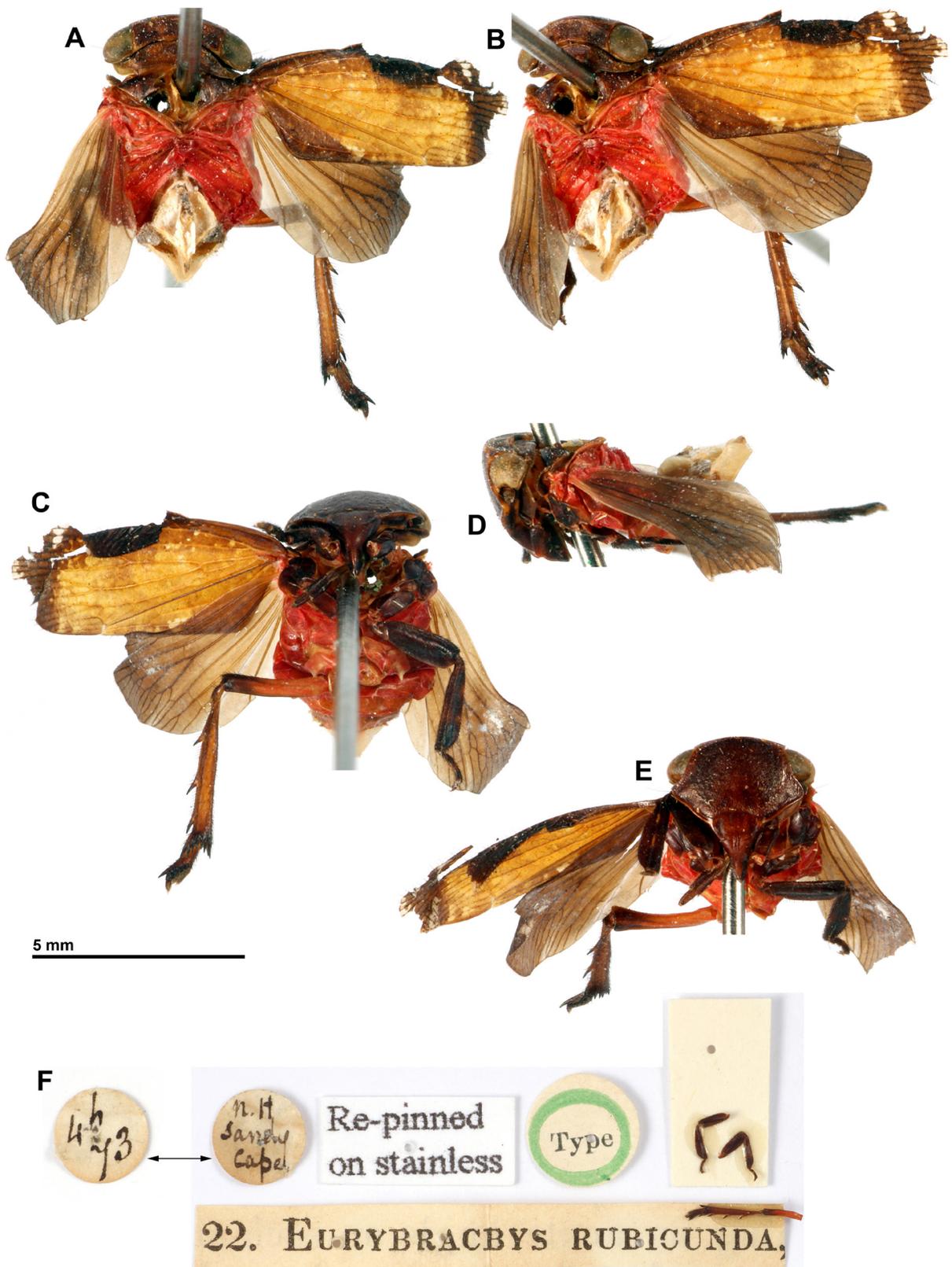


Fig. 44. *Olonia rubicunda* (Walker, 1851), holotype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, normal view of right tegmen. **C.** Ventral view. **D.** Habitus, lateral view. **E.** Habitus, normal view of frons. **F.** Labels (with three legs).

LEGS (Fig. 44A, C, E). Pro- and mesocoxae dark brown. Pro- and mesofemora black-brown with reddish spots marking obsolete rings. Pro- and mesotibiae black-brown with reddish markings on 3 obsolete rings. Pro- and mesotarsi black-brown, with basal half of third tarsomere paler. Metacoxae red; metafemora reddish with apex brown. Metatibiae brown, with 3 lateral spines paler basally and 8 apical black-brown spines. Metatarsi brown, with a ventral row of 6 black spines on first tarsomere.

ABDOMEN (FIG. 44A–D). Bright red with genital segments brown and sternite VII white.

Male

Unknown.

Distribution and biology

This species is known only from a single specimen from Fraser Island in southeastern Queensland (Fig. 3), in the Eastern Australian Temperate Forests bioregion. Biology unknown.

Olonia rylandae sp. nov.

[urn:lsid:zoobank.org:act:1F0498E0-6DD9-48AD-AB8D-2369364963CE](https://doi.org/10.1111/zoobank.org/act:1F0498E0-6DD9-48AD-AB8D-2369364963CE)

Figs 3, 45–46

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings without orange marking (Fig. 45E)
- (2) pro- and mesofemora and -tibiae mostly brown (Fig. 45A–D)
- (3) anal tube of male oblong, with posterior margin slightly concave and lateral margins sinuate (Fig. 46B)
- (4) centroventral part of gonostyli with elongate and narrow process strongly sinuate basally and with small apical hook pointing ventrally (Fig. 46A, C)
- (5) laterodorsal part of gonostyli with elongate and narrow process slightly curved lateroventrally (Fig. 46A, C–D)
- (6) rather small size: 7.7 mm

Etymology

This species is dedicated to Mrs Valry Ryland (Magnetic Island, Queensland, Australia) in acknowledgement for all her help and involvement in documenting the natural history of species of *Olonia*.

Material examined

Holotype

AUSTRALIA • ♂; Queensland, Bathurst Head; [14°16' S, 144°12' E]; “Bathurst Head, Q., Jan. 1927, Hale & Tindale”; dissected, right anterior leg missing, right posterior wing mounted; SAM.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 1): 7.7 mm; BV/LV = 4.38; BF/LF = 1.7; LP+LM/BT = 0.67; LTg/BTg = 2.5; LW/BW = 2.0.

Male

HEAD (Fig. 45A–D). Vertex concave, with anterior and posterior margins parallel, curved; brown, slightly wrinkled. Frons uniformly brown, slightly wrinkled. Clypeus elongate, brown. Genae yellowish with brown markings around eyes and under antennae. Labium dark brown, reaching metacoxae. Antennae black-brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

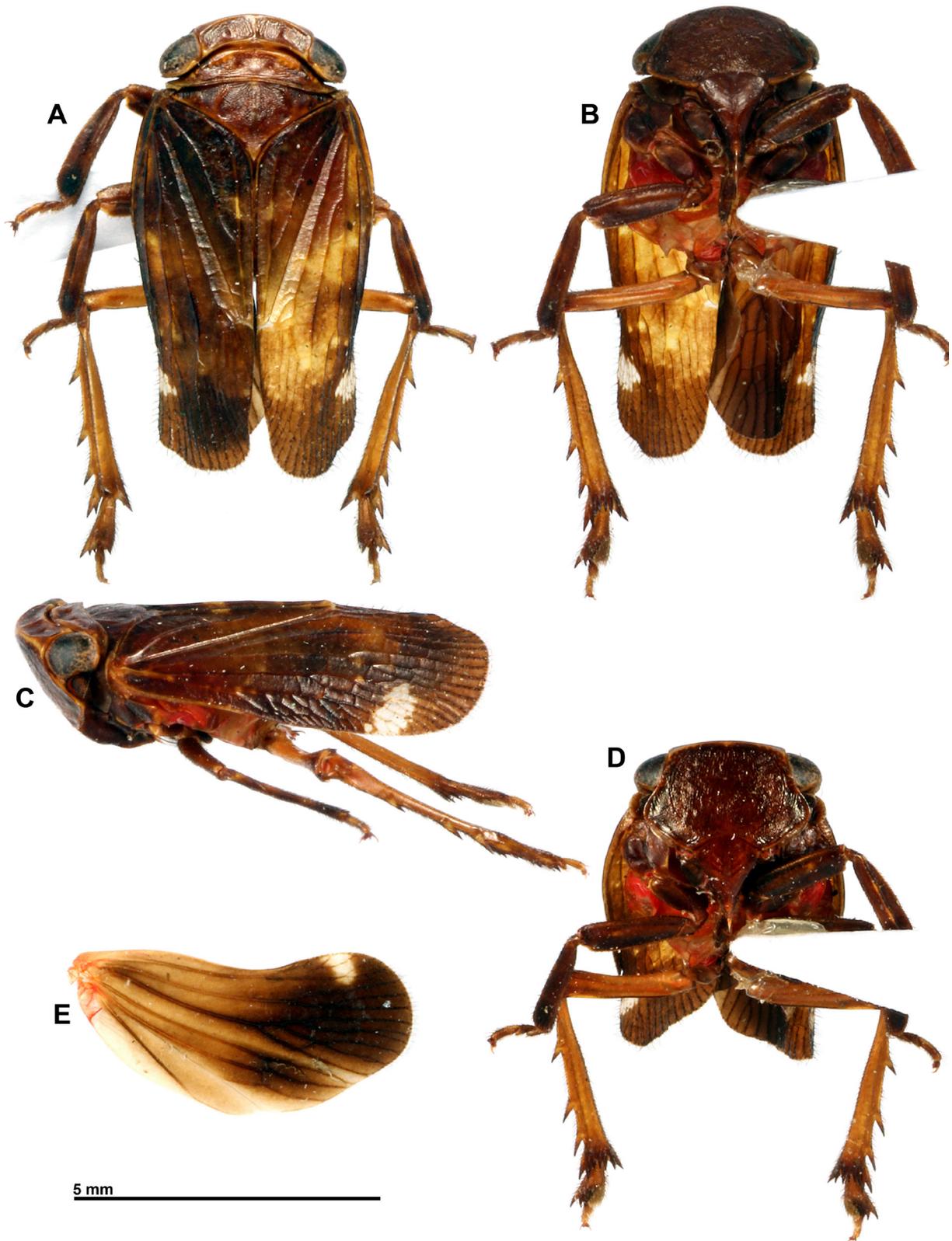


Fig. 45. *Olonia rylandae* sp. nov., holotype, ♂. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Posterior wing.

THORAX (Fig. 45A–C). Pronotum brown with small yellowish spot on each side; slightly wrinkled; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax brown. Mesonotum brown; yellowish spot on middle of anterior margin and at apex of scutellum; slightly rugulose; median and peridiscal carinae weakly marked; median carina stopping before scutellum; slight impression before scutellum. Red ventrally. Tegulae brown.

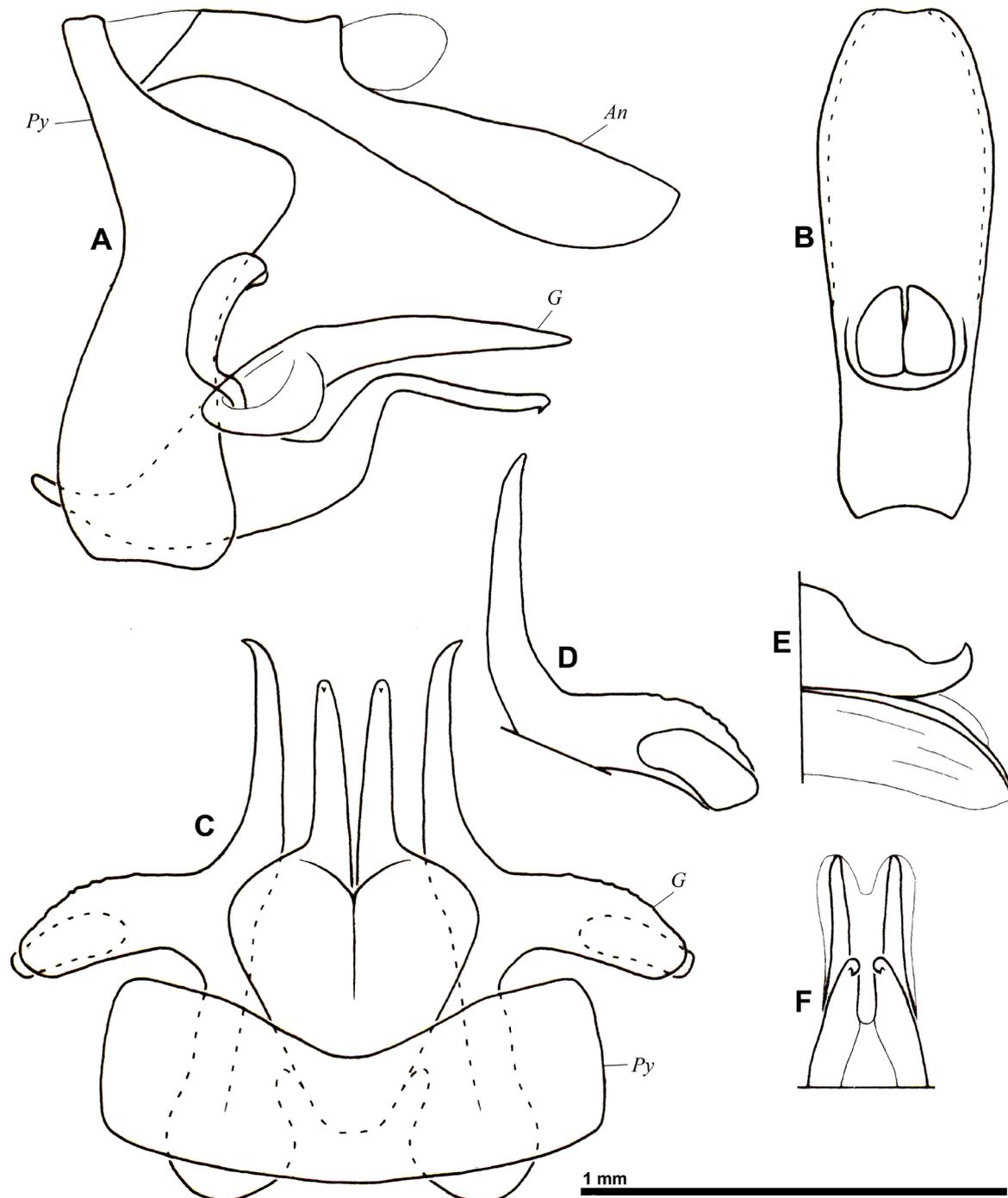


Fig. 46. *Olonia rylandae* sp. nov., holotype, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

TEGMINA (Fig. 45A, C). Brown, slightly variegated with yellowish; pale yellowish marking on vein A1 at midlength of clavus; marked with black along costal margin, more broadly so on posterior half, and along posterior margin. Triangular white marking on costal margin on nodal line; no white spot at apicosutural angle. Costal and sutural margins subparallel; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 45E). Brown, paler on anal area and with large blackish area reaching apical margin; elongate, transverse, subtriangular white marking at apicocostal angle, extending on 3–4 cells. Margin of anal area slightly sinuate; sutural margin with 1 cleft, cubital one nearly not marked.

LEGS (Fig. 45A–D). All coxae brown. Pro- and mesofemora dark brown. Pro- and mesotibiae brown, turning blackish apically, and with 3 very obsolete paler rings. Pro- and mesotarsi brown, with basal half of third tarsomere paler. Metafemora pale brown with apex darker. Metatibiae brown, with 3 lateral spines paler basally and 8 apical black-brown spines. Metatarsi brown, with a ventral row of 6 black spines on first tarsomere.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 46). Posterior margin of pygofer in lateral view strongly sinuate, rather angularly, roundly projecting at dorsal $\frac{1}{3}$ and rather broad ventrally (Fig. 46A, C). Anal tube oblong, 2.9 times as long as broad, broader at apical $\frac{2}{3}$, with lateral margins sinuate and apical margin concave; lateral margins curved ventrally on apical $\frac{2}{3}$; slightly curved ventrally near base in lateral view (Fig. 46A–B). Gonostyli fused on slightly less than basal half of length of centroventral part and projecting posteriorly (Fig. 46A, C). Centroventral part broad and dorsoventrally flattened on basal half, then strongly narrowing into elongate, narrow process, strongly sinuate basally in lateral view and ending in narrow hook curved anteroventrally (Fig. 46A, C). Laterodorsal part of gonostyli elongate and slightly curved posteroventrally, posteriorly slightly surpassing level of centroventral part; lateral process elongate, projecting posterolaterally, slightly concave dorsally and longer than spoon-shaped process (Fig. 46A, C–D). Dorsal portion of phallobase with hooked process on each side, narrowing in 2 steps from base to apex, directed posterocentrally and with apex pointing dorsally (Fig. 46E–F). Ventral portion of phallobase elongate and narrow, curved posteroventrally (Fig. 46E–F). Phallus dorsoventrally flattened, medially concave, broadening towards apex and emarginate apically (Fig. 46E–F).

Female

Unknown.

Distribution and biology

This species is only known from a single male from Bathurst Head, a headland covered with open forest close to Cape Melville on the eastern coast of Cape York Peninsula (Fig. 3), in the Cape York Peninsula Tropical Savanna bioregion. It was collected in January, nearly one century ago.

Olonia soulierae sp. nov.

[urn:lsid:zoobank.org:act:6670A1E7-F89F-4D93-901E-49DD861B6539](https://doi.org/10.21203/rs.3.rs-1111111/v1)

Figs 3, 47–49

Diagnosis

This species can be recognized by the following combination of characters:

- (1) hind wings with orange marking (Figs 47E, 49E)
- (2) pro- and mesofemora and -tibiae largely black-brown (Figs 47A–D, 49A–D)
- (3) anal tube of male narrow and parallel-sided on basal $\frac{1}{3}$, then ovate with posterior margin notched (Fig. 48B)

- (4) centroventral part of gonostyli with long and narrow process, slightly sinuate basally and with apical hook pointing dorsally (Fig. 48A, C)
- (5) laterodorsal part of gonostyli with hooked process curved lateroventrally (Fig. 48A, C–D)
- (6) large size: 9.4–11.2 mm

Etymology

This species is dedicated to Dr Adeline Soulier-Perkins (MNHN), who collected a part of the type series.

Material examined

Holotype

AUSTRALIA • ♂; Queensland, near Chillagoe Haunted Cave; 17°06' S, 144°25' E; “Muséum Paris, Australie (Queensland) Près de Chilagoe Hounted Cave - Cave 1”, “S17°06' E144°25', 13.III.1997, A. Soulier-Perkins & Th. Bourgoïn rec..”, “QM-244702”; QM.

Paratypes

AUSTRALIA • 2 ♂♂, 4 ♀♀; same collection data as for holotype; MNHN • 1 ♂, 1 ♀; same collection data as for holotype; RBINS • 1 ♀; “Australia: Qld., Chillagoe Caves Nat. Park, turnoff to Royal Arch Cave, Au 97 – 49, 60, 63, M. Asche & H. Hoch, 19.III.1997”; ZMHB.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 3): 9.8 mm (9.4–10.1); ♀ (n = 5): 10.8 mm (10.5–11.2); BV/LV = 4.0; BF/LF = 1.71; LP+LM/BT = 0.71; LTg/BTg = 2.1–2.3; LW/BW = 1.72.

Male

HEAD (Fig. 47A–D). Vertex concave, with anterior and posterior margins parallel, curved; dark brown variegated with yellowish in middle. Frons black-brown, slightly tinged with 2 transverse, irregular, variegated yellowish markings, more dorsal one not reaching sides. Clypeus elongate, dark reddish brown with 2 short, black, longitudinal lines at base, with apex darker. Genae yellowish with brown markings around eyes and under antennae. Labium brown, reaching metacoxae. Antennae black; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 47A–C). Pronotum dark brown with some yellowish markings laterally; slightly transversely wrinkled posteriorly; obsolete median carina and 2 small impressed points on disc. Lateral fields of prothorax brown. Mesonotum dark brown with reddish markings along posterior margin, median yellowish spot along anterior margin and at apex of scutellum; slightly rugulose; median and peridiscal carinae weakly marked; median carina ending before scutellum; slight impression before scutellum. Red ventrally. Tegulae brown.

TEGMINA (Fig. 47A, C). Dark brown with small yellowish or reddish spots; bigger, whitish, slightly transverse spot on middle of clavus; large triangular white marking on costal margin on nodal line and a smaller one anterior to it; white marking at apicosutural angle. Maximum breadth at nodal line; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 47E). Brown with apical $\frac{1}{3}$ black-brown; transverse, subtriangular white marking at apicocostal angle, extending on 5 cells; sutural margin broadly bordered with yellowish on basal half; orange marking between anal fold and vein CuA at about half length. Margin of anal area sinuate; sutural margin with 2 clefts, cubital one slightly marked.

LEGS (Fig. 47A–D). Pro- and mesocoxae reddish brown. Pro- and mesofemora black-brown with yellowish spots marking 2 obsolete rings. Pro- and mesotibiae black-brown with 3 obsolete ring-shaped yellowish markings, larger one near apex. Pro- and mesotarsi dark brown, with basal half of last segment

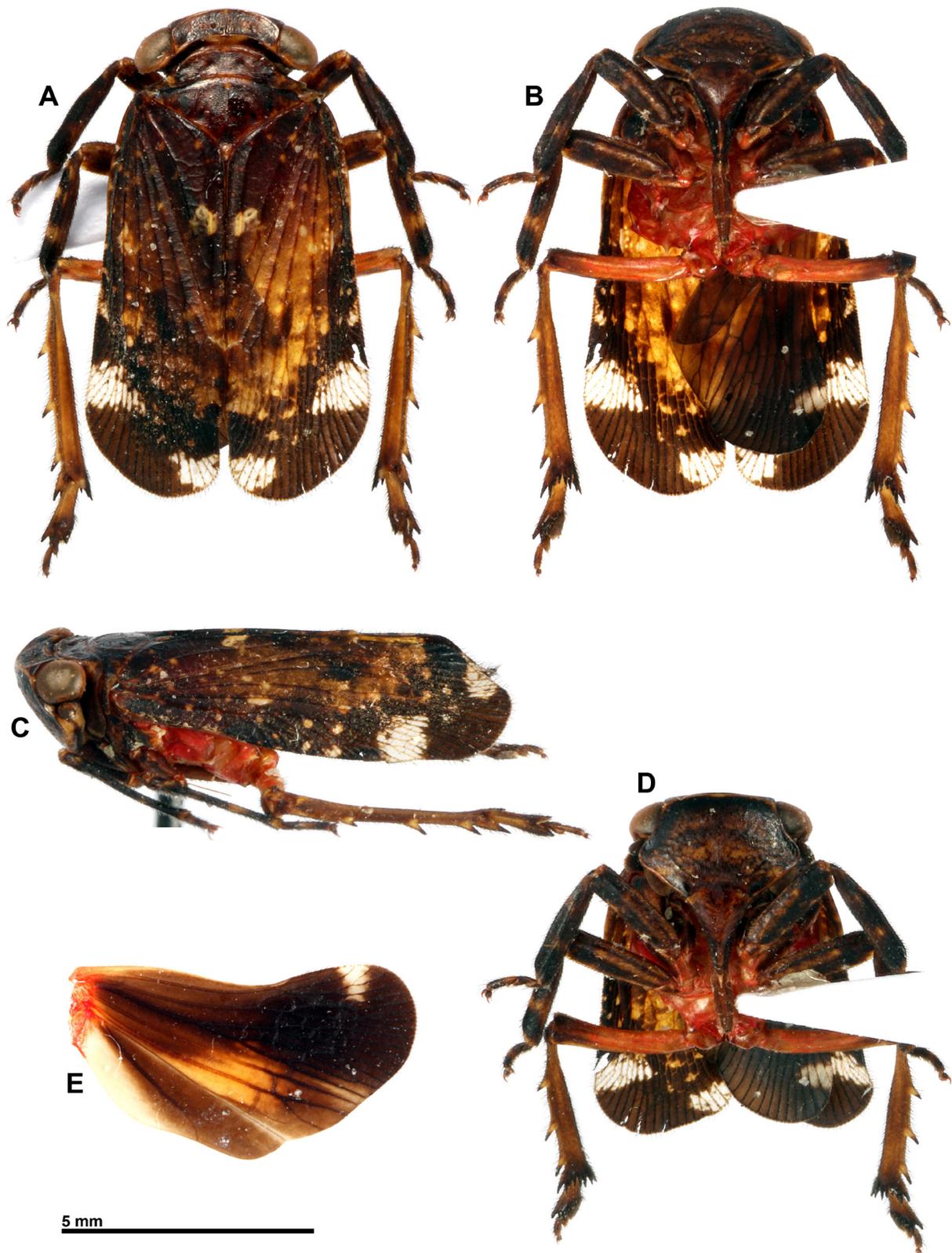


Fig. 47. *Olonia soulierae* sp. nov., holotype, ♂. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Posterior wing.

yellow-brown. Metacoxae reddish; metafemora reddish with apex brown. Metatibiae brown, with 3 lateral spines yellowish basally and 8 apical, black-brown spines. Metatarsi brown with a ventral row of 6 black spines on first tarsomere.

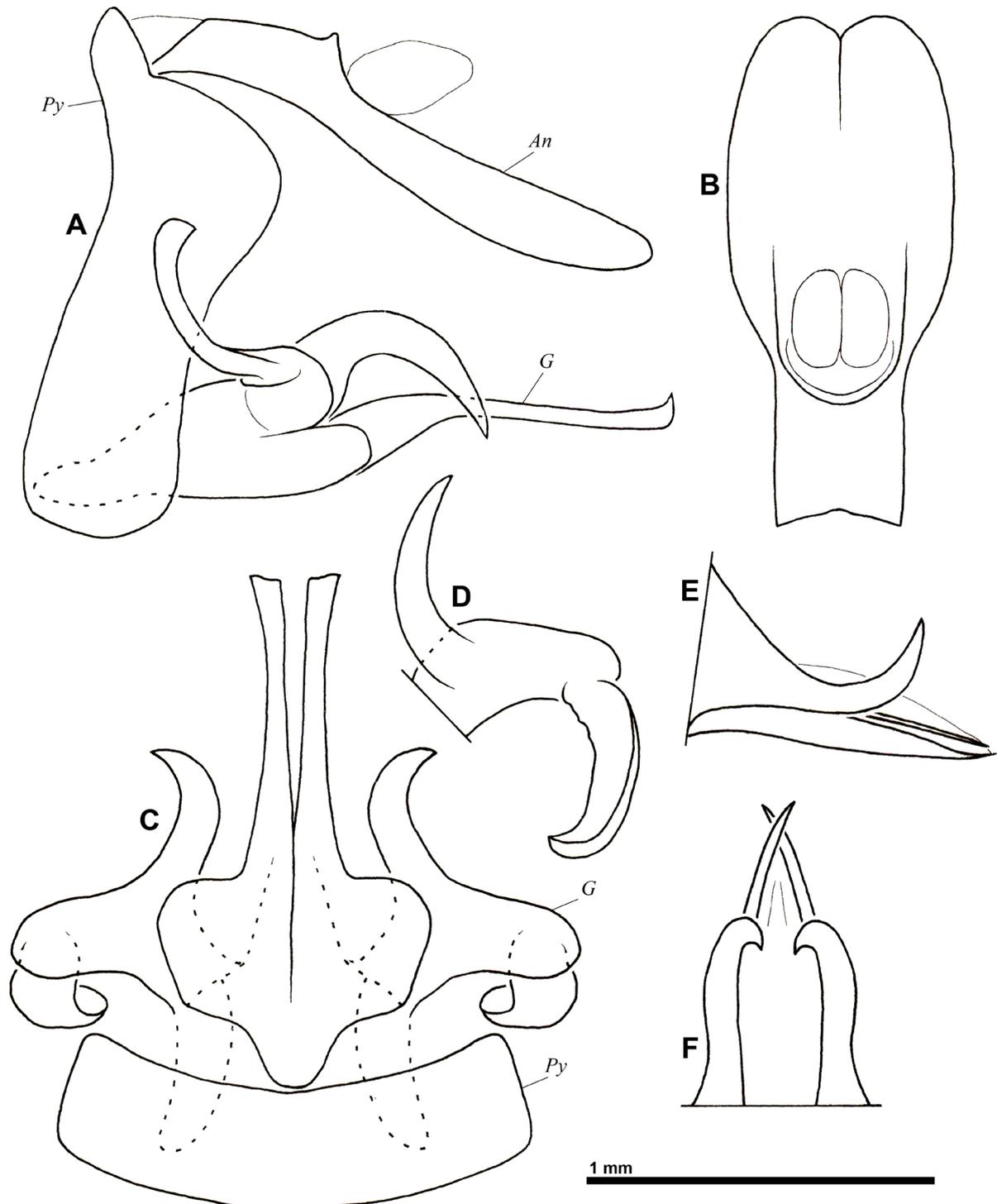


Fig. 48. *Olonia soulierae* sp. nov., holotype, ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

ABDOMEN. Bright red with genital segments black-brown.

MALE GENITALIA (Fig. 48). Posterior margin of pygofer in lateral view strongly sinuate, roundly projecting at dorsal $\frac{1}{3}$, rather broad ventrally (Fig. 48A, C). Anal tube elongate, 2.25 times as long as broad, with

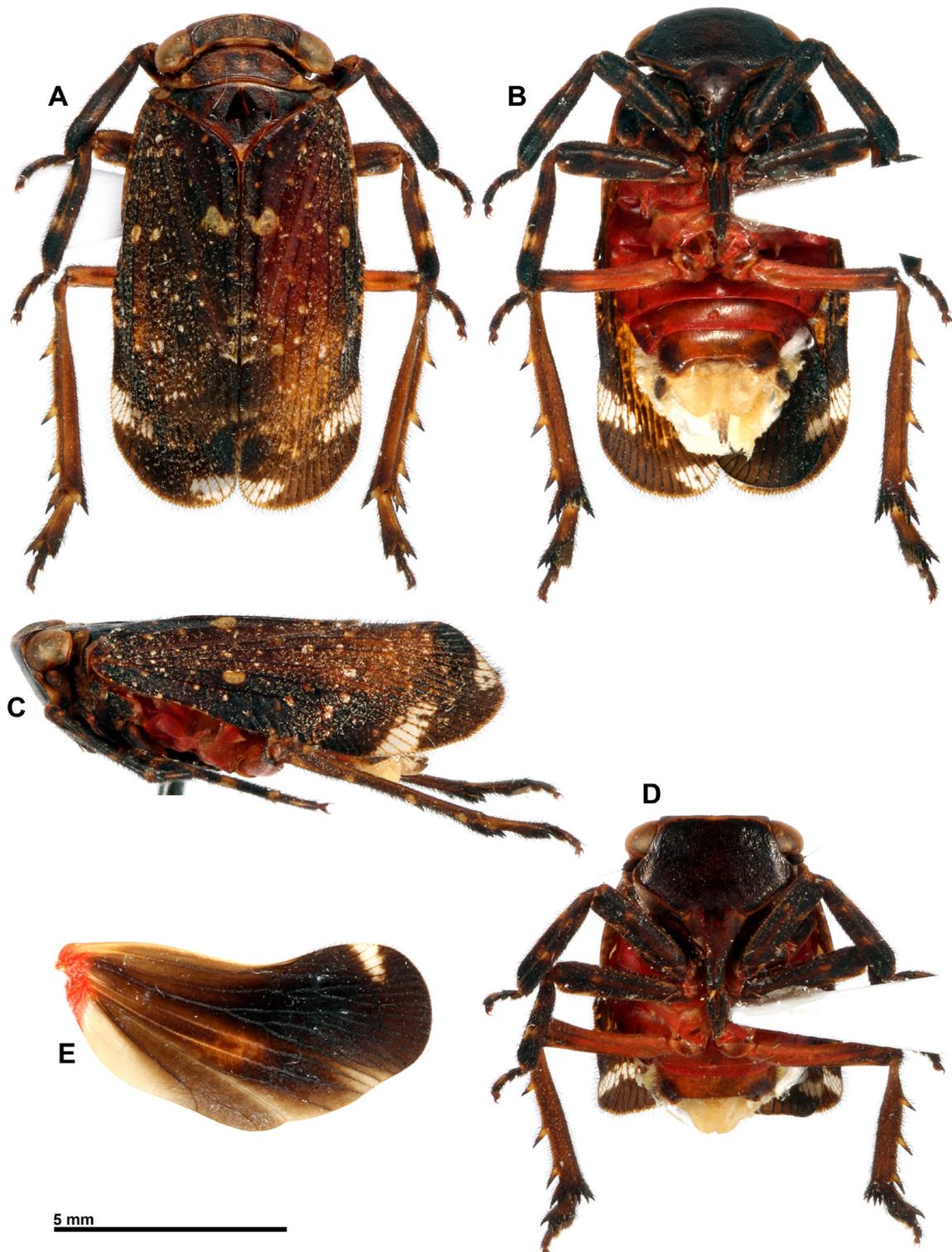


Fig. 49. *Olonia soulierae* sp. nov., paratype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, lateral view. **D.** Habitus, normal view of frons. **E.** Posterior wing.

basal $\frac{1}{3}$ parallel-sided and narrower, then broadly ovate with posterior margin notched; slightly sinuate in lateral view; lateral margins slightly curved ventrally on apical $\frac{2}{3}$ (Fig. 48A–B). Gonostyli fused to nearly half length of centroventral part and projecting posteriorly (Fig. 48A, C). Centroventral part dorsoventrally flattened on basal $\frac{1}{3}$, then abruptly narrowing and forming strongly elongated and narrow process, slightly diverging and ending in a small hook pointing dorsally; process slightly sinuate in lateral view (Fig. 48A, C). Laterodorsal part of gonostyli curved lateroventrally, hooked, with lateral process rather broad and moderately elongated, about as long as spoon-shaped process (Fig. 48A, C–D). Dorsal portion of phallobase with elongate process on each side, strongly narrowing on basal half in lateral view, then strongly hooked, pointing dorsally and slightly incurved (Fig. 48E–F). Ventral portion of phallobase along lateral margin of phallus on basal half, elongate and supassing phallus, crossing after it (Fig. 48E–F). Phallus dorsoventrally flattened, elongate, narrowing from base to apex (Fig. 48E–F).

Female

Similar to male, but darker; frons entirely black-brown; white markings on tegmina smaller; black-brown area of posterior wings larger, orange marking reduced (Fig. 49).

Distribution and biology

This species is known from a series of ten specimens collected in Chillagoe in North Queensland (Fig. 3), in the Einasleigh Upland Savanna bioregion. Nine specimens were collected on a single day in March; hence, the species is probably not scarce in its habitat. No host plant is recorded.

Genus *Stalobrachys* gen. nov.

[urn:lsid:zoobank.org:act:7BB4E8DC-4BF9-4A5B-B653-76EC7BD66201](https://doi.org/10.21203/rs.3.rs-1888881/v1)

Figs 50–51

Type species

Olonia alboapicata Jacobi, 1928, here designated.

Diagnosis

Rather small sized (8.5–10 mm), dark brown variegated with black and yellowish, tegmina with a white marking along costal margin on nodal line and posterior wings usually with a white marking near externoapical angle in males and with apex white in females. The genus can be recognized by the following set of characters:

- (1) posterior wings with anal area well developed, broad (LW/BW = 1.5) (Fig. 50A)
- (2) posterior margin of pygofer with strongly developed projection at laterodorsal angle
- (3) gonostyli moderately sclerified, separated and with elongate basodorsal process bearing dorsoapical, articulate, spoon-shaped process
- (4) aedeagus reduced, with dorsal portion of phallobase bearing two convergent elongate spinose processes

The genus shares having a spoon-shaped process on the gonostyli with the Australian genera *Chewobrachys* Constant, 2008, *Fletcherobrachys* Constant, 2006, *Hackerobrachys* Constant, 2006, *Maeniana* Metcalf, 1952, *Nirus* Jacobi, 1928 and *Olonia* Stål, 1862, but differs

– from *Chewobrachys* by the smaller size (8.5–10 mm; 12–16 mm in *Chewobrachys*), the dark brown colour (greyish brown in *Chewobrachys*) and the male gonostyli with an elongate basodorsal process (gonostyli without basodorsal process in *Chewobrachys*)

– from *Fletcherobrachys* by the bright red abdomen (green to orange in *Fletcherobrachys*), the hind wings being brown with a (sub-)apical white marking (basal half white in *Fletcherobrachys*), the absence of sexual dimorphism on the median tibiae (median tibiae with externodistal process in females of

Fletcherobrachys) and the male gonostyli separated and with an elongate basodorsal process (gonostyli fused along most of their length and without a basodorsal process in *Fletcherobrachys*)

– from *Hackerobrachys* by the frons being brown and slightly convex (bright yellow to red and strongly convex in *Hackerobrachys*), the posterior wings being brown with a (sub-)apical white marking and rounded apically (uniformly brown with apex subquadrate in *Hackerobrachys*) and the gonostyli being separated, with an elongate basodorsal process (gonostyli fused along most of their length and without a basodorsal process in *Hackerobrachys*)

– from *Maeniana* by the posterior wings having the anal area well developed (anal area of posterior wings reduced in *Maeniana*) and showing a characteristic sexual dimorphism (no sexual dimorphism in the posterior wings in *Maeniana*)

– from *Nirus* by the frons being slightly convex (concave in *Nirus*), the pygofer with the posterior margin strongly projecting posteriorly at dorsal $\frac{2}{3}$ (not projecting in *Nirus*) and the gonostyli being separated and with an elongate basodorsal process (gonostyli fused and with a short basodorsal process in *Nirus*)

– from *Olonia* by the pygofer having the posterior margin with an elongate laterodorsal process projecting posteriorly (posterior margin only strongly sinuate in *Olonia*), the gonostyli separated (fused along about $\frac{1}{3}$ of their length in *Olonia*) and the gonostyli having an elongate basodorsal process bearing a spoon-shaped process apically (spoon-shaped process attached to the lateral process of the laterodorsal part of the gonostylus, which bears a spine or hook, in *Olonia*)

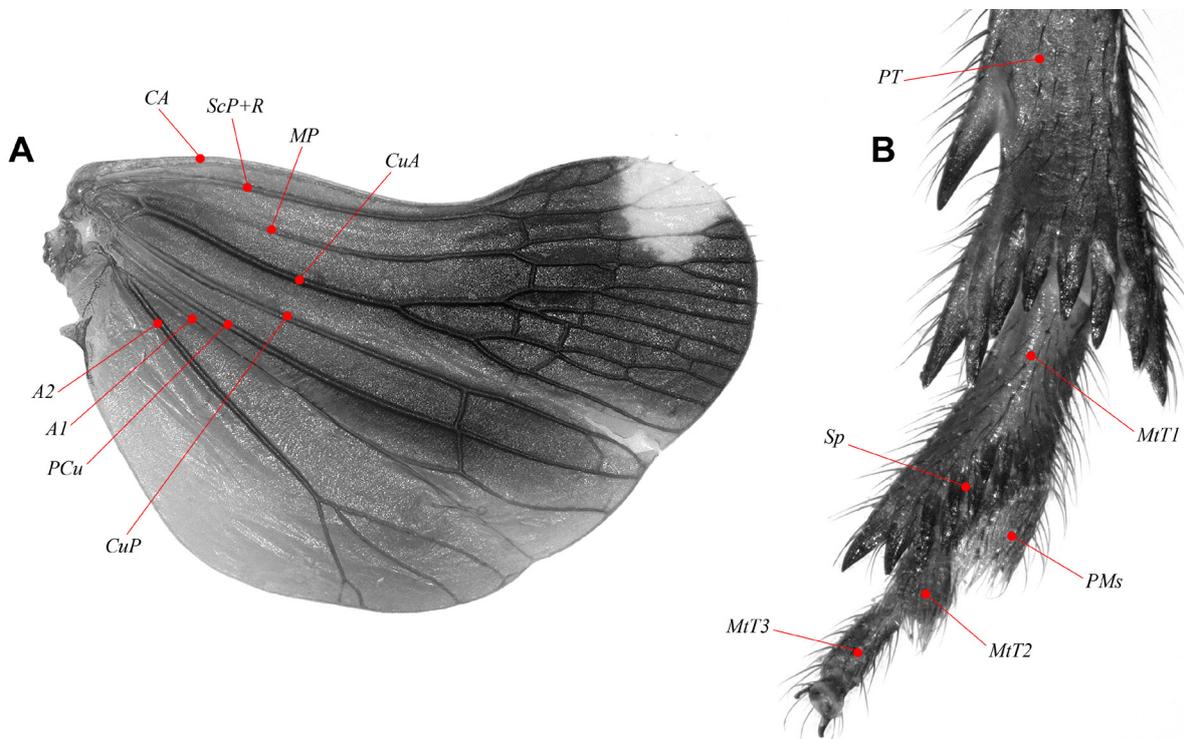


Fig. 50. *Stalobrachys* gen. nov. **A.** Right posterior wing, venation. **B.** Right posterior leg, distal portion, ventral view. Abbreviations: *A1* = first anal vein; *A2* = second anal vein; *CA* = costa anterior; *CuA* = cubitus anterior; *CuP* = cubitus posterior; *MP* = media posterior; *MtT1* = first metatarsomere; *MtT2* = second metatarsomere; *MtT3* = third metatarsomere; *PCu* = postcubitus; *PMs* = pad of microsetae; *PT* = posterior tibia; *ScP + R* = subcosta posterior + radius; *Sp* = spines.

The genus *Loisobrachys* Constant, 2008 is known from a single female and was placed by Constant (2008b) close to *Hackerobrachys*. Hence, the males of that genus potentially share having gonostyli with a spoon-shaped process. However, *Stalobrachys* gen. nov. can easily be separated from *Loisobrachys* by its slightly convex frons (frons strongly convex in *Loisobrachys*) and by the posterior wings having white markings (uniformly brown in *Loisobrachys*).

Etymology

The name of this genus is formed from ‘Stal’ in memory of the great contribution of the late Swedish entomologist Dr Carl Stål (1833–1878) to the study of Hemiptera, and *brachys* (Greek) = ‘short’, which is a common ending of generic names in Eurybrachidae. Gender feminine.

Description

COLOURATION. Head, pro- and mesonotum, and tegmina brown, from pale brown to nearly black, usually variegated with darker and paler. Tegmina with yellowish marking in middle of clavus on vein A1; triangular white marking along costal margin on nodal line; smaller white marking at apicosutural angle. Posterior wings black-brown, darker from base to distal part; with white subtriangular marking at apicocostal angle in males and with apex largely white in females. Pro- and mesofemora and pro- and mesotibiae with 2 paler rings; metafemora reddish, turning to brown apically; metatibiae brown. Abdomen and ventral face of thorax bright red; genital segments brown with ventral part of gonostyli white in males; abdominal segment VII white in females.

HEAD. AS broad as thorax; vertex 3.4–3.9 times as broad as long, concave, with all margins slightly carinate; anterior and posterior margins rounded, parallel; frons about 2.1 times as broad as long, slightly convex, slightly wrinkled to rugulose, with peridiscal carina slightly marked; upper margin of frons straight in normal view; clypeus slightly surpassing anterior trochanters, elongate, with median carina towards apex; labium reaching hind coxae, with apical segment longer than broad, acuminate, shorter and more slender than penultimate segment; no infra-ocular spines on genae; ocelli absent; antennae elongate, very slightly surpassing eye and visible from above; scape about as long as broad, pedicel elongate, barrel-shaped.

THORAX. About 1.55 times as broad as combined length of pro- and mesonotum; pronotum about half as long as mesonotum; pronotum with disc weakly wrinkled, smooth carina parallel to anterior margin and 2 slightly impressed points on disc; mesonotum with disc weakly wrinkled, median and peridiscal carinae slightly marked.

TEGMINA. Slightly convex; curving down at nodal line and with apex slightly curved upwards; elongate, about 2.15–2.5 times as long as broad; costal margin slightly sinuate; apical margin obliquely rounded; sutural margin slightly oblique after clavus. Venation: veins ScP+RA and RP separated close to base;

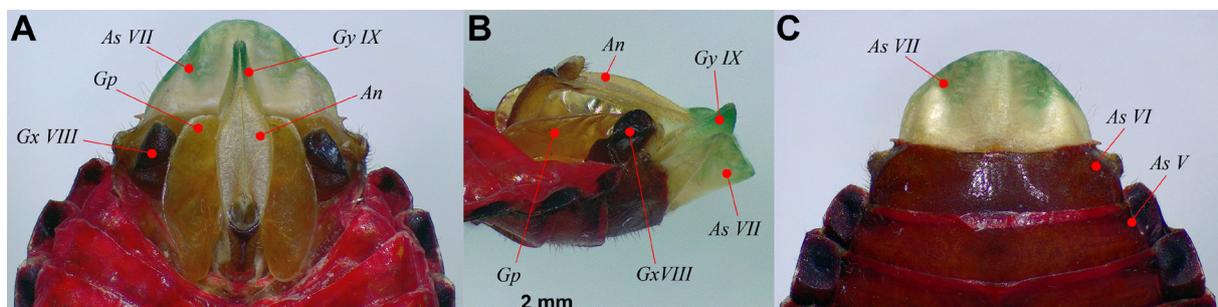


Fig. 51. *Stalobrachys* gen. nov., female genitalia. **A.** Dorsal view. **B.** Left lateral view. **C.** Ventral view. Abbreviations: *An* = anal tube; *As V* = fifth abdominal sternite; *As VI* = sixth abdominal sternite; *As VII* = seventh abdominal sternite; *Gp* = gonoplac; *Gx VIII* = gonocoxa VIII; *Gy IX* = gonapophysis IX.

first fork of MP on basal $\frac{1}{3}$; first fork of CuA slightly before apex of clavus; clavus closed; Pcu and A1 fused at $\frac{2}{3}$ of clavus length; Pcu+A1 reaching apical angle of clavus; numerous elongate cells along posterior half of costal margin and along apical margin.

POSTERIOR WINGS (Fig. 50A). Well developed, broader than tegmina, rounded apically; LW/BW = 1.5; anal area well developed; sutural margin very weakly trilobous; not reaching apex of tegmina at rest. All main veins visible from base, forked after nodal line and sometimes forming closed cells; transverse veinlets delimiting elongate cells on apical $\frac{1}{3}$; vein A2 multiforked, with 2–4 terminals. Arrangement of secondary veins and veinlets variable between specimens and sometimes between the two wings of the same specimen (Fig. 52A).

LEGS. Pro- and mesofemora and -tibiae dorsoventrally flattened, elongate and slender; metatibiae with 3 lateral and 9 apical spines; first metatarsomere ventrally with pad of microsetae at interno-apical angle and group of 9 spines distributed more or less in two rows (Fig. 50B). Metatibiotarsal formula: (3) 9/4/0.

MALE GENITALIA. Pygofer rather short, higher than long and with basal margin sinuate in lateral view; in lateral view, dorsal portion narrower than ventral portion, and strong posterior projection at half height; aedeagus attached to apex of posterior process. Anal tube dorsoventrally flattened, elongate; epiproct at basal $\frac{1}{3}$. Gonostyli separated, large, mostly dorsoventrally flattened, and with an elongate basidorsal process directed dorsally and bearing spoon-shaped process apically. Aedeagus simple; phallobase with dorsal process elongate, projecting posteriorly and with apical part incurving, and ventral part elongate and dorsoventrally flattened; phallus membranous, dorsoventrally flattened and notched apically.

FEMALE TERMINALIA (Fig. 51). Abdominal segment VI strongly narrowing apically, with posterior margin straight in ventral view and with a small posterolateral process projecting laterally (Fig. 51C); anal tube elongate and narrow, strongly curved posteroventrally before anal opening, v-shaped in cross section beyond anus, lanceolate in dorsal view (Fig. 51A–B); gonopods unilobed, projecting dorsolaterad, longer than high, not surpassing anal tube (Fig. 51A–B); gonapophysis IX large, elongate, apically rounded and curved dorsad (Fig. 51A–B); gonocoxae VIII looking like reniform inflated pouch (Fig. 51A–B); gonapophysis VIII dorsoventrally flattened and fused together, and with sternite VII in a large semicircular lobe transversely undulate and apically rounded (Fig. 51A–C).

SEXUAL DIMORPHISM. Females slightly larger than males. Colour dimorphism also affecting the colour pattern of posterior wings: males with a white spot at basicostal angle, females with apical margin largely bordered with white.

SIZE. ♂: 8.8 mm; ♀: 10.0 mm.

Distribution and biology

Australia: northern Queensland (Fig. 3). Biology unknown.

Species included (1)

S. alboapicata (Jacobi, 1928) gen. et comb. nov. (N Queensland)

Stalobrachys alboapicata (Jacobi, 1928) gen. et comb. nov.

Figs 3, 50–54

Olonia alboapicata Jacobi, 1928: 5 (described).

Olonia alboapicata – Metcalf 1956: 65 (catalogued).

Diagnosis

As for genus.

Etymology

The species epithet is formed from *albus* (adjective, Latin) = ‘white’ and ‘apex’. It refers to the white apical patch of the posterior wing.

Material examined

Holotype

AUSTRALIA • ♀; N Queensland, Cedar Creek; [coordinates of Ravenshoe: 17°36'33" S, 145°29'01" E]; “Queensl. Mjöberg”, “Cedar creek”, “*Olonia alboapicata* Jac., A. Jacobi determ.”, “Typus”, “Typus”; NHRS (Fig. 52).

Notes

Jacobi (1928) erroneously stated that the type specimen is a male. He was apparently prone to confusing males and females of Eurybrachidae, as he made the same mistake for the types of *Elthenus modestus* Jacobi, 1928 and *Olonia nigroapicata* Jacobi, 1928 (Constant 2005b, 2006b, respectively).

Cedar Creek was a Jirrbal aboriginal camp located at the southern edge of Ravenshoe (North Queensland). It covers 38 ha of grassland and woodland on a basalt plateau bordered by North Cedar Creek, South Cedar Creek and the Millstream River in the west. It represented an *Eucalyptus* “pocket” at the edge of the rainforest. Mjöberg and his staff sampled the location 29 Mar.–17 Apr. 1913 (Ferrier 2006, 2015).

Additional material

AUSTRALIA • 1 ♂; N Queensland, Ravenshoe; 17°36'33" S, 145°29'01" E; Jan. 1921; USNM.

Description

MEASUREMENTS AND RATIOS. LT: ♂ (n = 1): 8.8 mm; ♀ (n = 1): 10.0 mm; BV/LV = 3.9; BF/LF = 2.0; LP+LM/BT = 0.61; Ltg/BTg = 2.17; LW/BW = 1.6.

Male

HEAD (Fig. 53A–D). Vertex concave, with anterior and posterior margins parallel, curved, slightly wrinkled longitudinally; yellowish brown with an impression marked with darker colour at each side of disc. Frons dark brown variegated with blackish. Clypeus elongate, coloured as frons. Genae brown. Labium brown, reaching metacoxae. Antennae dark brown; scape short, ring-shaped; pedicel subcylindrical, slightly narrowing towards apex.

THORAX (Fig. 53A–C). Pronotum yellowish brown, dark brown at lateral angles; slightly wrinkled; median carina obsolete and 2 small, impressed points on disc. Lateral fields of prothorax dark brown. Mesonotum dark brown, transversely wrinkled; small yellowish spot at apex of scutellum; median and peridiscal carinae weakly marked. Red ventrally. Tegulae brown.

TEGMINA (Fig. 53A, C). Brown, darker on clavus and apicocostal half of membrane, irregularly marked with small yellowish spots; one larger pale yellowish marking on vein A1 at midlength of clavus; triangular white marking on costal margin after nodal line; white marking at apicosutural angle. Costal and sutural margins subparallel until nodal line, then slightly converging; costal margin slightly sinuate; apical margin obliquely rounded.

POSTERIOR WINGS (Fig. 53E). Dark brown, paler on anal area and becoming darker, nearly black on about distal third; elongate, transverse, subtriangular white marking at apicocostal angle, extending on 4 cells. Margin of anal area slightly sinuate; sutural margin with 2 clefts, cubital one not marked.

LEGS (Fig. 53A–D). Pro- and mesocoxae reddish brown. Pro- and mesofemora dark brown with reddish and yellowish spots marking obsolete rings. Pro- and mesotibiae dark brown with yellowish markings on 2 obsolete rings. Pro- and mesotarsi dark brown, with basal half of third tarsomere reddish. Metacoxae

reddish; metafemora reddish with apex brown. Metatibiae brown, with 3 lateral spines yellowish basally and black distally, and 9 apical black-brown spines. Metatarsi reddish brown, with first tarsomere darker distally and with a ventral group of 9 black spines in two irregular rows.

ABDOMEN. Bright red with genital segments yellowish white.

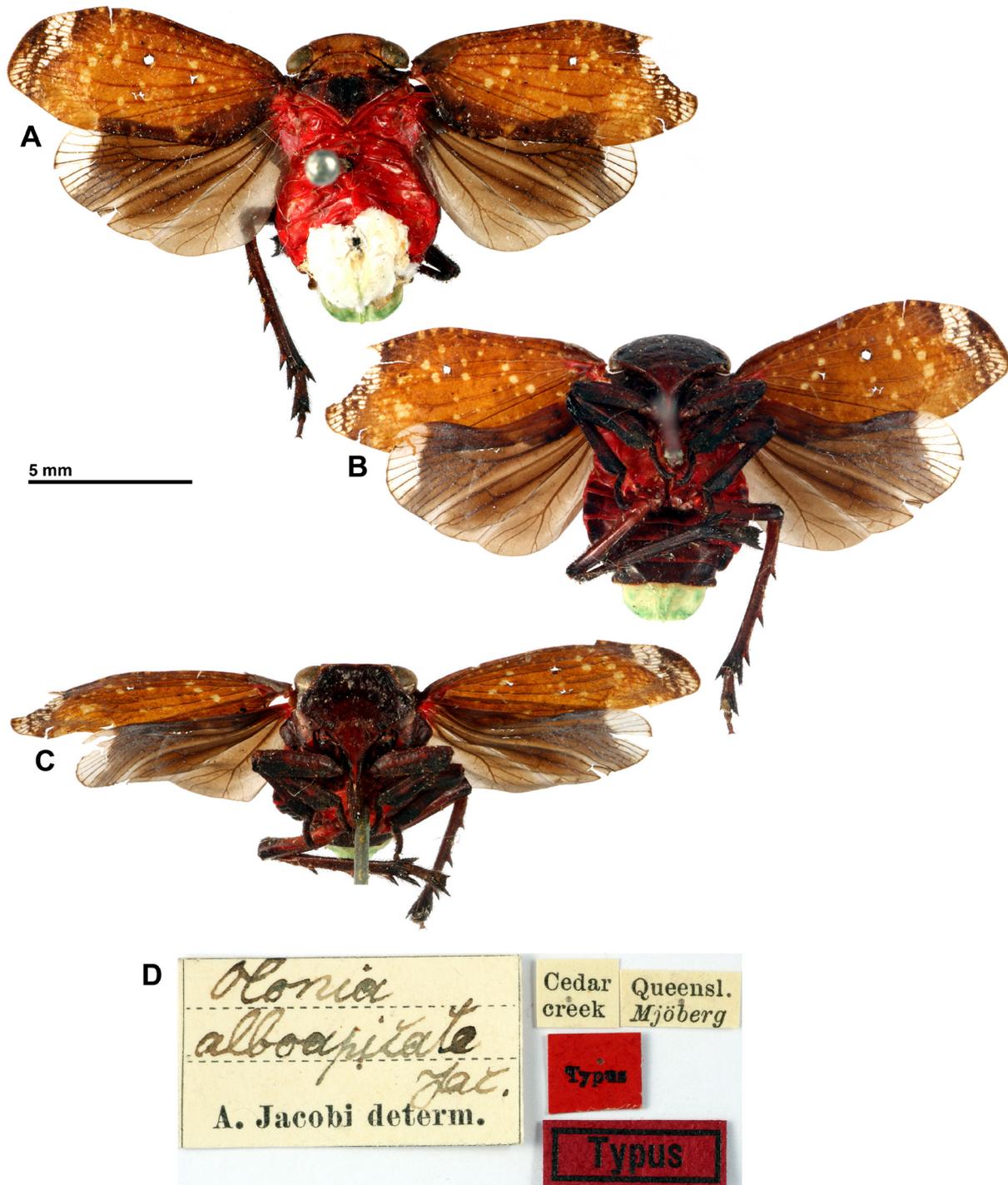


Fig. 52. *Stalobrachys alboapicata* gen. et comb. nov. (Jacobi, 1928), holotype, ♀. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, normal view of frons. D. Labels.

MALE GENITALIA (Fig. 54). Pygofer, anal tube and gonostyli whitish. Pygofer higher than long, with basal margin sinuate in lateral view; in lateral view, dorsal $\frac{2}{5}$ very narrow and ventral $\frac{2}{5}$ more than two times as broad as dorsal portion; strong posterior projection at half height, about twice as long as ventral portion (Fig. 54A, C). Anal tube dorsoventrally flattened, elongate, rounded apically in dorsal view and with posteromedian portion slightly depressed; epiproct at basal $\frac{1}{3}$ (Fig. 54A–B). Gonostyli separated, large, mostly dorsoventrally flattened, with elongate basidorsal process directed dorsally and bearing spoon-shaped process apically. Aedeagus simple; phallobase with dorsal process elongate, projecting

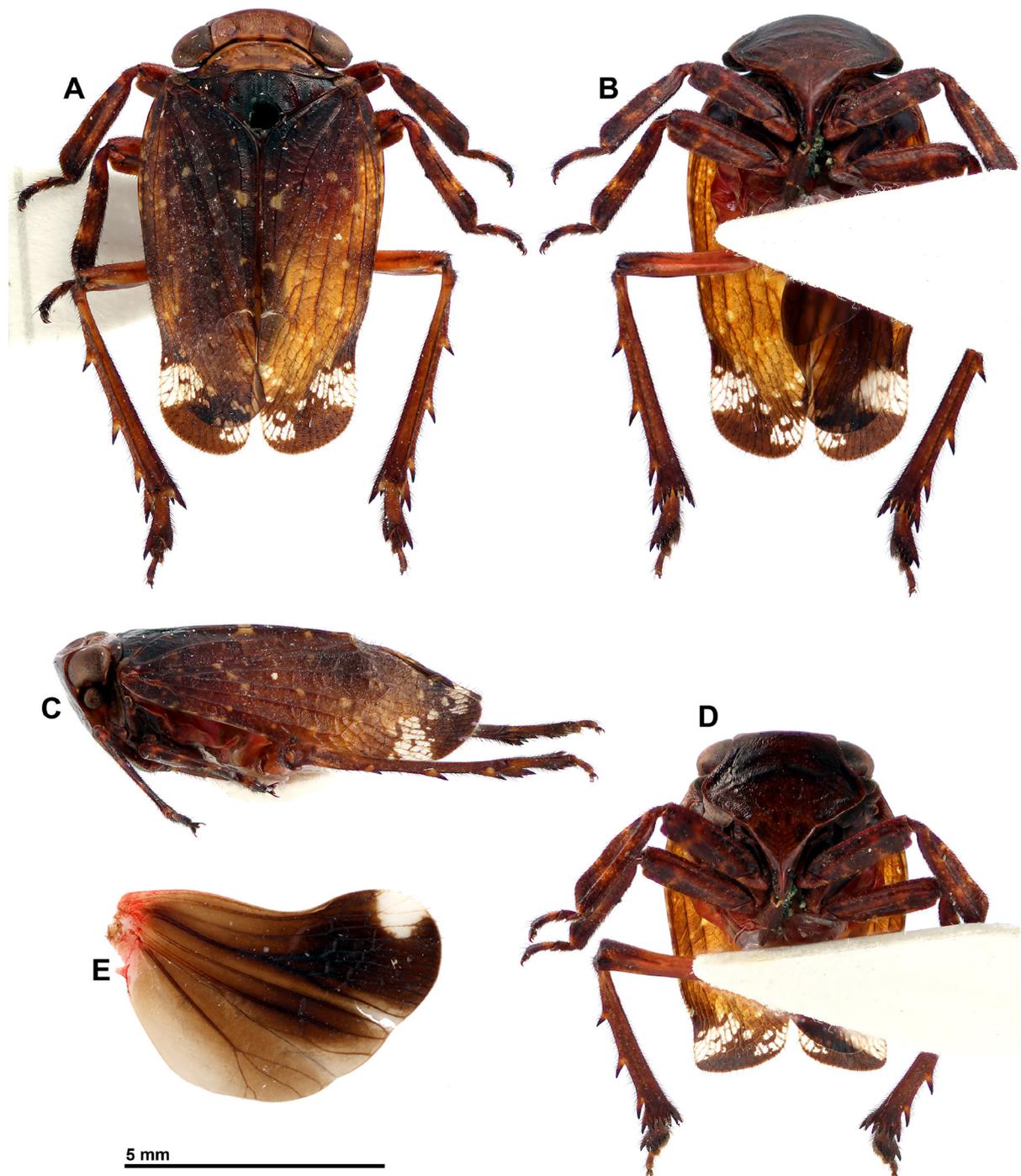


Fig. 53. *Stalobrachys alboapicata* gen. et comb. nov. (Jacobi, 1928), ♂. A. Habitus, dorsal view. B. Habitus, ventral view. C. Habitus, lateral view. D. Habitus, normal view of frons. E. Posterior wing.

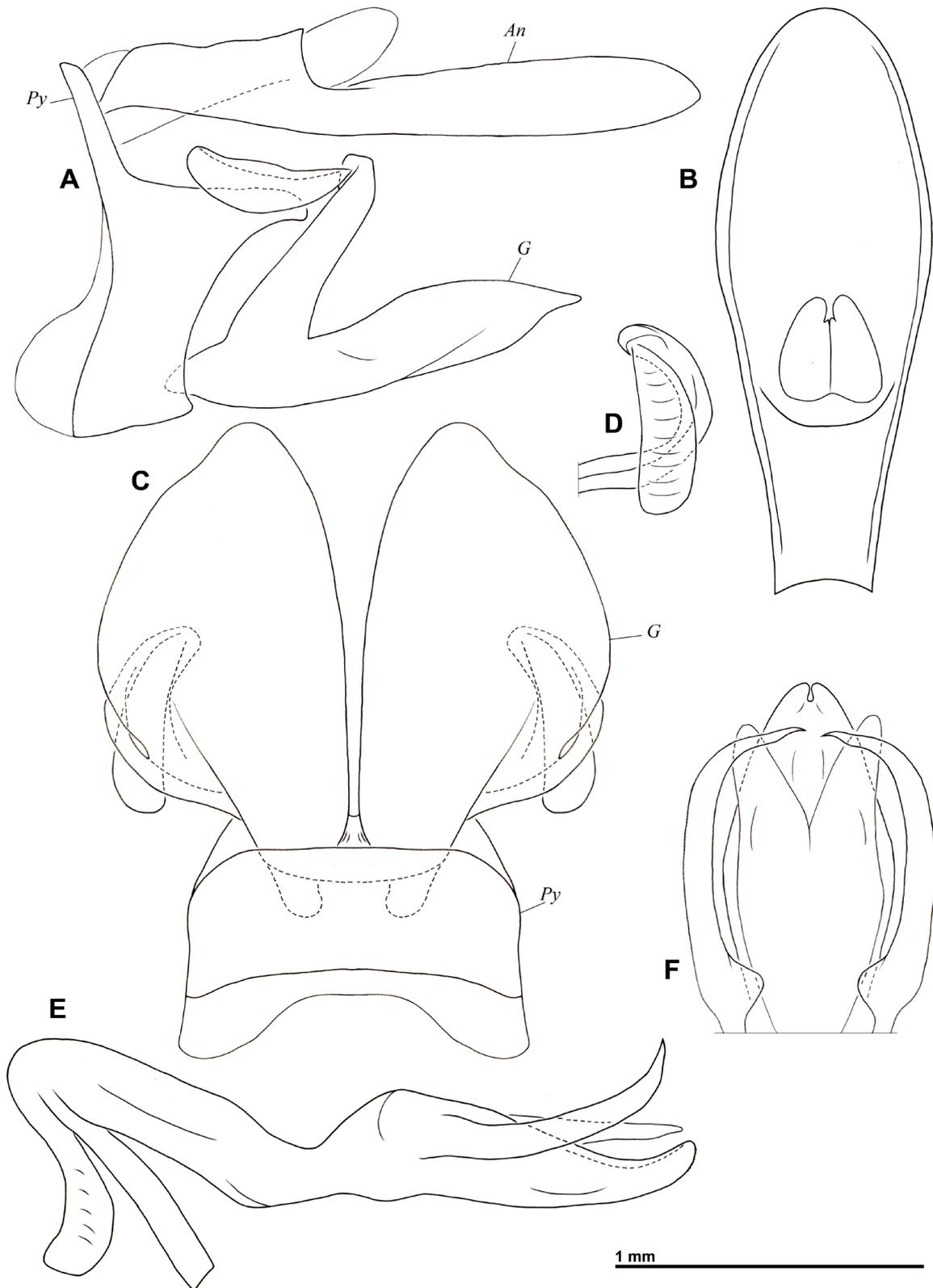


Fig. 54. *Stalobrachys alboapicata* gen. et comb. nov. (Jacobi, 1928), ♂, genitalia. **A.** Pygofer, anal tube and gonostyli, left lateral view. **B.** Anal tube, dorsal view. **C.** Pygofer and gonostyli, ventral view. **D.** Laterodorsal part of left gonostylus, dorsal view. **E.** Aedeagus, left lateral view. **F.** Aedeagus, dorsal view. Abbreviations: *An* = anal tube; *G* = gonostyli; *Py* = pygofer.

posteriorly and with apical part incurving, and ventral part elongate and dorsoventrally flattened and notched apically; phallus membranous, dorsoventrally flattened and bifid on distal $\frac{1}{3}$.

Female

Similar to male, but with distal cells of posterior wings white instead of having a white spot at the apicosutural angle (Fig. 52).

Distribution and biology

This species is recorded only from a small area around Ravenshoe, at the limit between the Queensland Tropical Rainforests and the Einasleigh Upland Savanna bioregions (Fig. 3). The specimens were found in January and April, one of them in a habitat of mixed grassland and woodland on a basalt plateau described as a *Eucalyptus* patch at the edge of the rainforest (Ferrier 2006, 2015). However, host plants remain undocumented.

Genus *Maeniana* Metcalf, 1952

Maenia Jacobi, 1928: 6 (nec *Maenia* Dalton, 1877) (type species: *Maenia hirsuta* Jacobi, 1928 by original designation).

Maeniana Metcalf, 1952: 228 (nom. nov. pro *Maenia* Jacobi, 1928 nec *Maenia* Dalton, 1877).

Maeniana – Metcalf 1956: 67 (catalogued).

Remarks

The most relevant diagnostic characters to recognize the genus *Maeniana* are these of the male genitalia (based on the male holotype of *M. hirsuta* (Jacobi, 1928)):

- (1) posterior margin of the pygofer with elongate laterodorsal process projecting posteriorly
- (2) gonostyli not or very shortly fused basally
- (3) laterodorsal part of the gonostyli elongate and laminate, without spine or hook, and bearing spoon-shaped process apically

Maeniana apicalis (Walker, 1851) comb. nov.

Fig. 55

Eurybrachys apicalis Walker, 1851: 393 (described).

Olonia apicalis – Stål 1862: 488 (transferred to *Olonia* in key to genera of Eurybrachidae). — Atkinson 1886: 13 (English translation of Stål's key). — Kirkaldy 1906: 445 (listed).

non *Olonia apicalis* – Lallemand 1935 (listed from Fergusson River, Katherine, Northern Territory, Australia (erroneous identification)).

Material examined

Holotype

AUSTRALIA • ♀; W Australia, NW coast; “N. Holl., N.W. Coast /on the reverse/ 44, 14”, “Type”, “*Eurybrachys apicalis*,” “Re-pinned on stainless”; BMNH.

Remarks

This species is known from a single female, but closely related species were examined in the course of this study and their male genitalia show that they belong to the genus *Maeniana*, a view supported by a comparison with the male genitalia of the type species, *Maeniana hirsuta* (Jacobi, 1928) (Constant, unpublished data).

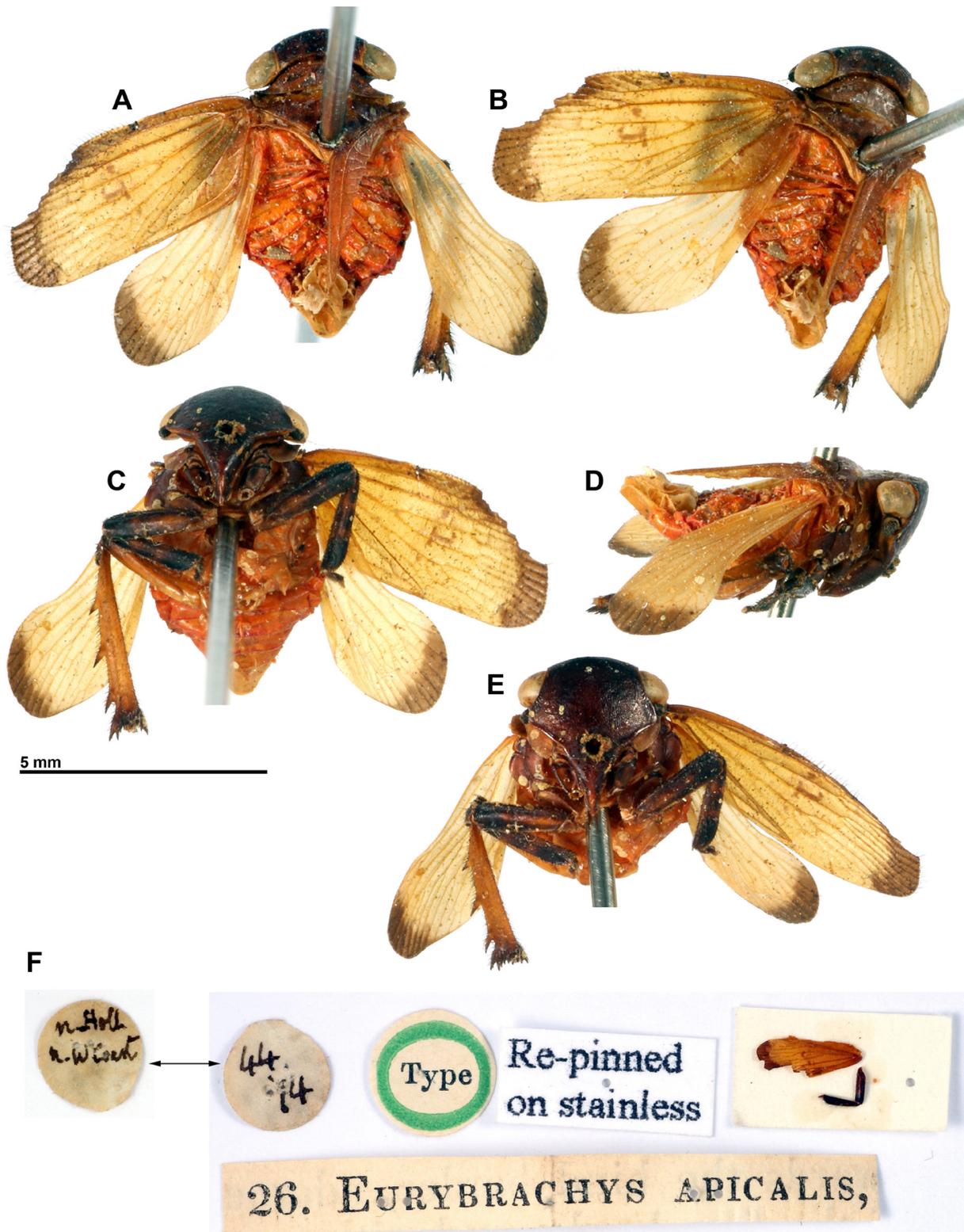


Fig. 55. *Maeniana apicalis* (Walker, 1851), holotype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, laterodorsal view. **C.** Habitus, ventral view. **D.** Habitus, lateral view. **E.** Habitus, normal view of frons. **F.** Labels (with leg and right tegmen).

Genus *Platybrachys* Stål, 1859

Platybrachys Stål, 1859: 280 (type species: *Platybrachys lanata* (Stål, 1854) by monotypy).

Platybrachys – Stål 1861: 210 (keyed); 1862: 488 (keyed, list of included species); 1863: 248 (compared with *Lyncilia* Stål, 1863), 249 (key to species). — Atkinson 1886: 13 (keyed). — Karsch 1890: 60 (compared with *Mesonitys* Karsch, 1890); 1895: 215 (compared with *Aspidonitys* Karsch, 1895). — Melichar 1903: 67 (comparative note). — Distant 1906: 206 (type species = *Platybrachys decemmacula* (Walker, 1851) (error)). — Kirkaldy 1906: 443 (keyed, notes); 1907: 6 (ecology: genus of *Eucalyptus* forests), 105 (problem of generic definition). — Schmidt 1908: 243 (listed). — Hacker 1924: 37 (natural history), 40 (compared with *Olonia* Stål, 1862). — Evans 1938: 4 (morphology). — Metcalf 1947: 163 (zoogeography); 1956: 67 (catalogued). — Fennah 1964: 159 (keyed), 160 (senior synonym of *Lyncilia* Stål, 1863 (erroneous)). — Constant 2006a: 47 (notes); 2006b: 31 (compared with *Olonia*); 2008a: 41 (compared with *Chewobrachys* Constant, 2008).

Platybrachys ornata (Lallemand, 1928) comb. nov.

Fig. 56

Olonia ornata Lallemand, 1928: 393 (described).

Olonia ornata – Metcalf 1956: 65 (catalogued).

Material examined

Holotype

AUSTRALIA • ♀; Northern Territory, Hermansburg; “Hermansburg, N.T., S. Australia, H.J. Hillier. 1907–233.”, “Type”, “*Olonia ornata* Lallemand, Type, dét. V. Lallemand 19”, “10”; BMNH.

Remarks

The species is known from a single female and the key to the genera proposed by Fennah (1964) leads to the genus *Platybrachys* Stål, 1859. It is quite similar to *P. leucostigma* (Walker, 1851) (illustrations of the latter species available in Hacker 1924: fig. 14) by showing an apical white border on the posterior wings and numerous elongate cells in the costal area. The species is here transferred to *Platybrachys*, although it is obvious that a complete revision of the genus will be necessary (Constant 2008a).

Discussion

Taxonomic issues

I recently described two genera (Constant 2006b) to include species previously placed in *Olonia*:

- (1) *Hackerobrachys* Constant, 2006 to accommodate *O. viridiventris* Stål, 1863, and
- (2) *Fletcherobrachys* Constant, 2006 to accommodate *Platybrachys stillata* Bergroth, 1907, under which I synonymized *O. nigroapicata* Jacobi, 1928

The study of the type specimens and available material from many museums raised the issue of the inconsistency of the genus *Olonia*, as already partly treated in Constant (2006b). The study of the male genitalia revealed that the species *Olonia alboapicata* Jacobi, 1928 does not belong in *Olonia* and cannot be placed in one of the existing genera. The new genus *Stalobrachys* gen. nov. is hence necessary to accommodate this species.

The species *Olonia ornata* Lallemand, 1928, described from a female specimen from Hermansburg (erroneously spelled “Hermansburg” by Lallemand 1928) in the Northern Territory near Alice Springs,

does not belong in *Olonia* and is transferred to *Platybrachys*. *Olonia apicalis*, described from the northwestern coast of Australia by Walker (1851), does not belong in *Olonia* either and is transferred to *Maeniana* Metcalf, 1952.

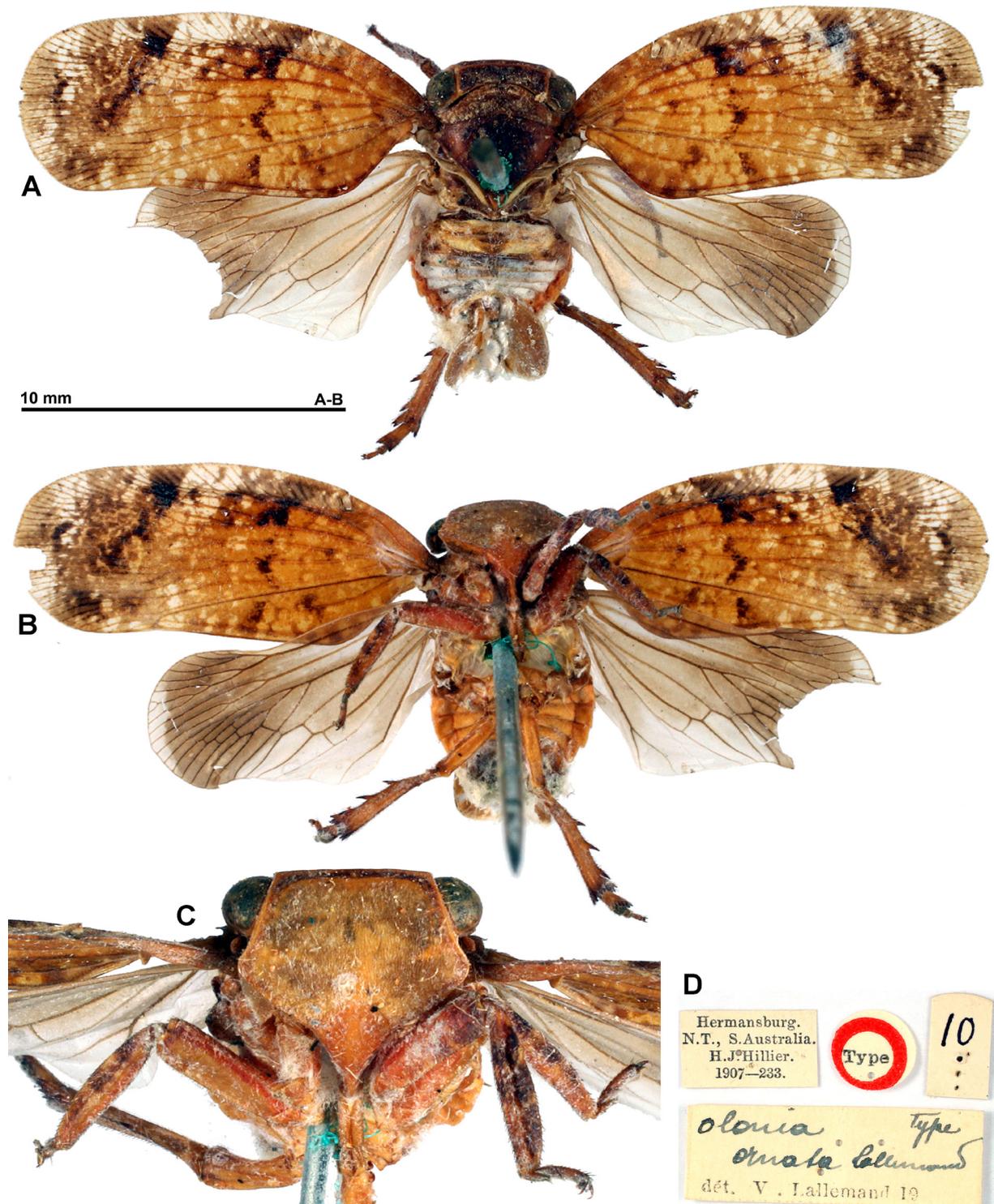


Fig. 56. *Platybrachys ornata* (Lallemand, 1928) comb. nov., holotype, ♀. **A.** Habitus, dorsal view. **B.** Habitus, ventral view. **C.** Habitus, normal view of frons. **D.** Labels.

The review of the specimens reported by Jacobi (1928) as belonging to *Olonia* revealed the following errors:

- (1) the specimens identified as *Olonia rubicunda* (Walker, 1851) from Kimberley District (Western Australia instead of Northern Territory, as erroneously stated by Jacobi) represent a new species, the generic attribution of which will be treated in another paper (Constant, unpublished data)
- (2) the specimens mentioned under *Olonia transversa* (Walker, 1858) were misidentified and the synonymy of *Olonia picea* Kirkaldy, 1906 is erroneous (Constant 2005b)
- (3) *Olonia nigroapicata* Jacobi, 1928 is actually a junior synonym of *Fletcherobrachys stillata* (Bergroth, 1907) (Constant 2006b)

Male genitalia

The genus *Olonia* is well characterized by the shape of the male genitalia, with a reduced aedeagus and very specialized, strongly sclerotized gonostyli. The male genitalia also offer very relevant characters for species separation. Although externally very similar, the genus *Stalobrachys* gen. nov. can be separated from *Olonia* at first glance if the male genitalia are considered. However, both genera show a peculiar spoon-shaped process on the gonostyli, a character shared also with the genera *Chewobrachys* Constant, 2008, *Fletcherobrachys* Constant, 2006, *Hackerobrachys* Constant, 2006, *Maeniana* Metcalf, 1952 and *Nirus* Jacobi, 1928, and probably also *Loisobrachys* Constant, 2008 although the latter is known only from a single female. All these genera are from Australia and it seems that the “spoon-shaped process character” allows the separation of a consistent group among the Eurybrachidae from Australia, the status of which will need further study. More than 70% of the Australian eurybrachids remain undescribed, both at specific and generic levels (Constant, unpublished data), and molecular data would provide a great help for the assessment of suprageneric relationships. The role of the spoon-shaped process remains unknown and requires the observation of living specimens.

Biological data

The present work also provides a series of new host plant data, which clearly shows that species of *Olonia* are polyphagous, as opposed to the other Australian genera for which host plants have been recorded, e.g., *Chewobrachys* Constant, 2008, *Gelastopsis* Kirkaldy, 1906 and *Hackerobrachys* Constant, 2006 being recorded only on *Acacia* spp. (Fabaceae), and *Platybrachys* Stål, 1859 recorded only on *Eucalyptus* spp. (Myrtaceae) (Hacker 1924; Constant 2006b, 2008a; Bourgoin 2018). The list of host plants of *Olonia* spp. contains species of the family Myrtaceae: *Melaleuca quinquenervia* (Cav.) S.T.Blake, *Eucalyptus* sp. and *Corymbia tessellaris* K.D.Hill & L.A.S.Johnson; Bursaceae: *Canarium australianum* F.Muell.; Vitaceae: *Vitis vinifera* L.; Urticaceae: *Pipturus argenteus* Wedd.; Solanaceae: *Solanum tubiferum* L.; Convolvulaceae: *Ipomoea pes-caprae* (L.) R.Br. More host plants will likely be recorded if more field research is conducted. On the other hand, the host plants of *Stalobrachys* gen. nov. remain undocumented. From the available data, the species of *Olonia* seem to be present as adults all year round, but this requires confirmation from additional field work. The habitat requirements of the different species need to be defined from field observations rather than by extrapolation from the general habitat represented in each bioregion, e.g., *O. marginata* is present in the Queensland tropical rainforests bioregion, but was recorded from open coastal areas and vineyards rather than from the rainforest. It is also interesting to note that the genus seems to be restricted to the eastern part of Queensland, extending to the Great Dividing Range to the west, and mountains might have played a role in the speciation within the genus; however, this hypothesis requires a much more complete knowledge of the actual distribution of the species before any conclusion can be proposed.

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References

- Atkinson E.T. 1886. Notes on Indian Rhynchota. No. 5. *Journal of the Asiatic Society of Bengal* 55: 12–83. Available from <https://www.biodiversitylibrary.org/page/35545361> [accessed 23 Sep. 2018].
- Banks J. 1771. *The Endeavour Journal of Sir Joseph Banks*. Available from <http://gutenberg.net.au/ebooks05/0501141h.html#jul1770> [accessed 6 Sep. 2017].
- Bourgoin T. 1993. Female genitalia in Hemiptera Fulgoromorpha, morphological and phylogenetic data. *Annales de la Société entomologique de France* 29: 225–244.
- Bourgoin T. 2018. FLOW (Fulgoromorpha Lists On the Web): a World Knowledge Base Dedicated to Fulgoromorpha. Ver. 8, updated. Available from <http://hemiptera-databases.org/flow/> [accessed 18 Jun. 2018].
- Bourgoin T., Wang R.R., Asche M., Hoch H., Soulier-Perkins A., Stroinski A., Yap S. & Szwedo J. 2015. From micropterism to hyperpterism: recognition strategy and standardized homology-driven terminology of the fore wing venation patterns in planthoppers (Hemiptera: Fulgoromorpha). *Zoomorphology* 134 (1): 63–77. <https://doi.org/10.1007/s00435-014-0243-6>
- Carayon J. 1969. Emploi du noir chlorazol en anatomie microscopique des insectes. *Annales de la Société entomologique de France* 5: 179–193.
- Constant J. 2004. Révision des Eurybrachidae (I). Le genre *Amychodes* Karsch, 1895 (Homoptera: Fulgoromorpha: Eurybrachidae). *Bulletin de l'Institut royal des Sciences naturelles de Belgique* 74: 11–28.
- Constant J. 2005a. Revision of the Eurybrachidae (III). The Afrotropical genus *Metoponitys* Karsch, 1890 (Hemiptera: Fulgoromorpha: Eurybrachidae). *Bulletin de l'Institut royal des Sciences naturelles de Belgique* 75: 41–56.
- Constant J. 2005b. Revision of the Eurybrachidae (IV). The Australian genus *Gelastopsis* Kirkaldy, 1906 (Hemiptera: Fulgoromorpha: Eurybrachidae). *Bulletin de l'Institut royal des Sciences naturelles de Belgique* 75: 57–69.
- Constant J. 2006a. Revision of the Eurybrachidae (V). Description of the new Australian genus *Kirkaldybrachys* Constant (Hemiptera: Fulgoromorpha: Eurybrachidae). *Bulletin de la Société royale belge d'Entomologie* 142: 47–54.

- Constant J. 2006b. Revision of the Eurybrachidae (VII). The Australian genera *Hackerobrachys* and *Fletcherobrachys* (Hemiptera: Fulgoromorpha: Eurybrachidae). *Bulletin de l'Institut royal des Sciences naturelles de Belgique* 76: 31–40.
- Constant J. 2008a. Revision of the Eurybrachidae (XIII). The new Australian genus *Chewobrachys* (Hemiptera: Fulgoromorpha). *Zootaxa* 1898: 41–54.
- Constant J. 2008b. Revision of the Eurybrachidae (XIV). The new Australian genus *Loisobrachys* (Hemiptera: Fulgoromorpha). *Annales Zoologici* 58 (4): 755–759.
- Distant W.L. 1906. Rhynchotal notes xxxix. *Annals and Magazine of Natural History, Series 7* 18: 191–208. <https://doi.org/10.1080/00222930608562600>
- Distant W.L. 1908. On some Australian Homoptera. Synonymical notes. *Annales de la Société entomologique de Belgique* 52: 97–111. Available from <http://biodiversitylibrary.org/page/13193447> [accessed 23 Sep. 2018].
- Donovan E. 1805. *An Epitome of the Natural History of the Insects of New Holland, New Zealand, New Guinea, Otaheite, and Other Islands in the Indian, Southern, and Pacific Oceans*. F.C. & J. Rivington, London.
- Donovan E. 1820. Cicada. In: Rees A. (ed.) *The Cyclopaedia; or, Universal Dictionary of Arts, Sciences and Literature* 8: 115–121. Available from <https://biodiversitylibrary.org/page/38741207> [accessed 23 Sep. 2018].
- Evans J.W. 1933. A revision of the Eurymelini (Homoptera, Bythoscopidae). *Transactions and Proceedings of the Royal Society of South Australia* 57: 73–90. Available from <https://biodiversitylibrary.org/page/41569875> [accessed 23 Sep. 2018].
- Evans J.W. 1938. Australian leaf-hoppers (Homoptera, Jassoidea), Part VIII. *Papers and Proceedings of the Royal Society of Tasmania* 1938: 1–18.
- Fabricius J.C. 1775. Ryngota. In: Fabricius J.C. 1775. *Systema Entomologiae, Sistens Insectorum Classes, Ordines, Genera, Species, Adiectis Synonymis, Locis, Descriptionibus, Observationibus*: 673–689. Kortius, Flensburg and Leipzig. Available from <https://biodiversitylibrary.org/page/25552060> [accessed 1 Oct. 2018].
- Fabricius J.C. 1781. *Species Insectorum Exhibentes eorum Differentias Specificas, Synonyma Auctorum, Loca Natalia, Metamorphosin Adiectis Observationibus, Descriptionibus*. Bohnius, Hamburg and Kiel. <https://doi.org/10.5962/bhl.title.36509>
- Fabricius J.C. 1787. *Mantissa Insectorum Sistens Species Nuper Detectas Adiectis Synonymis, Observationibus, Descriptionibus, Emendationibus* 2. Proft, Havniae [Copenhagen]. <https://doi.org/10.5962/bhl.title.36471>
- Fabricius J.C. 1794. *Entomologia Systematica Emendata et Aucta: Secundum Classes, Ordines, Genera, Species, Adiectis Synonymis, Locis, Observationibus, Descriptionibus* 4: 1–472. Proft, Havniae [Copenhagen]. <http://dx.doi.org/10.5962/bhl.title.36532>
- Fabricius J.C. 1803. *Systema Rhyngotorum Secundum Ordines, Genera, Species, Adiectis Synonymis, Locis, Observationibus, Descriptionibus*. Carolus Reichard, Braunschweig. <https://doi.org/10.5962/bhl.title.11644>
- Fennah R.G. 1964. Three new genera of Eurybrachidae (Homoptera: Fulgoroidea) from West Africa and Australia. *Proceedings of the Entomological Society of London B* 33 (9–10): 157–162.
- Ferrier Å. 2006. Dr Eric Mjöberg's 1913 scientific exploration of North Queensland's rainforest region. *Memoirs of the Queensland Museum, Cultural Heritage Series* 4 (1): 1–27.

- Ferrier Å. 2015. Journeys into the rainforest: archaeology of culture change and continuity on the Evelyn Tableland, North Queensland. *Terra Australis* 43: 1–174.
- Germar E.F. 1830. Species cicadarium enumeratae et sub genera distributae. *Thon's Entomologisches Archiv* 2 (2): 1–57.
- Gmelin J.F. 1789. *Caroli a Linné. Systema Naturae per Regna tria Naturae: Secundum Classes, Ordines, Genera, Species, cum Characteribus, Differentiis, Synonymis, Locis* 1 (4): 1517–2224. Emanuel Beer, Leipzig. <https://biodiversitylibrary.org/page/25745483>
- Goeze J.A.E. 1778. *Entomologische Beyträge zu des Ritter Linné zwölften Ausgabe des Natursystems*. Vol. 2. Weidmanns Erben und Reich, Leipzig. <https://doi.org/10.5962/bhl.title.45974>
- Hacker H. 1924. Field notes on *Platybrachys*, & c. (Homoptera). *Memoirs of the Queensland Museum* 8: 37–42.
- Ingram G. 1998. *Trip Report: Bakers Blue Mountain (NE Queensland, Australia), July 17, 1997*. Available from <http://www.camacdonald.com/birding/tripreports/BakersMt97.html> [accessed 26 Dec. 2015].
- Jacobi A. 1928. Results of Dr E. Mjöberg's Swedish Scientific Expeditions to Australia 1910–1913. Rhynchota Homoptera. 1. Fulgoridae und Cercopidae. *Arkiv för Zoologi* 19A (28): 1–50.
- Karsch F.A.F. 1890. Afrikanische Fulgoriden. *Berliner Entomologische Zeitschrift* 35: 57–70. Available from <https://www.biodiversitylibrary.org/page/25049326> [accessed 23 Sep. 2018].
- Karsch F.A.F. 1895. Aethiopische Eurybrachiden. *Entomologische Nachrichten* 21: 209–217. Available from <https://www.biodiversitylibrary.org/page/25411449> [accessed 23 Sep. 2018].
- Kershaw J.C. & Muir F. 1922. The genitalia of the auchenorhynchous Homoptera. *Annals of the Entomological Society of America* 15 (3): 201–211.
- Kirkaldy G.W. 1906. Leaf hoppers and their natural enemies. *Bulletin of the Experiment Station of the Hawaiian Sugar Planters' Association, Division of Entomology* 1 (9): 271–479. Available from <https://biodiversitylibrary.org/page/15499805> [accessed 24 Sep. 2018].
- Kirkaldy G.W. 1907. Leaf hoppers supplement (Hemiptera). *Bulletin of the Experiment Station of the Hawaiian Sugar Planters' Association, Division of Entomology* 3: 1–186. Available from <https://biodiversitylibrary.org/page/15500139> [accessed 24 Sep. 2018].
- Lallemand V. 1928. Fulgorides nouveaux provenant de la collection du British Museum. *Annals and Magazine of Natural History, Series 10* 1 (2): 241–249. <https://doi.org/10.1080/00222932808672768>
- Lallemand V. 1935. Homoptères des Iles de la Sonde et de l'Australie du Nord. *Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève* 42: 661–681.
- Melichar L. 1903. Homopteren-Fauna von Ceylon. F.L. Dames, Berlin.
- Metcalf Z.P. 1936. Part 2. Cixidae. In: Metcalf Z.P. (ed.) *General Catalogue of the Homoptera, Fascicule IV*: 1–267. North Carolina State College, Raleigh, NC, USA.
- Metcalf Z.P. 1938. The Fulgorina of Barro Colorado and other parts of Panama. *Bulletin of the Museum of Comparative Zoology* 82: 277–423.
- Metcalf Z.P. 1947. The center of the origin. *Journal of the Elisha Mitchell Science Society* 62: 149–175.
- Metcalf Z.P. 1952. New names in the Homoptera. *Journal of the Washington Academy of Sciences* 42 (7): 226–231.

Metcalf Z.P. 1956. Part 18. Eurybrachidae and Gengidae. In: Metcalf Z.P. (ed.) *General Catalogue of the Homoptera. Fascicle IV, Fulgoroidea*. North Carolina State College, Raleigh, NC, USA.

Muir F. 1923. On the classification of the Fulgoroidea (Homoptera). *Proceedings of the Hawaiian Entomological Society* 5: 205–247.

Muir F. 1925. On the genera of Cixiidae, Meenoplidae and Kinnaridae. *Pan-Pacific Entomologist* 1: 97–110, 156–163.

Schaum H.R. 1850. Fulgorellae. Erste Section A–G. In: Ersch I.S. & Gruber I.G. (eds) *Allgemeine Encyclopädie der Wissenschaften und Künste in alphabetischer Folge von genannten Schriftstellern bearbeitet und herausgegeben* 51: 58–73. Johann Friedrich Gleditsch, Leipzig.

Schmidt E. 1908. Beitrag zur Kenntnis der Eurybrachinen (Hemiptera – Homoptera). *Zoologischer Anzeiger* 33: 241–247.

Shorthouse D.P. 2010. SimpleMappr, an Online Tool to Produce Publication-Quality Point Maps. Available from <http://www.simplemappr.net> [accessed 13 Dec. 2017].

Signoret V. 1850. Notice sur le groupe des euryméliides. *Annales de la Société entomologique de France, Série 2* 8: 497–513.

Signoret V. 1858. Rectifications synonymiques relatives à divers Hémiptères. *Bulletin de la Société entomologique de France, Série 3* 6: xxxii.

Available from <http://biodiversitylibrary.org/page/8375865> [accessed 23 Sep. 2018].

Soulier-Perkins A. 1997. *Systématique phylogénétique et test d'hypothèses biogéographiques chez les Lophopidae (Homoptera, Fulgoromorpha)*. PhD Thesis. Muséum national d'Histoire naturelle, Paris.

Soulier-Perkins A. & Bourgoïn T. 1998. Copulatory mechanisms and sexual selection in the Lophopidae (Hemiptera: Fulgoromorpha). *Annales de la Société entomologique de France, N.S.* 34 (2): 149–162.

Stål C. 1859. Hemiptera. Species novas descripsit. Fregatten Eugenies Resa. *Arkiv för Zoologi* 4: 219–298.

Stål C. 1861. Nova methodus familias quasdam hemipterorum disponendi genera. *Öfversigt af Kongliga Svenska Vetenskaps-Akademiens Förhandlingar* 18: 195–212.

Available from <https://www.biodiversitylibrary.org/page/15376759> [accessed 23 Sep. 2018].

Stål C. 1862. Synonymiska och systematiska anteckningar öfver Hemiptera. *Öfversigt af Kongliga Svenska Vetenskaps-Akademiens Förhandlingar* 19: 479–504.

Available from <https://www.biodiversitylibrary.org/page/15376267> [accessed 23 Sep. 2018].

Stål C. 1863. Beitrag zur Kenntnis der Fulgoriden. *Entomologische Zeitung, herausgegeben von dem entomologischen Vereine zu Stettin* 24: 230–251.

Available from <https://www.biodiversitylibrary.org/page/35950043> [accessed 23 Sep. 2018].

Turton W. 1806. *A General System of Nature, through the Three Grand Kingdoms of Animals, Vegetables, and Minerals, Systematically Divided into their Several Classes, Orders, Genera, Species, and Varieties, with their Habitations, Manners, Economy, Structure and Peculiarities*. Lackington, Allen and Co., Temple of the Muses, London. <https://doi.org/10.5962/bhl.title.37018>

Walker F. 1851. *List of the Specimens of Homopterous Insects in the Collection of the British Museum* 2: 261–636. Trustees of the British Museum, London.

Available from <https://www.biodiversitylibrary.org/page/9620260> [accessed 23 Sep. 2018].

Walker F. 1852. *List of the Specimens of Homopterous Insects in the Collection of the British Museum, Supplement* 4: 1119–1188. Trustees of the British Museum, London.

Available from <https://www.biodiversitylibrary.org/page/9692970> [accessed 23 Sep. 2018].

Walker F. 1858. *Supplement. List of the Homopterous Insects in the Collections of the British Museum* 1–307. Trustees of the British Museum, London.

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